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Environmental Friendly Vector Control Potentials of Plant Secondary Metabolites

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Environmental Friendly Vector Control Potentials of Plant Secondary Metabolites

A THESIS SUBMITTED TO THE INSTITUTE OF ENVIRONMENTAL SCIENCE UNIVERSITY OF RAJSHAHI, BANGLADESH IN PARTIAL FULFILLMENT OF THE REQUIREMENTS FOR THE DEGREE OF DOCTOR OF PHILOSOPHY

Submitted By

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Certificate

It is my pleasure to certify that the thesis entitled, "Environmental friendly vector control potentials of plant secondary metabolites" submitted to the Institute of Environmental Science, University of Rajshahi by Nayeema Parvin in partial fulfillment of the requirements for the degree of Doctor of Philosophy is a dissertation of the perfect study which she carried out in my laboratory with much success. It contains no materials previously published, or submitted for any other purpose, or written by any other person except, wherever, due references are made.

I hereby clarify that the author completed her work under my direct supervision and contributed some new ideas and openings by adding most recent information in this research arena. She also enriched results of screening of the selected plants for biological activity and reintroduced most common test organisms of their 'vector' identity with reasonable references as well.

September, 2015 R A J S H A H I

Motihaar Green (Dr. Md. Nurul Islam)

Declaration *I hereby declare that the whole work submitted as a entitled "Environmental friendly vector control potentials of plant secondary metabolites" to the Institute of Environmental Science, University of Rajshahi in partial fulfillment of the requirements for the degree of Doctor of Philosophy is the result of my own investigation. The thesis contains no materials previously published or accepted by any institution for the award of any other degree or diploma elsewhere except wherever due references are made.* (Nayeema Parvin)

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The Author

CONTENTS

A checklist of the Tables provided

A checklist of the Figures provided

A checklist of the plates provided

ABBREVIATIONS OF THE SPECIAL WORDS USED IN THE TEXT

ABBREVIATIONS OF THE SPECIAL WORDS USED IN THE TEXT

ABSTRACT

Whole plants (wp) of *Evolvulus nummularius*, *Mentha piperita*, *Mimosa pudica*, *Parthenium hysterophorus*, *Phyllanthus niruri*, *Polygonum hydropiper*, *Pouzolzia zeylanica*, *Synedrella nodiflora*, aerial part (ap) and roots (r) of *Lantana camara*, and aerial part and rhizome (rh) of *Zingiber zerumbet* were extracted in petroleum ether (PetE), chloroform $(CHCl₃)$ and methanol (CH₃OH), and were screened against four vectors *i.e.* eggplant aphid, *Aphis gossypii*; larvae of the mosquito, *Culex quinquefasciatus*; red flour beetle, *Tribolium castaneum* and brine shrimp, *Artemia salina* nauplii under laboratory conditions to yield their efficacy through dose-mortality assay and repellent activity against *A. gossypii* and *T. castaneum*; lethality against *A. salina*; larvicidal activity against *C. quinquefasciatus* larvae with much success. Antimicrobial activity tests were carried out in this connection for further justification of efficacy of the extractives. Isolation, purification and characterization of plant component(s) were also done to establish step-forward positive control measure(s) against four selected vectors from the pest control and pharmaceutical points of view.

The mortality of *A. gossypii* were found in the descending order started with the *E.* nummularius (wp/PetE) giving LD₅₀ 0.034 mg/cm² and ended with *L. camara* $(ap/PetE)$ giving LD_{50} 0.080mg/cm².

The larvicidal activity against *C. quinquefasciatus* larvae the descending order of intensity was started with *Ph. niruri* (wp/CHCl₃) provided with the LC₅₀ 3.220ppm and ended with *Mi. pudica* (wp/PetE) giving LC₅₀ 88.187ppm). Methanol extract of *Mi. pudica* (wp), *Pz. zeylanica* (wp) and *Z. zerumbet* (ap) did not show any larvicidal effect.

Brine shrimp lethality against *A. salina* nauplii the efficacy was in the descending order of *Po. hydropiper* (wp/CHCl₃) provided with the LC₅₀ 1.590ppm and ended with *Ph. niruri* (wp/CHCl₃) giving LC₅₀ 24.331ppm.

The mortality of *T. castaneum* adults through dose-mortality assay were found in the descending order of *M. piperita* (wp/CH₃OH) provided with the LD₅₀ 0.238mg/cm² and ended with L. camara (r/PetE) giving LD₅₀ 2.672mg/cm². While, no activity was traced for whole plant of *Mi. pudica*; and the methanol extract of *Pz. zeylanica* (wp);

methanol and CHCl₃ extracts of *L. camara* (r) and CHCl₃ extract of *S. nodiflora* (wp) didn't show any mortality against the adult beetles of *T. castaneum*.

Against the eggplant aphids CH3OH extracts of *E. nummularius* (wp), *L. camara* (ap), *Mi. pudica* (wp) and *Pz. zeylanica* (wp) offered repellent activity, while the PetE and CHCl₃ extracts of the same didn't show repellency; however PetE, CHCl₃ and CH₃OH the extracts of *Ph. niruri* (wp), *S. nodiflora* (wp) and *Z. zerumbet* (ap); PetE extracts of *M. piperita* (wp), *P. hysterophorus* (wp) and *Z. zerumbet* (rh); CHCl₃ extracts of *L. camara* (r) and *Po. hydropiper* (wp) and CH3OH extracts of *P. hysterophorus* (wp) and *Po. hydropiper* (wp) offered no repellent activity at all.

For the repellency against *T. castaneum* adults CH3OH extracts of *Po. hydropiper* (wp) offered the most promising activity, however except *L. camara* (ap/CHCl₃ and CH3OH); *L. camara* (r/PetE); *M. piperita* (wp/PetE); *Mi. pudica* (wp/PetE and CH3OH); *P. hysterophorus* (wp/CH3OH); *Ph. niruri* (wp/CHCl3 and CH3OH); *Pz. zeylanica* (wp/CHCl3) showed repellent activity of different degree; and aerial and rhizome part of *Z. zerumbet* extracts of all the three solvents showed no repellent activity.

Activity tests of the extracts against eight selected pathogenic bacteria at concentrations of 200µg disc⁻¹ and 400µg disc⁻¹ compared with the standard antibiotic, Ampicilin 10µg disc⁻¹ provided a promising outcome. The chloroform extract was found most effective in comparison to PetE and CH3OH extracts. Finding potential activity the PetE extract of *E. nummularius* (wp) and CH3OH extract of *Po. hydropiper* (wp) were attempted for chromatographic fractionation to isolate bioactive compound(s) and as a result two compounds named ENP and POM were isolated, and only the ENP was determined as palmitic acid.

All the selected arthropod test agents are vectors [Aphid (Ng and Perry, 2004), Mosquito larvae (Michigan Mosquito Control Organization, 2013; Wilcox and Ellis, 2006: Janet, 2010), Red flour beetle (Channaiah *et al*., 2009); Brine shrimp (Sudhakaran *et al*., 2006: Hameed *et al*., 2002)] and the data achieved from the bioassays clearly showed the presence of insecticidal properties and vector control potentials in the test plants*.* Thus, comprehensive phytochemical analyses of the test plants for their insecticidal, insect repellent, cytotoxic and larvicidal leads, as well as the pharmacological studies of the active ingredients are very much to be solicited for their possible use in the future vector control and pharmaceutical endeavors.

INTRODUCTION

Pesticides represent the only group of chemicals that are purposely applied to the environment with an aim to suppress pests of plants and animals and to protect agricultural and industrial products. However, the majority of pesticides are not specifically targeting the pests, they also affect non-target organisms. Thus, repeated application of pesticides leads to loss of biodiversity. Many pesticides do not degrade easily, and they persist in soil, leach to ground and surface water and contaminate the wide environment. Depending on their chemical properties they can enter the organism, bio-accumulate in food chains and consequently influence human health. Therefore, intensive pesticide application results in several negative effects in the environment that cannot be ignored (Pesticide action network, 2010).

The use of pesticides by Bangladeshi farmers increased by 328 percent during the past 10 years, posing a serious health hazards on human being due to its long-term residual effect, according to a study released by Bangladesh Rice Research Institute (Islam, 2010). The survey, studying the use of pesticides in farmland during 1997 to 2008, showed that in 1997 the use of pesticides in Bangladesh was more than 8,000 tons; it doubled to 16,000 tons in 2000; in 2005-06, it increased to nearly 20,000 tons and in 2008 it rose up to 48,690 tons. The insecticides, being the dominant item, account for 76 percent of the pesticides, and per hectare use of pesticides increased around 598.8 percent and its annual import cost stands nearly at 171.43 million US \$. According to the study, the intensity of pesticide use was found especially higher in vegetable crops in Bangladesh, compared to other countries in the world. Recently, the use of insecticides has considerably increased in vegetables like eggplant, country bean, cucurbits, yard long bean, etc. particularly in their intensive growing areas. The overuse of pesticides has been identified as one of the reasons for the decline in the overall export of vegetables from Bangladesh, the survey said. The study said the residual effect of these toxic chemicals on vegetables and cereals are likely to create different diseases in humans including cancer, skin diseases, kidney diseases and hypertension as its long term effect (Islam, 2010).

First warning signals about pesticides' danger appeared in 1962, Rachel Carson, an American courageous woman and scientist, wrote down her nature observation and pointed out sudden dying of birds caused by indiscriminate spraying of pesticides (DDT**,** half-life of it in soil ranges from 22 to 30 years). Her book, Silent Spring, became a landmark. It changed the existing view on pesticides and has stimulated public concern on pesticides and their impact on health and the environment. Silent Spring facilitated the ban of the DDT in 1972 in the United States. More researches have been done and several dangerous and persistent organic pesticides like Dieldrin, Endosulfan and Lindane have been banned or restricted since early seventies. Environment affects occurs by pesticides in different compartments. Pesticides enter the soil via spray drift during foliage treatment, wash-off from treated foliage, release from granulates or from treated seeds in soil. Some pesticides such as soil fumigants and nematicides are applied directly into soil to control pests and plant diseases presented in soil.

Pesticides can get into water via drift during pesticide spraying, by run-off from treated area, leaching through the soil. In some cases pesticides can be applied directly onto water surface e.g. for control of mosquitoes. Water contamination depends mainly on nature of pesticides (water solubility, hydrophobicity), soil properties, weather conditions, landscape and also on the distance from an application site to a water source. Rapid transport to groundwater may be caused by heavy rainfall shortly after application of the pesticide to wet soils.

Soil microorganisms play a key role in soil. They are essential for the maintenance of soil structure, transformation and mineralization of organic matter, making nutrients available for plants. Soil microorganisms are also able to metabolise and degrade a lot of pollutants and pesticides and thus are of great concern for using in biotechnology. On the other hand, microbial degradation can lead to formation of more toxic and persistent metabolites. Although soil microbial population are characterized by fast flexibility and adaptability to changed environmental condition, the application of pesticides (especially long-term) can cause significant irreversible changes in their population. Inhibition of species, which provide key process, can have a significant impact on function of whole terrestrial ecosystem. Fungicides were found to be toxic to soil fungi and actinomycetes and caused changes in microbial community structure (Liebich *et al*., 2003; Pal *et al*., 2005). Other bacterial species, such as nitrification bacteria, are very sensitive to pesticides influence. Intensive pesticides and fertilizers usage, loss of natural and semi-natural habitats and decreased habitat heterogeneity and all other aspects of agricultural intensification have undoubted impact on biodiversity decline during last few decades.

Plant is a natural source for providing mankind various secondary metabolites with antiviral or vector control activity, which are organic compounds that are not directly involved in the normal growth, development, or reproduction of an organism (Fraenkel, 1959) and often play an important role in plant defense against herbivores (Nancy, 2003; Crozier *et al*., 2006) and other interspecies defenses (Samuni-Blank *et al*., 2012). Secondary metabolites are also of interest because of their use as dyes, fibres, glues, oils, waxes, flavoring agents, drugs and perfumes and they are viewed as potential sources of new natural drugs, antibiotics, insecticides and herbicides.

Secondary metabolites from plants include alkaloids, terpenoids, phenolics, flavonoids, chromenes and other minor chemicals that can affect insect life in several ways. They may disrupt major metabolic pathways and cause rapid death. Essential oils and especially their main compounds monoterpenoids offer promising alternatives to classical fumigants (Papachristos and Stamopoulos, 2003). These compounds may act as contact insecticides (Huang and Ho, 1998; Chun *et al*., 2000; Tripathi *et al*., 2000; Papachristos and Stamopoulos, 2002a), antifeedant or repellent effects (Kim *et al*., 2003a,b; Park *et al*., 2003a,b) and may also affect some biological parameters, such as growth rate, life span and reproduction (Regnault-Roger and Hamaoui, 1995; Gurusubramanian and Krishna, 1996; Pascual-Villalobos, 1996).

Like pests there are vectors that may also cause damage to crops. Vector is an organism, often an invertebrate arthropod that transmits a pathogen from reservoir to host. Viruses that are transmitted between vertebrate or plant hosts by feeding insects (vectors) can replicate within both their host and their vector. Viruses cause many diseases of international importance. Amongst the human vector like mosquitoes, houseflies carry several diseases like malaria, yellow fever, dengue, diarrhea, dysentery, conjunctivitis, typhoid fever, etc.

Viruses also cause many important plant diseases and are responsible for huge losses in crop production and quality in all parts of the world. Infected plants may show a range of symptoms depending on the disease but often there is leaf yellowing (either of the whole leaf or in a pattern of stripes or blotches), leaf distortion (e.g. curling) and/or other growth distortions (e.g. stunting of the whole plant, abnormalities in flower or fruit formation). Plant viruses pose some of the most severe threats to world agriculture. Because they invade the crop's cells and cloak themselves with the plant's normal life processes, they are far more difficult to control than free-living organisms, such as bacteria, protozoa, or fungi. Plant viruses can cause severe yield loses to the cereal, vegetable, fruit and floral industries and substantially lessen the quality of crop products. Due to virus infection, losses of over 1.5 billion \$ are reported in rice in South-east Asia (Hull, 2002).

Vectors of plant viruses are taxonomically very diverse and can be found among arthropods, nematodes, fungi and plasmodiophorids (Froissart *et al*., 2002 ; Hull, 2002). Arthropod vectors that transmit most plant viruses are aphids, whiteflies, leafhoppers, thrips, beetles, mealybugs, mirids and mites (Spence, 2001), the most common being aphids with more than 200 vector species identified (Ng and Perry, 2004). More than half of the nearly 550 vector-transmitted virus species recorded so far are disseminated by aphids (55%), 11% by leafhoppers, 11% by beetles, 9% by whiteflies, 7% by nematodes, 5% by fungi and plasmodiophorids and the remaining 2% by thrips, mites, mirids or mealybugs (Astier *et al*., 2001).

Aphids or plant lice are the most destructive insect pests on cultivated plants in temperate regions (McGavin, 1993). The damage they do to plants has made them enemies of farmers and gardeners the world over, though from a zoological standpoint they are a highly successful group of organisms (Piper, 2007). Their success is due in part to the asexual reproductive capabilities of some species. About 4,400 species of 10 families are known. Historically, far fewer families were recognized, as most species were included in the family Aphididae. Around 250 species are serious pests for agriculture and forestry as well as an annoyance for gardeners. They vary in length from 1 to 10mm (0.04 to 0.39 inch). Many of the virus diseases are transmitted by homopterous insects, of which aphids constitute an important group (Miyazaki, 2001).

The mosquitoes are a family of small, midge-like flies: the Culicidae. Although a few species are harmless or even useful to humanity, most are considered a nuisance because they consume blood from living vertebrates, including humans. The females of many species of mosquitoes are blood-sucking pests. In feeding on blood, some of them transmit extremely harmful human and livestock diseases, such as malaria, yellow fever and filariasis. Some authorities argue accordingly that mosquitoes are the most dangerous insects on Earth (Michigan Mosquito Control Organization, 2013). Over 3,500 species of mosquitoes have already been described from various parts of the world (Lesley, 1993). Many scientists have suggested that complete eradication of mosquitoes would not have serious ecological consequences (Wilcox and Ellis, 2006; Janet, 2010).

Enterococcus faecalis can be potentially acquired and transmitted to fresh feed by *Tribolium castaneum* adults. It is a worldwide pest of stored products, particularly food grains, and a model organism for food safety research. These adults can serve as potential vectors of antibiotic-resistant *Enterococci* within the feed manufacturing environment. Therefore, it is important to follow proper pest management practices to reduce potential insect vectors in feeds and in the feed manufacturing environments (Channaiah *et al*., 2009). These disease-bearing organisms are known as vectors.

An investigation of *Artemia salina* (Brine shrimp nauplii) as a possible vector for white spot syndrome virus (WSSV) transmission to *Penaeus indicus* (Indian prawn) and *Macrobrachium rosenbergii* nodavirus (MrNV) and its associated extra small virus (XSV) transmission to *M. rosenbergii* (giant river prawn) post-larvae (Sudhakaran *et al*., 2006; Hameed *et al*., 2002). These diseases are highly lethal and contagious, killing shrimps quickly. Outbreaks of this disease have wiped out within a few days the entire populations of many shrimp farm throughout the world.

Vector control is a method to limit the population of certain mammals, birds, insects or arthropods which transmit disease pathogens. The most frequent type of vector control is mosquito control using a variety of strategies. For diseases where there is no effective cure, such as West Nile Virus and Dengue fever, vector control remains the only way to protect populations. However, even for vector-borne diseases with effective treatments the high cost of treatment remains a huge barrier to large amounts of developing world populations. Despite being treatable, malaria has by far the greatest impact on human health from vectors. In Africa, a child dies every 45 seconds of malaria (World Health Organization, 2010). In countries where malaria is well established the World Health Organization estimates countries lose 1.3% annual economic income due to the disease. Both prevention through vector control and treatment are needed to protect populations.

As the impacts of disease and virus are devastating, the need to control the vectors in which they carried is prioritized. Vector control in many Third World countries can have tremendous impacts as it increases mortality rates, especially among infants (World Health Organization, 2009). Because of the high movement of the population, disease spread is also a greater issue in these areas (Walsh *et al.*, 1980).

Curbing population of the vector species can be done in many ways, such as:

i) Habitat control- while removing or reducing areas where the vector species can easily breed, e.g. stagnant water removal, destruction of old tires and cans which were serving as mosquito breeding grounds. Habitat modification includes harbourage alteration and source reduction can be used for mosquito control. Harbourage alteration renders the sites unsuitable for resting of adult mosquitoes and source reduction changes the larval habitat so that mosquito oviposition, hatching and larval development are prevented. Accessibility of water to adult mosquito can be altered or eliminated by ditching, draining, covering and filling. Shredding of disused tires, proper disposal of water containers, alteration of flow rate of water, disturbance of water surface, removal of shelters, such as vegetation and refuse in water bodies etc. can interfere the breeding of mosquitoes. Larval habitats of mosquito vary in size. Some of the water bodies cannot be covered, filled or drained because of ecological or technical reasons. It may be too costly to drain or fill the water bodies. Converting sloping edges of ponds/pools with exposure of muddy areas to almost vertical banks with deepwater can reduce the breeding of *Aedes* mosquitoes. Increase sunlight on water by trimming overhanging vegetation prevents breeding of mosquitoes which prefer shaded habitats. Removal of rooted and floating vegetation also reduces breeding of mosquitoes.

ii) Reducing contact- while limiting exposure to insects or animals that are known disease vectors can reduce infection risks significantly, e.g., bed nets, window screens on homes, or protective clothing can help reduce the likelihood contact with vectors (to be effective, this requires education and promotion of methods amongst the population to raise the awareness of vector threats).

iii) Chemical control- insecticides, larvicides, rodenticides and repellents can be used to control vectors, e.g. larvicides can be used in mosquito breeding zones, insecticides can be applied to house walls or bed nets, and use of repellents can reduce incidence of insect bites and thus curb infection.

iv) Biological control- means the use of natural vector predators, such as bacterial toxins or botanical compounds, can help control vector populations, e.g. using fish that eat mosquito larvae or reducing breeding rates by introducing sterilized male tsetse flies have been shown to control vector populations and reduce infection risks (Vreysen *et al.*,2000). Shreth *et al.*, (2009) suggested use of Neem products and

Lantana products to protect plants against aphids. Ladybugs and their larvae have a voracious appetite for aphids. A single ladybird can eat about 100 aphids a day. Lacewing larvae are also quite fond of eating aphids.

v) Genetic control- this method is usually directed against adult mosquito, while sterile-male release technique is being studied in some countries.

vi) Other methods- giving emphasis on environmental modifications including straightening of watercourse and maintaining a sunlit water surface is made for prevention of malaria vectors breeding. Proper management of small containers, clearing of choked drains, filling of small holes etc. are the methods adopted and promoted for preventing the breeding of dengue fever vectors locally. Draining of water and keeping ditches and ponds free from aquatic vegetation are the methods used for controlling the vectors of Japanese encephalitis breeding in the territory.

Traps may be used for capturing adult mosquitoes. The attractant used could be the carbon dioxide released by breaking down propane into water and carbon dioxide. The warm water vapours with carbon dioxide attract biting insects, such as mosquitoes. Octenol, 1-octen-3-ol, has been used as attractant for attracting mosquitoes over a distance of about 30m down-wind from the trap. This attractant mainly attracts zoophagous mosquitoes. Some traps have a dim light as attractant. As light is not very attractive to mosquitoes, some mosquito traps have a fan to suck the insects flying nearby into some collection chamber or bag. Setting of the traps can, however, be included as one of the protective measures for people against mosquito attacks. Water traps can be used for aphid control. A yellow pan or bowl filled with water and a few drops of liquid dish soap placed in areas where aphid control is required are a good method for killing aphids with wings.

It has been demonstrated that device emitting sonic energy, in the frequency range of 18 to 36 kilohertz, could cause the air in the spaces inside mosquito larvae to resonate violently. The internal membranes and organs of mosquito larvae are disrupted and air bubbles are formed in the bodies of the larvae. The larvae stop moving quickly and die. However, further tests on the effects of the sonic energy on related non-target aquatic insects or other invertebrates have to be conducted.

By the way, amongst the various methods and techniques mentioned above nonhazardous chemical control method would be the most effective way to control the vector population, while non-hazardous chemical refers to the chemical components

collected from natural resources, i.e. plants. These natural chemicals have considerable potential for vector management because these chemicals are safer than conventional insecticides (Mann and Kaufman, 2012). At the present time there are a number of botanical insecticides being marketed, which are extracted from neem, grapefruit seeds and garlic, among other plants. Bangladesh has a great treasure of promising plants. More than 500 plants growing or available in Bangladesh have been reported to possess medicinal properties of some description or other and have been enumerated in the literature of indigenous drug (Ghani, 1998). Many plant extracts of terrestrial origin have been reported to suppress mosquito larval population.

In recent years, use of environment friendly and biodegradable natural insecticides of plant origin has received renewed attention as agents for the control of vectors. During a screening program for new agrochemicals from Chinese medicinal herbs and local wild plants, the ethanol extract of *Evodia rutaecarpa* (Rutaceae) unripe fruits were found to possess larvicidal activity against the mosquito larvae of the Culicidae mosquito *Aedes albopictus* (Liu *et al*., 2012). Essential oil from the leaves of *L. camara* was reported to possess adulticidal activity against *Aedes aegypti*, *Culex quinquefasciatus*, *Anopheles culicifacies*, *An*. *fluvialitis* and *An*. *stephensi* mosquitoes (Dua *et al.*, 2010). Kamaraj *et al*., 2011 reported that the ethyl acetate and methanol extract of *Annona squamosa* stem bark, ethyl acetate and methanol extract of *Chrysanthemum indicum* leaf, acetone and ethyl acetate extract of *Tridax procumbens* whole plant have the potential to be used as an eco-friendly approach for the control of *Anopheles subpictus* and *Culex tritaeniorhynchus*.

The larvicidal effect of CH3OH extracts of *Cleome viscosa* against the mosquito larvae of *Culex* sp. were found promising (Islam *et al*., 2014). Methanol extract of *Azadirachta indica* was found most potential against mosquito larvae and it can be use as alternate potential to synthetic insecticides (Batabyal *et al*., 2007). The acetone, chloroform, ethyl acetate, hexane and methanol extracts of leaf and flower of *Ocimum sanctum* were studied against the fourth instar larvae of *Aedes aegypti* and *Culex quinquefasciatus*. The highest larval mortality was found in case of leaf extract against *A. aegypti* and *C. quinquefasciatus* (Anees, 2008). The effective adult mortality was observed in case of methanol extract of *A. indica*, ethyl acetate extract of *D. biflorus* and ethyl acetate and hexane extract of *Z. zerumbet* against *C. gelidus* and *C. quinquefasciatus.* The promising larval mortality was found in case of hexane

extract of *Z. zerumbet*, ethyl acetate extract of *D. biflorus* and methanol extracts of *A. indica* against *C. gelidus* and *C. quinquefasciatus* after 24h of exposures (Kamaraj *et al*., 2010). Crude and ethyl acetate extracts of matured seed coat of *Cassia sophera* was tested against *Culex quinquefaciatus*. All the graded concentration (0.6%, 0.7%, 0.8%, 0.9%, 1%) showed significant larval mortality (p<0.05). The results support that the tested plant extract can be used for the control of *Culex quinquefaciatus* (Kundu *et al*., 2013) if treated at the larval stage. Mosquito Larval mortality up to 93.33% and reduction of egg hatchability was observed in case of *Acacia nilotica* extract by Zaitoun *et al*. (2012). The leaf extract of *Ageratina adenophora* was more toxic to both the *Aedes. aegypti* and *Culex quinquefasciatus* and could be effectively used for the control of mosquito (Mohan and Ramaswamy, 2007).

Herbaceous plant like *Polygonum hydropiper* L. is important for antiviral activity due to the presence of some secondary metabolites like alkaloids, glycosides, tannins, volatile oils, minerals and vitamins in their cells and tissues. According to Das *et al*., 2008, aphidicidal activity of hot water extracts of *P. hydropiper* and *A. indica* were found to be effective $(87.6-94.5 \text{ and } 80.47-89.6\% \text{ mortality respectively, } P < 0.01$. Ethanol and water extracts of five medicinal and ornamental plant species namely, *Aerva lanata*, *Ruta chalepensis*, *Fagonia arabica*, *Malva parviflora* and *Calotropis procera* were evaluated against pomegranate aphid, *Aphis punicae* Passerini under laboratory conditions*.* Results indicated that the ethanol extract of *R. chalepensis* (wp) showed the highest repellency (75) and mortality (79.5) at 0.015% concentration (Moawad and Barty, 2011).

Insects cause extensive damage to stored grains and their value added products. Among the stored grain pests *Sitophilus oryzae* (L.), *Callosobruchus chinensis* (Fab.) and *Tribolium castaneum* (Herbst) are considered as destructive pests in India. Plants may provide alternatives to currently used insect control agents as they constitute rich source in bioactive molecules. *Lantana camara*, an erect shrub, which grows widely in the tropics, exhibits insecticidal activity against several insects (Rajashekar *et al*., 2012). Plants are the natural factories for the synthesis of variety of bioactive compounds. This diverse chemical setup of the plants speaks their important role as biomedicine. These biomolecules are often toxic to both plants and animals. *Polygonum persicaria* and *Polygonum plebejum* show significant insecticidal activities against *Tribolium castaneum* (Hussain *et al*., 2010). Toxicity, repellency and residual effects of chloroform and ethylalcohol extracts of Bishkatali were evaluated

against the red flour beetle and showed strong repellency against *T. castaneum* in which chloroform extract was better than ethyl alcohol extract (Kundu *et al.*, 2007). Three plant species, leaf of *Polygonum hydropiper* Linn. (Bishkatali), *Vitex negundo* Linn. (Nishinda) and *Aphanamixis polystachya* (Pithraj) extracted with water and acetone were evaluated for their repellent and feeding deterrent activity against adult red flour beetle, *Tribolium castaneum* (Herbst). It was observed that Biskatali (Leaf/water extract) have strong repellent and feeding deterrent effect followed by Nishinda and Pithraj (Islam *et al.*, 2000). Gallardo *et al.* (2011) showed that essential oils (EOs) from *Tagetes lucida*, *Lepechinia betonicifolia*, *Lippia alba*, *Cananga odorata*, and *Rosmarinus* were repellent, followed a dose-response relationship and EOs from *C. odorata* and *L. alba* were the most active repellents against *T. castaneum*. The methanolic extract of *Calendula arvensis* was screened for its toxic potential against *Lemna minor*, *Artimia salina* (Brine shrimps) and exhibited moderate level of cytotoxicity (LD₅₀ value 9.23μg/ml) (Ullah, 2012). Ethanol extract of *Pouzolzia* zeylanica (L.) Benn possesses significant cytotoxic activity with the LC₅₀ value of 6.1 μg/ml and the LC⁹⁰ value of 12.2μg/ml (Saha *et al*., 2012a). Chloroform extracts of the fruit shell, leaves, root bark, root wood, seeds, stem bark and stem wood of *Derris indica* Bennet were tested against the brine shrimp, A*rtemia salina* nauplii. All the test extracts of *D. indica* were found to be effective (Mondal *et al*., 2012b)*.* For brine shrimp lethality bioassay different concentrations (10, 100 and 1000ug/ml) of the methanolic extract of medicinal herb *Ajuga parviflora* were used and toxicity was found (Rahman *et al*., 2013). The Petroleum Ether, chloroform and Methanol extracts of *Cleome viscosa* (root, aerial part and fruit) have been thoroughly screened through residual film assay and repellent activity test against brine shrimp lethality test against *A. salina nauplii*; The cytotoxic effect of *C. viscosa* extracts against the brine shrimp nauplii were found promising reported by Islam (2014). Crude ethanol extracts of the rhizome of *Z. zerumbet* (L) Smith showed the highest cytotoxicity (LC⁵⁰ was 1.24μg/mL) against brine shrimp nauplii (Hossain *et al*., 2012).

Under the shade of some previous works, similar to which are mentioned above, a basic pathway from the plants to the bioactive constituents has been established by Hostettmann and his group in 1995. However, the extensive use of synthetic organic insecticides during the last five decades have resulted in environmental hazards and also in the development of physiological resistance in major pest insect and vector species.

Fig. 1.1: The basic pathway from the plant to the bioactive constituents (Hostettmann *et al.,* 1995)
This has necessitated the need for search and development of environmentally safe, biodegradable, low cost, indigenous methods for vector control, which can be used with minimum care by individual and communities in specific situations.

Being situated in the tropics Bangladesh has a biodiversity rich enough and a huge number of its plants are being used in the traditional system of folk medicine, as well as in the control of crop pests. Thus, in this research endeavor plants of Bangladesh having vector control potential will be selected for the investigation of their bioactive properties as well as of other toxicological aspects used in the pesticide technology. The selected plants will be subjected for extraction with different organic solvents for the isolation, purification (by using chromatographic techniques through activityguided fractionation) followed by characterization of the bioactive constituents.

However, a primary screening through insecticidal, larvicidal, insect repellency and antibacterial tests will be carried out to trace presence of biologically active components in some selected indigenous plants and the plant(s) found to possess promising bioactive component(s) (especially of vector control potential) given special reference in this dissertation. Thus, through literature search and taking into consideration the results obtained by the *Ad Hoc* experiments done in the Crop Protection and Toxicology Laboratory, University of Rajshahi, Bangladesh the following plants were selected: *Evolvulus nummularius* (wp) [01: 05.07.2012], *Lantana camara* (ap and r) [27: 30.06.2007], *Mentha piperita* (wp) [18: 03.04.2006], *Mimosa pudica* (wp) [37: 10.12.2011], *Parthenium hysterophorus* (wp) [48: 23.07.2008], *Phyllanthus niruri* (wp) [11: 09.07.2010], *Polygonum hydropiper* (wp) [37: 15.04.2013], *Pouzolzia zeylanica* (wp) [70: 07.06.2014], *Synedrella nodiflora* (wp) [27: 01.11.1991], *Zingiber zerumbet* (ap and rh) [not available]. Here, number along with date besides the names of plants are the voucher numbers of specimens and dates of collection which are kept in the herbarium of the Department of Botany, University of Rajshahi; and these collection helped us to identify the test plants through direct comparison.

1.1. Background information on the test plants

1.1.1. *Evolvulus nummularius*

1.1.1.1. Synonym: *Convolvulus nummularius* L.; **English name:** Round leaf Bindweed**; Local name:** Bhui Akra (ভূঁই আকরা).

1.1.1.2. Habitat and geographical distribution: *E. nummularius* can be seen on the hill slopes, edges of fields, roadsides and railway embankments. It is widely distributed in India, Nepal, Bhutan, tropical America and Africa. In Bangladesh, it is found in all districts.

1.1.1.3. Morphology: A perennial herb with prostrate stem, often pilose at the nodes with short tricomes to glabrate. Leaves 0.5 -1.3 x 0.5 -1.1cm, broadly ovate to orbicular, petioles 1-6mm long, apex rounded to slightly emerginate, base rounded to subcordate, glabrous to puberulent beneath. Flower 1-2 in axils, rarely more, up to 5mm high, pedicels 2-6mm long. Flowers are very tinny, around 0.5-0.7cm size, color is milky white. Sepals elliptic ovate to elliptic-oblong, pubescent, ciliate. Corolla broadly campanulate, up to 10mm wide, white. Fruit a globose capsule, 3-4mm across, often reflexed at maturity. Seeds 1.5mm long, brownish to black, subglobose. Flowering and fruiting throughout the year. It binds the ground while creeping around (Ahmed *et al*., 2009c).

1.1.1.4. Systematic position:

Kingdom: Plantae

Superdivision: Spermatophyta Division: Magnoliophyta Class: Magnoliopsida Order: Solanales Family: Convolvulaceae Genus: *Evolvulus* L. Species: *E. nummularius* (L.)

Plate 1.1: *Evolvulus nummularius*

1.1.1.5. Uses: The whole plant is used as a medicine for hysteria, to cure burns, cuts, wounds and scropion stings (Jain, 1991). In Nepal, the paste of the plant is used to treat scabies (Manandhar, 2002). *E. nummularius* has been pharmacologically reported to possess antihelminthic activity (Dash *et al.*, 2003) wound healing activity (Saini *et al.*, 2007), poor sedative and anticonvulsant properties (Chitralekha *et al.*, 1964). Three new compounds, 1-3 along with βsitosterol and its glucoside, stigmasterol, *d*-mannitol, ursolic acid and oleanolic acid have been isolated from the aerial parts of *E. nummularius* (Biswanath *et al.*, 2007). Methanol extract of *E. nummularius* has antibacterial activity against *Escherichia coli* and *Bacillus subtilus* (Pavithra *et al.*, 2009).

1.1.2. *Lantana camara*

1.1.2.1. Synonym: *Lantana aculeata* L., *Lantana armata*; **English name:** Lantana; **Local name: Chotra (**চাতরা).

1.1.2.2. Habitat and geographical distribution: *L. camara* are very much common in the wild and along footpaths, deserted fields and farms. It is distributed in Mexico, Central America, the Greater Antilles, The Bahamas, Colombia and Venezuela, Texas in the United States, tropical and warm regions worldwide.

1.1.2.3. Morphology: *L. camara* is a low erect or subscandent vigorous shrub with tetrangular stem. It has a strong odor of black currents. Plant grows up to 1 to 3m and it can spread to 2.5m in width. Leaves are ovate or ovate oblong, 3-8cm long by 3-6cm wide and green in color. Leaves and stem are covered with rough hairs. Small flower held in clusters (called umbels). Color usually orange, the calyx is small, corolla tube slender, the limb spreading 6 to 7mm wide and divided in to unequal lobes. Stemen four in two pairs, included and ovary two celled, two ovuled. Inflorescences are produced in pairs in the axils of opposite leaves. Inflorescences are compact, dome shaped 2-3cm across. Root system is very strong and it gives out new fresh shoots even after repeated cuttings (Sastri, 1962).

1.1.2.4. Systematic position:

Kingdom: Plantae Superdivision: Spermatophyta Division: Magnoliophyta Class: Magnoliopsida Order: Lamiales Family: Verbenaceae Genus: *Lantana* L. Species: *L. camara* (L.)

Plate 1.2: *Lantana camara*

1.1.2.5. Uses: Foliage contains pentacyclic triterpenoids cause hepatotoxicity and photosensitivity in grazing animals such as sheep, goats, bovines and horses. The berries are edible when ripe. Ingestion of *L. camara* (including unripe berries) is not associated with significant human toxicity (Carstairs *et al.*, 2010). The methanolic extract of *Lantana camara* leaves shown healing of gastric ulcers and also prevents development of duodenal ulcers in rats (Sathish *et al.*, 2011). Extracts of the fresh leaves are antibacterial and are traditionally used in Brazil as an antipyretic, carminative and in the treatment of respiratory system infections (Barreto, 2010).

1.1.3. *Mentha piperita*

1.1.3.1. Synonym: *Mentha crispa* L.; **English name:** Peppermint, Mint; **Local name:** Pudina (পুদিনা).

1.1.3.2. Habitat and geographical distribution: Peppermint typically occurs in moist habitats including stream sides and drainage ditches. It is native to Europe and Asia. Outside of its native range, areas where peppermint was formerly grown for oil often have an abundance of feral plants and it is considered invasive in Australia, the Galapagos Islands, New Zealand and in the United States in the Great Lakes region, noted since 1843.

1.1.3.3. Morphology: Peppermint is an herbaceous rhizomatous perennial plant growing to 30–90cm (12–35 inch) tall, with smooth stems, square in cross section. The rhizomes are wide-spreading, fleshy and bare fibrous roots. The leaves are from 4–9cm (1.6–3.5 inch) long and 1.5–4cm (0.59–1.6 inch) broad, dark green with reddish veins and with an acute apex and coarsely toothed margins. The leaves and stems are usually slightly fuzzy. The flowers are purple, 6–8mm (0.24–0.31 inch) long, with a four-lobed corolla about 5mm (0.20 inch) diameter; they are produced in whorls (Verticillasters) around the stem, forming thick, blunt spikes. Flowering is from mid- to late summer. Both leaves and flowers have a characteristic, aromatic fragrance. Peppermint is a fast growing plant once it sprouts, it spreads very quickly.

1.1.3.4. Systematic position:

- Kingdom: Plantae
	- Superdivision: Spermatophyta Division: Magnoliophyta Class: Magnoliopsida Order: Lamiales Family: Lamiaceae Genus: *Mentha* L.

Species: *M. piperita* (L.) **Plate 1.3:** *Mentha piperita*

1.1.3.5. Uses: Peppermint is considered to have astringent, antiseptic, emetic and stimulant qualities. It has a long history of medicinal use, especially in treatment of digestive complaints. Tea made from leaves and flowers can be an excellent remedy for treatment of indigestion, cramps, flatulence, nausea, vomiting and colic. It has a soothing effect on the stomach, and can also be an appetite stimulant. Topical application of peppermint oil can reduce arthritis, rheumatism and chronic joint pain. Due to its antiseptic properties, peppermint can be helpful in the relief of toothache and in treatments of cavities. Peppermint vapors and inhalers are very helpful in cases of nasal and sinus congestions, laryngitis and bronchitis. Peppermint oil has a high concentration of natural pesticides, mainly pulegone (Found in *Mentha arvensis var. piperascens* Cornmint, Field Mint, Japanese Mint and to a lesser extent-6,530 ppm in *Mentha piperita* (Krieger, 2001). The chemical composition of the essential oil from peppermint (*Mentha piperita* L.) was analyzed by GC/FID and GC-MS. The main constituents were menthol (40.7%) and menthone (23.4%). Further components were (+/-)-menthyl acetate, 1,8-cineole, limonene, beta-pinene and beta-caryophyllene (Schmidt *et al.*, 2009).

1.1.4. *Mimosa pudica*

1.1.4.1. Synonyms: *Mimosa tetrandra* Humb. & Bonpl. ex Willd. *Mimosa pudica* L. var. *tetrandra* (Willd.) DC. *Mimosa unijuga* Duch. & Walp. *Mimosa pudica* L. var. U*nijuga* (Duch. &Walp.) Griseb.; **Common name:** The touch-me-not and sensitive plant.; **English name:** Lajjabati (লǵাবতী).

1.1.4.2. Habitat and geographical distribution: *M. pudica* are found in the dry open grassy fields, pastures, roadsides and fallow lands. It is pantropical weed of South American origin, distributed to all the tropical countries of the world. In Bangladesh, it is a common weed, growing all over the country.

1.1.4.3. Morphology: A low, spreading and prostrate annual or perennial herb, up to 1m tall, sometimes sub-shrubby herb, erect or scrambling, sometimes rooting at the nodes, branches more or less herbaceous but old stem woody, glandular hairy and prickly, prickles curved, compressed. Leaves sub-digitately pinnately compound, very sensitive to touch, stipulate, stipules linear-lanceolate, 7-8mm long, rachis 2.5-3.0cm long, sometimes up to 5cm long, sulcate, hispid and prickly, without throne at the junctions of the pinnae but sometimes with a few recurved prickles on the internodes, pinnae l-2 pairs, about 2.5-5.0cm long, sessile, at the end of the rachis, leaflets 12-25 pairs, sessile, 5-10×1.5-2.0mm, linear-oblong to sub-falcate, acute, coriaceous, glabrous above, faintly hairy on margin and beneath. In florescence of axillary pedunculate globose heads, solitary or paired in the axils of the distal leaves, peduncles about 2.5-3.5cm long, yellowish-green, densely hirsute, bearing heads 6- 8mm across, solitary or in a axillary pair. Flowers small, sessile, tubular, pinkish. Calyx minute, less than 0.lmm long, inconspicuous. Corolla pink, l.5-2.5mm long, narrowly campanulate, lobes 4, oblong-ovate, obtuse or rounded. Stamens 4, much exserted. Ovary 0.3-0.5mm long, glabrous. Fruit a pod, l5-l8×2-4mm, oblong, flat, straight or recurved wavy margin on both sides, dark brown with bright brown prickly bristle margin when dry, develop in cluster from the same head, indehiscent. Seeds 3-5 per pod, light brown, smooth, glossy. Flowering and fruiting: September-December (sometimes throughout the year) (Ahmed *et al*., 2009a).

1.1.4.4. Systematic position:

Kingdom: Plantae Superdivision: Spermatophyta Division: Magnoliophyta Class: Magnoliopsida Order: Fabales Family: Fabaceae Genus: *Mimosa* L. Species: *Mi. pudica L.*

Plate 1.4: *Mimosa pudica*

1.1.4.5. Uses: *Mimosa pudica* is a good soil binder and it has been tried as green manure. In Thailand, the species was introduced to check soil erosion along embankments (Nielsen, 1985). The roots contain tannin, which is used as medicine in the Philippines; also the plant is used as a cover crop along roadside in Thailand (Nielsen, I992). Due to the sensitivity of leaves it is valued as an interesting ornamental plant. The local people of Orissa, India take extract of powdered roots and leaves to cure fever due to spleen enlargement, the residue is also applied externally for the same purpose over the stomach (Srivastava and Rout, 1994). It is used in the treatment of leprosy, dysentery, vaginal and uterine complaints and inflammations, burning sensation, asthma, leucoderma, fatigue and blood diseases. Decoction of root is used as gargle to reduce toothache. It is very useful in diarrhea, amoebic dysentery, bleeding piles and urinary infections. This review gives a brief compilation of its phytochemical and pharmacological activities (Joseph *et al.*, 2013).

1.1.5. *Parthenium hysterophorus*

1.1.5.1. Synonym: Bastard feverfew; **English name:** Bitter weed, *Parthenium* weed; **Local name:** Gajor Ghash Ful, *Parthenium* (পােথŪিনয়াম).

1.1.5.2. Habitat and geographical distribution: This noxious weed is often spotted on abandoned lands, developing residential colonies around the towns, railway tracks, roads, drainage and irrigation canals, etc. This weed grows luxuriantly in established gardens, plantations and vegetable crops. This erect, ephemeral herb known for its vigorous growth and high fecundity especially in warmer climates is a native of north-east Mexico and is endemic in America. Within the past century it has found its way to Africa, Australia, Asia and Pacific Islands and has now become one of the world's seven most devastating and hazardous weeds. Due to its high fecundity a single plant can produce 10,000 to 15,000 viable seeds and these seeds can disperse and germinate to cover large areas.

1.1.5.3. Morphology: *Parthenium hysterophorus* is a prolific weed belonging to Asteraceae family. This erect ephemeral herb can grow up to 1.5-2m high and has a deep tap root. It is light green with branching stems, finely lobed leaves, 3-20cm long and 2-10cm wide. Once stem elongation is initiated, smaller leaves are produced and the plant becomes multi-branched in its extremities. Flower heads are small (4mm across) and numerous in open panicles. *P. hysterophorus* reproduces by seeds and is known to be highly prolific, and produces 15,000 seeds (GISD Database, 2010).

1.1.5.4. Systematic position:

Kingdom: Plantae

Superdivision: Spermatophyta Division: Magnoliophyta Class: Magnoliopsida Order: Asterales Family: Asteraceae Genus: *Parthenium* L.

Species: *P. hysterophorus* L. **Plate 1.5:** *Parthenium hysterophorus*

1.1.5.5. Uses: *P. hysterophorus* is a noxious weed in America, Asia, Africa and Australia. This weed is considered to be a cause of allergic respiratory problems, contact dermatitis, mutagenicity in human and livestock. Crop production is drastically reduced owing to its allelopathy. Also aggressive dominance of this weed threatens biodiversity. Eradication of *P. hysterophorus* by burning, chemical herbicides, eucalyptus oil and biological control by leaf-feeding beetle, stem-galling moth, stem-boring weevil and fungi have been carried out with variable degrees of success. Recently many innovative uses of this hitherto notorious plant have been discovered. *P. hysterophorus* confers many health benefits, *viz* remedy for skin inflammation, rheumatic pain, diarrhoea, urinary tract infections, dysentery, malaria and neuralgia. Its prospect as nano-medicine is being carried out with some preliminary success so far. Removal of heavy metals and dye from the environment, eradication of aquatic weeds, use as substrate for commercial enzyme production, additives in cattle manure for biogas production, as biopesticide, as green manure and compost are to name a few of some other potentials. The active compounds responsible for hazardous properties have been summarized.

Isolation and structural elucidation of the active principles of *P. hysterophorus* is required to determine their chemical properties. Chemical analysis of *P. hysterophorus* has indicated that all its parts including trichomes and pollen contain toxins called sesquiterpene lactones (SQL).

Maishi *et al.*, (1998) reported that *P. hysterophorus* contains a bitter glycoside parthenin, a major sesquiterpene lactone. Other phytotoxic compounds or allelochemicals are hysterin, ambrosin, flavonoids such as quercelagetin 3,7 dimethylether, 6-hydroxyl kaempferol 3-0 arabinoglucoside, fumaric acid. P-hydroxy benzoin and vanillic acid, caffeic acid, p courmaric, anisic acid, p-anisic acid, chlorogenic acid, ferulic acid, sitosterol and some unidentified alcohols. Parthenin, hymenin and ambrosin are found to be the culprits behind the menacing role of this weed in provoking health hazards (Patel, 2011).

1.1.6. *Phyllanthus niruri*

1.1.6.1. Synonyms: *Phyllanthus fraternus* Webster, *Ph*, *sellowianus*, *Ph. carolinianus*, *Ph. kirganella*, *Ph. lathyroides*, *Ph. lonphali, Nymphanthus niruri*; **English name:** Stone-breaker; **Local name:** Bhui-amla (ভু*u*ইআমলা)**.**

1.1.6.2. Habitat and geographical distribution: It occurs in Sandy clay soil in moist habitat. It is available in Africa, India, Pakistan, Saudi Arabia and the West Indies. In Bangladesh, this species is found throughout the country.

1.1.6.3. Morphology: A monoecious, erect annual herb, up to 70cm high, branches angular. Leaves stipulate, stipules 1.0-1.2mm long, lanceolate, scarious, acute,, petiolate, petioles very short, leaf blade elliptic-oblong to elliptic-oblanceolate, 5-12 x 2-5mm, obtuse or rounded at the apex and base or sometimes tapering to the base,, membranous, lateral nerves 4-7 pairs, indistinct, dark green above, paler and greyish beneath. Flowers yellowish, very numerous, axillary, the males l-3, the female solitary. Male flowers pedicellate, pedicels l mm long, sepals 6, suborbicular-obovate, c 0.5 x 0.5mm, rounded, midrib yellow, disc glands 6, verruculose, lobulate, stamens 3, filaments united into a short column, anthers more or less horizontal, dehiscing transversely. Female flowers with pedicels 1.4-1.9mm long, sepals 6, unequal, l.0-l.5 x 0.4-0.5mm, oblong-oblanceolate, rounded, white, disk thin, flat, irregularly deeply lobed into 6-l0 segments, some crenate and broad, some triangular and bifid, some others linear and entire, ovary subglobose, c 1mm in diameter, smooth, styles minute, free, adpressed or ascending, the lobes recurved. Fruits trilobatesubglobose, l.5-2.5mm in diameter, smooth, olivaceous or stramineous. Seeds l.0 x 0.6mm, longitudinally 7-8 ridged on the back, ochreous-fulvous (Ahmed *et al*., 2009d).

1.1.6.4. Systematic position:

Kingdom: Plantae Superdivision: Spermatophyta Division: Magnoliophyta Class: Magnoliopsida Order: Magnoliopsida Family: Euphorbiales Genus: *Phyllanthus* L.

Species: *Ph. niruri* L. **Plate 1.6:** *Phyllanthus niruri*

1.1.6.5. Uses: The plant is used as diuretic in gonorrhoea and other ailments of genito-urinary tract. The fresh root is administered as a remedy of jaundice. Infusion of young shoots is prescribed in dysentery. A poultice of the leaves is applied to bruises and wounds, and with salt it cures scabby infections. An infusion of root and leaves is a good tonic and diuretic. The milky juice is effective for offensive sores. The bark is applied as a purgative (Kirtikar *et al*., I935). In India, the ethnic people of Lodha use the juice of the plant with curd to cure jaundice. They apply a decoction of root with paste of long peppers in the treatment of dysentery, and stem decoction made with water that is obtained after washing rice is used to treat menorrhagia. The Santal ethnic people prescribe a decoction of plant with black peppers 'for genital diseases. In Ghana, the pounded leaves are given to cure gonorrhoea. A decoction of the root and leaves is used in stomach-ache in Haiti (Pal and Jain, 1998). Various bioactivities of this medicinal plant such as antidiabetic (Okoli *et al.*, 2011), antihepatotoxicity, (Ravikumar *et al.*, 2011) antilithic, anti-hypertensive, anti-HIV and antihepatitis B (Bagalkotkar *et al*., 2006; Naik and Juvekar, 2003) have been reported.

1.1.7. *Polygonum hydropiper*

1.1.7.1. Synonyms: *Persicaria fastigiatoramosa*, *Persicaria hydropiper*, *Polygonum fastigiatoramosum*, *Polygonum maximowiczii*; **English name:** Water papper; **Local name:** Bishkatali (িবষকাটালী).

1.1.7.2. Habitat and geographical distribution: It can be seen on shallow water in ponds, ditches etc and in wet places on land. It is distributed in Europe, including Britain, from Norway south and east to N. Africa and temperate Asia. Origin - Native to Eurasia.

1.1.7.3. Morphology: Leaves are alternate, short-petiolate or subsessile, lanceolate to linear-oblong, glabrous, acuminate. Ocrea with ciliate bristles on margin, glabrous to scabrous. Inflorescence are terminal and axillary racemes, loosely arranged, often nodding at tip. Stems are 1m tall, herbaceous, glabrous or with some pubescence above, typically green or reddish, erect to spreading, multiple or single from base, simple to few-branching. The roots are thin and fibrous as the plant is an annual. Flowers are small, sessile in terminal, generally unbranched spikes and blooming in the month of June-July.

1.1.7.4. Systematic position:

Kingdom: Plantae Superdivision: Spermatophyta Division: Magnoliophyta Class: Magnoliopsida Order: Polygonales Family: Polygonaceae Genus: *Polygonum* L. Species: *Po. hydropiper* L.

Plate 1.7: *Polygonum hydropiper*

1.1.7.5. Uses: Smartweed has a long history of herbal use, both in Eastern and Western herbalism. It is not used very often and is seen more as a domestic remedy being valued especially for its astringent properties which makes it useful in treating bleeding, skin problems, diarrhoea, etc. The leaves are anti-inflammatory, astringent, carminative, diaphoretic, diuretic, emmenagogue, stimulant, stomachic, styptic. They contain rutin, which helps strengthen fragile capillaries and thus helps prevent bleeding. The seed is carminative, diuretic and stimulant. The whole plant, either on its own or mixed with other herbs, is decocted and used in the treatment of a wide range of ailments including diarrhoea, dyspepsia, itching skin, excessive menstrual bleeding and haemorrhoids. A poultice of the plant is used in treating swollen and inflamed areas. In Chinese tests, the plant was ranked 20th in a survey of 250 potential antifertility drugs. A homeopathic remedy is made from the leaves. It is used in the treatment of piles, menstrual pains and other menstrual complaints.

Leaves and stems can be used as raw or cooked. They can also be made into an acid peppery condiment. They are very hot. The leaves contain about 7.5% protein, 1.9% fat, 8% carbohydrate, 2% ash. The leaves are said to contain rutin. Seed can be used as raw or cooked. It is rather small and fiddly to utilize. The seed is used as a condiment - a pepper substitute. The sprouted seeds or young seedlings can be used as a garnish or added to salads, they are commonly sold in Japanese markets. They are very hot. A yellow-gold dye is obtained from the stalks.

1.1.8. *Pouzolzia zeylanica*

1.1.8.1. Synonym: *Pouzolzia arnhemica* F. Muell.; *Pouzolzia indica* Gaud; *Parietaria zeylanica* L.; *Parietaria indica* L; **English name:** Graceful Pouzolz's Bush; **Local name:** *Pouzolzia* (পওজলিজয়া).

1.1.8.2. Habitat and geographical distribution: *Pz. zeylanica* is very much mesophytic (*i.e.*, Mesophytes are terrestrial plants which are adapted to neither a particularly dry nor particularly wet environment). It likes to grow roadsides, old fields, waste places, disturbed areas; grasslands, thickets by streams, wet places, sunny and somewhat moist places by rice fields. *Pz. zeylanica* is available throughout Asia. It also found in Polynesia, Papua New Guinea, Australia and around its Islands; it also recorded from Yemen (Socotra). It introduced in Africa and the New World. In Bangladesh, it found throughout the country in fallow lands.

1.1.8.3. Morphology: Graceful Pouzolz's Bush is a perennial herb, very variable in size and habit; sparsely to moderately cover on all parts with appressed and spreading hairs. Stem is erect to sub-erect, rarely prostrate, almost simple or few branched at base, 12-40cm tall hairy to glabrescent herb, growing up to 12-40cm tall but occasionally grows into a shrub about 1m tall; rootstock often tuberous; branches sometimes with short branchlets, strigillose. Leaves are opposite or rarely alternate, with 4-12mm long stalk. Leaf blade is lanceshaped- rhomboid ovate, 1-2.5cm long, 4-15mm broad, wedge-shaped or flat at the base, entire, pointed. Stipules are broadly ovate, 3-5mm long, 3-4mm broad, acuminate, ciliate, clothed in long white hairs. Leaf blades 1-5 \times 0.6-2cm, base rounded to broadly cuneate, apex subobtuse, acute or nearly acuminate. Flowering time July to October. Flowers are minute, in small axillary androgynous clusters, covered with hairs, female stalkless, male with 2mm long stalk. Flower colour varies from green to pale green, beige, white. Sepal cup is 4-lobed and gibbous in male; 4-toothed in female flowers. Calyx 4-lobed and gibbous in male; 4-toothed in female flowers. Four stamens. Anthers reniform, about 0.9 x 0.6mm, reflexed in the bud. Filaments about 1-2mm long. Achenes are white, light to dark yellow or light brown, ovate, 1.5 mm long, enveloped by ribbed sepal cup. Fruits enclosed in the persistent, hairy perianth. Fruits pyriform, about 2.5 x 2mm. Seeds about 2 x 1.8mm, testa thin and glabrous. Cotyledons about 1 x 1.5mm, broader than long, slightly cordate at the base. Radicle straight and thick, about 0.6 x 0.5mm.Cotyledons orbicular, about 7-8mm diam. First pair of leaves opposite, leaf blades ovate, upper surface clothed in numerous 'glandular' humps and long, pale hairs. Stipules hairy; at the tenth leaf stage, Midrib and lateral veins depressed on the upper surface. Leaf blade 3-veined at the base.

1.1.8.4. Systematic position:

Kingdom: Plantae Superdivision: Spermatophyta Division: Magnoliophyta Class: Magnoliopsida Order: Urticales Family: Urticaceae Genus: *Pouzolzia* Gaudich.

Species: *Pz. zeylanica* (L.) Benn **Plate 1.8:** *Pouzolzia zeylanica*

1.1.8.5. Uses: *Pz. zeylanica* is a perennial herbaceous plant and has a reputation for its folk used as a remedy for diarrhea, indigestion, infantile malnutrition, urination difficulties and injuries from falls. Moreover, it is especially useful in conditions such as acute mastitis and pyogenic infections (Tsao and Deng, 2004). However, no chemical and biochemical information concerning *Pz. zeylanica* has been reported.

This species has been used medicinally in Malaysia and Indonesia. Leaves are anthelmintic and vulnerary; used as a cicatrizant for gangrenous ulcers, in syphilis and gonorrhoea. Leaf juice is used as galactagogue. Poultice of the herb is applied to sores, boils and to relieve stomachache (Yusuf *et al.*, 1994).The aerial part of *Pz. zeylanica* was studied by Paul and Saha (2012) to fix the parameters for pharmacognostical standards and they also ensured the presence of alkaloids, glycosides, tannins and flavonoids. *Pz. zeylanica* has also fluorescence properties (Kokoshi *et al.*, 1958; Chase and Pratt 1949). Recently organoleptic study has been done by Paul and Saha (2012), which offer a scientific basis for the traditional use of *Pz. zeylanica* which possess characters like greenish grey, characteristic odour and mucilaginous and slightly bitter taste. Presence of antitumor or pesticidal compounds in this plant is also suggested by Meyer *et al.* (1982).

Other current studies ensured the Anti-Bacterial activity of *Pz. zeylanica*. The ethanol extracts of *Pz. zeylanica* was showing anti-bacterial activity against both gram positive and gram negative organisms (Saha *et al.*, 2012b; Jenny *et al.*, 2012). The ethanolic extracts of *Pz. zeylanica* possessed antifungal activity against six fungal

strains fungal strains (Saha and Paul, 2012a). *Pz. zeylanica* contains flavones, flavonoids, tannin, carotene, carotenoids, ascorbic acid, tartaric, malic and pectic acids, gum, minerals and their salts (Ghani, 2003); alkaloids, glycosides (Paul and Saha, 2012).

1.1.9. *Synedrella nodiflora*

1.1.9.1. Synonym: *Verbesina nodiflora* L.; **English name:** Node weed, Cindrella weed**; Local name:** Cindrella (সিনড়েলা) .

1.1.9.2. Habitat and geographical distribution: *S. nodiflora* originates from the New World tropics, but became naturalized in the Old World in the 19th Century. This plant is found in tropical Africa, tropical and sub-tropical Asia, and introduced in the West Indies. In Bangladesh it is found all over the country.

1.1.9.3. Morphology: A short-lived or long-lived herbaceous plant with weak sprawling stems growing up to 60cm long. Its paired leaves (1-6cm long) are eggshaped in outline or somewhat triangular in shape with sparsely toothed margins. Its stems and leaves are loosely covered in short close-lying hairs. Its small yellow flower-heads (5-10mm across) are borne singly in the upper leaf forks. Its 'seeds' (2- 4mm long) are topped with a pair of spreading awns 1-3mm long. This species reproduces mainly by seed.

1.1.9.4. Systematic position:

Kingdom: Plantae Superdivision: Spermatophyta Division: Magnoliophyta Class: Magnoliopsida Order: Asterales Family: Asteraceae Genus: *Synedrella* Gaertn. Species: *S. nodiflora* (L) Gaertn.

Plate 1.9: *Synedrella nodiflora*

1.1.9.5. Uses: In Indonesia, the leaves of *S. nodiflora* are used as a poultice for sore legs and rheumatism; in Africa the leaves are applied as an embrocation for different oedemas. In Ghana, an infusion of young leaves is used as a laxative. In Indonesia, the juice of the leaves is used for the treatment of earache, and in Africa for treatment of mouth affections such as infected gums. In Papua New Guinea, the root

is chewed against diarrhoea, together with some other herbs. Dislocated bones are massaged daily with sap from the squeezed leaves. In Fiji, a decoction of the leaves is used to treat haemorrhoids and diarrhoea. A decoction of the pounded and cooked roots is drunk as a cough-mixture in Africa and in Barbados. In Colombia, the entire plant is used as an emmenagogue. *S. nodiflora* is not known to be used in Indo-China. In Indonesia tender leaves are used in salad.

Upon steam distillation of the leaves, *S. nodiflora* yields a yellow coloured essential oil (0.02%), with the terpenes ' β -caryophyllene, ' β -farnesene, germacrene-D and ' β cubebene as major components. From the ethanol extract of the whole plant, the triterpenoid saponin nodifloside A (oleanolic acid-3-O- β -D xylopyranosyl-(1forward 4 -' β '-D-glucopyranuronosyl methylate) was isolated, together with the triterpenoid oleanic acid-3-O-' β '-D-glucopyranuronosyl methylate, and the steroids ' β '-sitosterol, stigmasterol, stigmasterol-3-O-'B[']-D-glycoside and rosasterol. *S. nodiflora* also contains a high content of estradiol. An orally administered dried leaf extract of *S. nodiflora* was found to be active as an anti-inflammatory against adjuvantcarrageenan-induced inflammation in rats. It inhibited both acute and chronic phases, especially the chronic phase. The ethanol extract of the entire plant showed analgesic and antipyretic activity in rodents. Furthermore, chloroform extracts of the foliage of *S. nodiflora* acted as a deterrent when tested on three pests of stored grain products: larvae and imagos of *Tribolium confusum*, larvae of *Trigoderma granarium*, and imagos of *Sitophilus granarius*.

1.1.10. *Zingiber zerumbet*

1.1.10.1 Synonym: *Cardamomum spurium* (J. Konig) Kuntze, *Zingiber amaricans* Blume; **English name:** Variegated-leaved zerumbet ginger; **Local name:** Bon-ada (বন*-*আদা)*.*

1.1.10.2. Habitat and geographical distribution: *Z. zerumbet* is found in partial shade in the forests and village thickets. It is distributed in India, Malaysia, Nepal and Sri Lanka. ln Bangladesh, it grows all over the country in forests and village thickets.

1.1.10.3. Morphology: A rhizomatous herb with annual aerial leafy stems and underground tuberous rhizome, pale yellow inside. Leafy stem 0.6-2.0m high. Leaves sessile or subsessile, $20-30 \times 4$ -8cm, lanceolate or oblanceolate, acuminate, usually with some hairs on the lower surface, silky when young, ligules 1.5-3.0cm long, membranous, entire, with a few scattered hairs. Inflorescence radical, peduncles 30- 70cm long, clothed with sheathing bracts, 4-6cm long, lightly pubescent, rounded at the apex, spikes $7-12 \times 4-6$ cm, ovate-oblong, often rounded at the tip. Calyx spathaceous, c 2cm long, shortly 3-toothed, hairy at the base and few bristles at the tip, unilaterally split. Corolla tube 2.5-3.0cm long, petals 3, white or yellowish, 2.0- 2.3cm long, narrowed at the apex. Labellum light yellow or creamy, 3-lobed, mid-lobe c 1.8cm long, broad, suborbicular, bifid, central portion raised, margin crinkled, lateral lobes (staminodes) rotundate or ovate, c 1cm long. Filaments very short, c 3mm long, anthers c 1cm long, beak c 7mm long, curved. Ovary 4 x 2-3mm, glabrous, epigynous gland yellow, 3-5mm long, linear, free (Ahmed *et al.,* 2009b).

1.1.10.4. Systematic position:

Kingdom: Plantae

Superdivision: Spermatophyta Division: Magnoliophyta Class: Magnoliopsida Order: Zingiberales Family: Zingiberaceae Genus: *Zingiber* Mill.

Species: *Z. zirumbet* L. Roscoe

Plate 1.10: *Zingiber zerumbet*

1.1.10.5. Uses: The plant is sold in the market in the trade name "Ekangi", a traditional drug used in the preparation of Ayurvedic and Unani medicines. ln lndia, the rhizome is used in cough, stomachache, asthma, worm infestation, leprosy and skin diseases. Dried powdered rhizome is used as an antidiarrhoeal agent in the Philippines. Decoction is used in asthma and rheumatism. The plant was tested in India against different pathogens. Rhizome was found antiviral and antiprotozoal and the leaf as diuretic. The rhizome contains volatile oil, main constituent of which is zerumbone 35.48%, other major constituents are humulene 17.29% and camphene 16.04% (Oliveros and Cantoria, 1982; Prakash and Mehrotra, 1996; Riswan and Setyowati, 1996.) Rhizome is used for cough and cold in the Chittagong Hill Tracts by the ethnic people. The plant can be easily propagated through rhizome segments.

1.2. Background information on the test organisms

The whole experiment has been designed to carry on screening of the crude extracts of the test plant species on several test organisms for the detection of biological activity keeping an option to show extent of activity by analyzing the data statistically that read on various parameters during the course of the work. The following test steps have been taken into consideration:

Table 1.1. List of the test agents

1.2.1. 'Rust-red flour beetle' (*Tribolium castaneum***)**

T. castaneum (Hbst.), the red flour beetle is Indo-Australian origin (Smith and Whitman, 1992) and is found in temperate region, but will survive the winter in protected places, especially where there is central heat (Tripathi *et al.*, 2001). In the United States, it is found primarily in the southern states. *T. castaneum* is a worldwide and commonest pest of wheat-flour. It is commonly known as 'Rust-red flour beetle'. It is an insect of the family 'Tenebrionidae' under the order 'Coleoptera'.

It is one of the serious pests of stored products. Mouthparts of this pest insect are not adapted to feed on hard whole grains and they are thus found in almost any kind of flour, cracked grains etc. The specific foods of *T. castaneum*, which include wholewheat flour, bran, rice flour, cornmeal, barley flour and oat meals. It also feeds upon dried fruits, dried plant roots, nuts, chocolates, drugs, snuff, cayenne pepper, pulses and prepared cereal foods such as corn flakes (Metcalf and Flint, 1962). Not only pulses and millets, but also cereals are also been attacked by this beetle (Purthi and

Singh, 1950). *T. castaneum*, attack meal, crackers, beans, spices, pasta, cake mix, dried pet food, dried flowers and even dried museum specimens (Via, 1999; Weston and Rattlingroud, 2000).

Although small beetles, about $\frac{1}{4}$ of an inch long, the adults are long-lived and may live for more than three years (Walter, 1990). The red flour beetle is reddish brown in color and its antennae end in a three segmented club (Bousquet *et al.*, 1990). The head of the red flour beetle is visible from above, does not have a beak and the thorax has slightly curved sides.

Plate 1.11: Life cycle of *T. castaneum*

Rearing temperatures	30° C	$34^\circ C$
Egg	3 days	2 days
Larva	20 days	15 days
Pupa	4 days	3 days
Reproductive maturation	5 days	4 days
Total time from egg to egg	32 days	24 days

Table 1.2. Developmental rates of *T. castaneum*

The eggs are white, microscopic and often have bits of flour stuck to their surface. The slender larvae are creamy yellow to light brown in color. They have two dark pointed projections on the last body segment. The young larva is yellowish white and measures 1mm in length. As it matures, it turns reddish yellow, becomes hairy and measures over 6mm in length. Its head, appendages and the last abdominal segment are darker. The adult is a small reddish-brown beetle, measuring about 3.5mm in length and 1.2mm in width. Its antennae are bent and bear a distinct club formed by the three enlarged terminal joints. The last antennal segment is transversely rounded. It was commonly found in wild state in rotting wood and in loose bark of trees in India. This insect is now widely distributed all over the world mainly through commerce.

1.2.1.1 Systemic position:

Kingdom: Animalia Phylum: Arthopoda Class: Insecta Order: Coleoptera Family: Tenebrionidae Genus: *Tribolium* Species: *T. castaneum* (Herbst)

Plate 1.12: *T. castaneum*

The red flour beetle may be present in large numbers in infested grain, but are unable to attack sound or undamaged grain (Walter, 1990). Both the larva and the adults cause damage. They are found in great numbers on infested materials and caused serious losses and considerable damage to flour and grains that have previously been attacked by other pests. Much of the damage done by *T. castaneum* is directly to kernels (germplasm and endoplasm). In case of severe infestation flour or other materials invaded may have a characteristics pungent odor as a result of the gaseous secretion exuded by the beetle. Such flour has an exceedingly low viscosity and its elasticity is markedly affected which may cause gastric disturbance if used as food. In severe infestation, the flour turn grayish and moldy and has a pungent, disagreeable odor making it unfit for human consumption (Good, 1936). Infested material will show many elongate reddish brown beetles, about 1/7 inch long crawling over the material when it is disturbed and brownish white (somewhat flattened) sixlegged larval bedding on the inside of the grain kernels and crawling over the infested seeds. They are generally known among millers as "bran bugs". *T. castaneum* contaminates more than they consume.

According to Khan (1981), this contamination results from, the presence of living or dead insects or insect parts; cast exuvae, eggshell and pupal cases; fecal and persistent odor; and webbing of food.

Tribolium species are major pests of stored grains and grain products in the tropics. Control of these insects relies heavily on the use of synthetic insecticides and fumigants, which has led to problems such as disturbances of the environment, increasing costs of application, pest resurgence, pest resistance to pesticides and lethal effects on non-target organisms in addition to direct toxicity to users (Jembere *et al.*, 1995). Thus, repellents, fumigants, feeding deterrents and insecticides of natural origin are rational alternatives to synthetic insecticides.

1.2.2. Brine shrimp (*Artemia salina***) nauplii**

Brine shrimp lethality bioassay is a recent development in the bioassay for the bioactive compounds, which indicates cytotoxicity, as well as, a wide range of pharmacological activities (e.g. anticancer, antiviral, pesticidal, anti-AIDS, etc.) of the compounds. Bioactive compounds are almost always toxic in high doses. Pharmacology is simply toxicology at a lower dose or toxicology is simply pharmacology at a higher dose. Brine shrimp lethality bioassay is a bench top bioassay method for evaluating anticancer, anti-microbial and pharmacological activities of natural products. Natural product extracts, fractions or pure compounds can be tested for their bioactivity by this method. Here *in vivo* lethality of a simple monitor for screening a fractionation in the discovery of new bioactive natural products.

The Brine shrimp belongs to a genus of very primordial crustacean (crawfish crayfish) the *Anostraca* (Fairy shrimp). Crawfish of this genus just have a divided exoskeleton made of Chitin enhanced protein, no usual crust of chitin (escutcheon) as other crawfish have. There are many species within the genus of *Anostraca*, but the *Artemia salina* is very nice to grow, since the rate of successful hatches is very high. To carry on toxicity tests of certain materials these nauplii are very easy to grow from its marketed cysts and to set experiments thereby.

1.2.2.1. Systemic position:

Kingdom: Animalia Phylum: Arthopoda Class: Branchiopoda Order: Anostraca Family: Artemiidae Genus: *Artemia*

Species: *A. salina* L. **Plate 1.13:** *A. salina* nauplii

1.2.3. Mosquito larvae (*Culex quinquefasciatus***)**

There are over 2500 different species of mosquitoes throughout the world. There are four common groups of mosquitoes living in the Bay Area. They are *Aedes*, *Anopheles*, *Culex*, and *Culiseta* (McCafferty, 1983). Mosquitoes are very important to serve as vectors of important diseases, such as West Nile virus, dengue, filariasis, Japanese encephalitis, St. Louis encephalitis and avian malaria.

1.2.3.1. Systemic position:

Kingdom: Animalia Phylum: Arthopoda Class: Insecta Order: Diptera Family: Culicidae Genus: *Culex* Species: *C. quinquefasciatus* Say

Plate 1.14: *C. quinquefasciatus* female

All mosquitoes must have water in which to complete their life cycle. The type of water in which the mosquito larvae is found can be an aid to the identification of which species it may be. Also, the adult mosquitoes show a very distinct preference for the types of sources in which to lay their eggs. They lay their eggs in such places such as tree holes that periodically hold water, tide water pools in salt marshes, sewage effluent ponds, irrigated pastures, rain water ponds, etc. Each species therefore has unique environmental requirements for the maintenance of its life cycle.

The feeding habits of mosquitoes are quite unique in that it is only the adult females that bite man and other animals. The male mosquitoes feed only on plant juices. Some female mosquitoes prefer to feed on only one type of animal or they can feed on a variety of animals. Female mosquitoes feed on man, domesticated animals, such as cattle, horses, goats, etc; all types of birds including chickens; all types of wild animals including deer, rabbits; and they also feed on snakes, lizards, frogs, and toads. Most female mosquitoes have to feed on an animal and get a sufficient blood meal before she can develop eggs. If they do not get this blood meal, then they will die without laying viable eggs. However, some species of mosquitoes have developed the means to lay viable eggs without getting a blood meal.

The flight habits of mosquitoes depend on the species. Most domestic species remain fairly close to their point of origin while some species known for their migration habits are often an annoyance far from their breeding place. The flight range for females is usually longer than that of males. Many times wind is a factor in the dispersal or migration of mosquitoes. Most mosquitoes stay within a mile or two of their source. However, some have been recorded as far as 75 miles from their breeding source (McCafferty, 1983).

Mosquito have complete metamorphosis in their life cycle. The length of life of the adult mosquito usually depends on several factors: temperature, humidity, sex of the mosquito and time of year. Most males live a very short time, about a week; and females live about a month depending on the above factors. The mosquito goes through four separate and distinct stages of its life cycle and they are as follows: Egg, Larva, pupa, and adult. Each of these stages can be easily recognized by their special appearance.

Eggs are laid one at a time and they float on the surface of the water. In the case of *Culex* and *Culiseta* species, the eggs are stuck together in rafts of a hundred or more eggs. *Culex*, *Culiseta*, laid their eggs on water. Most eggs hatch into larvae within 48 hours. Mosquito larvae are commonly referred to as "Wrigglers", these newly hatched insects can be seen wriggling up and down from the surface of the water. The larva lives in the water and come to the surface to breathe. They shed their skin four times growing larger after each molting. Most larvae have siphon tubes for breathing and hang from the water surface. The larva feed on micro-organisms and organic matter in the water. On the fourth molt the larva changes into a pupa. The pupal stage is a resting, non-feeding stage. This is the time the mosquito turns into an adult. It takes about two days before the adult is fully developed. When development is complete, the pupal skin splits and the mosquito emerges as an adult. The newly emerged adult rests on the surface of the water for a short time to allow itself to dry and all its parts to harden. Also, the wings have to spread out and dry properly before it can fly. The adult mosquito can measure from 4–10mm (McCafferty, 1983) and morphologically has the three body parts common to insects: head, thorax, and abdomen.

Plate 1.15: Mosquito egg raft (left) and larvae (right) in natural condition

Plate 1.16: Life cycle of mosquito

The egg, larvae and pupae stages depend on temperature and species characteristics as to how long it takes for development. For instance, *Culex tarsalis* might go through its life cycle in 14 days at 70°F and take only 10 days at 80°F (McCafferty, 1983). Also, some species have naturally adapted to go through their entire life cycle in as little as four days or as long as one month.

1.2.4. Eggplant aphids (*Aphis gossypii***)**

The aphids constitute a large group of small, soft-bodied insects that are frequently found in large number sucking the sap from the stem or leaves of plants. Melon and green peach aphids attack a number of crops and are vectors of many viruses attacking sweet-potato, brinjal and other crops.

1.2.4.1. Systemic position:

Kingdom: Animalia Phylum: Arthropoda Class: Insecta Order: Homoptera Family: Aphididae Genus: *Aphis* Species: *A. gossypii* Glover

Plate 1.17: *Aphis gossypii* Glover

A. gossypii found in Worldwide. Their host range apart from sweet-potato. This species also damages citrus, cocoa, coffee, cotton, cucurbits, eggplant, okra, pepper, potato and also ornamentals like Hibiscus (Amalin *et al.,* 1993; Ames *et al.,* 1996).

These insects can be recognized by its pear-like shape, a pair of cornicles at the posterior end of the abdomen and fairly long antennae; winged forms can usually be recognized by the venation and relative size of the front and hind wings. The cornicles of aphids are tube-like structures arising from the dorsal side of the fifth or sixth abdominal segment. The nymphs are green to brown and moult four times before reaching the adult stage. The nymphs look like the wingless adults except for their small sizes and softer body. The adults are small to medium sized, 1.2-2.5mm long. They are usually green with darker thorax. The antennae are two-thirds as long as the body.

The cornicles are clavate and fairly long. The face when viewed dorsally has a characteristic shape. This species generally produces little honeydew. The life cycle of aphids is rather unusual and complex.

Plate 1.18: Life cycle of Aphid

The females reproduce parthenogenetically. Several generations may be produced in short period of time. The first generation usually consists of wingless individuals; however, when a colony becomes too crowded, winged individuals appear.

The winged forms migrate to a different host plant and begin new colonies, a generation consisting of both males and females is produced, and the reproductive process continues. Aphids secrete honeydew which is emitted from the anus; the honeydew consists mainly of excess sap ingested by the insect, to which are added excess sugars and waste materials. Aphids usually attack the growing shoots and expanding leaves. They feed on the lower surface of the leaves and injure the plants by sucking the sap. The leaves become deformed as they expand. They may curl down at the edges, and become wrinkled or puckered. Feeding on expanded leaves (more common with green peach aphid) may result in pale stippled areas of feeding damage between the veins. During heavy infestation, the vigour of the plant is greatly reduced, stunting growth of the plants. Leaves of such stunted plants are pale and may have yellow interveinal areas (Vasquez *et al.,* 1990).

Eggplant aphid spends part of the winter on weed hosts and in gardens on cold tolerant plants. During warm periods, they continue feeding until cold weather inactivates them. In spring, winged females fly to suitable host plants and give birth to living young. Each female produces an average of 84 nymphs. Under favorable conditions, a nymph will mature in about 5 days and begin producing its own progeny. Most nymphs develop into wingless female adults. However, when crowding occurs or food becomes scarce, winged adults develop and fly to new host plants. Reproduction continues through the winter as in the summer but at a much slower rate. Many overlapping generation are produced each year.

1.2.5. Agents for antibacterial tests

To carry on screening of pesticidal properties of the extracts of all the test samples eight pathogenic bacteria were selected for the test, five of which were Gram negative and the remaining were Gram positive. These organisms of pure culture were collected from the Department of Biochemistry, University of Rajshahi -6205, Bangladesh.

It is very important to determine whether the crude extracts are active against various types of test organisms or not and thus a preliminary antibacterial screening of the crude extract was very much necessary. Therefore, screening was done against various test pathogenic bacteria by disc diffusion assay (Bauer *et al.*, 1966; Barry *et al.*, 1976) method.

Table 1.3. List of the test pathogenic bacteria

1.3. Hypothesis of this research work

For the control of vectors active ingredients of plants could be used through furnishing their biological activity (against different vectors agents responsible for plant, human and animal diseases); isolation, purification and characterization of the detected promising bioactive compound(s); further bioassay with the purified compound(s) (if a sufficient amount is received) will be done if the laboratory support permit.

1.4. Aim of this study

To promote environmentally safe pesticides to replace chemical ones.

1.5. Objectives of this work

Quite a good number of plants have been identified and utilized for insecticidal and medicinal purpose till to date. But it is true that a large number of plants have still remain untouched or less investigated from which significant results can be obtained to control the pest of crops and disease problems of human beings. In this proposition *Evolvulus nummularius* (wp), *Lantana camara* (ap & r), *Mentha piperita* (wp), *Mimosa pudica* (wp), *Parthenium hysterophorus* (wp), *Phyllanthus niruri* (wp), *Polygonum hydropiper* (wp), *Pouzolzia zeylanica* (wp), *Synedrella nodiflora* (wp), *Zingiber zerumbet* (ap & rh) were attemted since some of these plants have been studied phytochemically, but only a few studies have been done on its medicinal properties, and a very few works have been attempted on its uses for the control of vectors.

Objectives:

- 1. A thorough screening program will be attempted to trace presence of bioactive potentials (insecticidal, larvicidal, insect repellency, etc.) with the study materials by-
	- **1999** using mosquito (human disease vector) larvae for assessing larvicidal activity of the extracts through establishing LC_{50} values;
	- **3** using eggplant aphid (plant disease vector) as a test agent to evaluate the insect repellent and insecticidal activity of the promising extracts through establishing LD₅₀ values;
	- **W** using the stored product pest and insect vector, *Tribolium castaneum* to evaluate efficacy of the promising extracts through dose-mortality tests by establishing LD_{50} values through surface film assay and also to evaluate the repellent activity;
	- using *Artemia salina* (a shrimp disease vector)*,* the recognized test agent for cytotoxic effect to evaluate cytotoxicity of the extracts by establishing LC_{50} values;

(For the evaluation of further bioactive potentials of the test plants the following experiment has also been done.)

- **图 Using pathogenic bacteria to know the presence of antibacterial properties in** the study plants.
- 2. To summarize the potentials of the test materials through searching literature and web information in comparison to the results of the investigation to be carried out on their possible use in the vector control strategy.
- 3. Isolation, purification and characterization of the promising bioactive compound(s) from the test plants for their possible use in the vector control sector.
	- **W** To comment on the future perspectives of the test plants depending on the achieved results.

MATERIALS AND METHODS

Plants are the most suitable source in the field of pesticide technology while some plants in different parts of the world are considered toxic and some are used in the traditional medicine. A literature search on the title plant offered some essential openings that these species bears repellent and toxicological properties which is subjected to go thorough screening to develop natural non-hazardous biodegradable pesticides. The approach adopted to obtain an exploitable pure plant constituent involves interdisciplinary work in Botany, Zoology, Pharmacognosy, Pharmacology, Phytochemistry, Chemistry and Toxicology as described by Hostettmann *et al.* (1995).

2.1. Selection and collection of plant materials

Evolvulus nummularius (wp), *Lantana camara* (ap and r)*, Mentha piperita* (wp)*, Mimosa pudica* (wp)*, Parthenium hysterophorus* (wp)*, Phyllanthus niruri* (wp)*, Polygonum hydropiper* (wp)*, Pouzolzia zeylanica* (wp)*, Synedrella nodiflora* (wp)*,* and *Zingiber zerumbet* (ap and rh) have been collected for the presence of toxic, as well as, bio-active constituents since the plants are well known as medicinal plants and also considered to contain toxic constituents. In case of very small plants, such as herbs, grass, etc. normally the whole plant is subjected for extraction, because the distribution of constituents, generally in different parts of such small plants not varies too much. But, it varies in case of a large timber plants. The distribution of compounds in different parts of higher plants is obviously different. The presence of constituents in the heart-wood may disappear in the leaves; similarly constituents in the roots may not be the same that present there in the fruits.

2.1.1. Preparation of plant materials for extraction

The fresh materials of all test plants excepting *Z. zerumbet* were collected from the RU campus. *Z. zerumbet* was collected from Mirzapur Rajshahi. Whole plants of *E. nummularius*, *M. piperita*, *Mi. pudica*, *P. hysterophorus*, *Ph. niruri*, *Po. hydropiper*, *Pz. zeylanica*, *S. nodiflora* were collected together with roots. In case of *L. camara* aerial part and roots, and in case of *Z. zerumbet* aerial part and rhizome was separated. Excess soils from collected parts were removed, without washing and then cut into small pieces using a knife and spread out to dry without heaping the material together. This was done under shade of the sun (beware of rain) or in well-ventilated room. After drying well the plant materials were powdered in a grinder machine avoiding excess heat during grinding.

Po. hydropiper (whole plant) *Pz. zeylanica* (whole plant)

Plate 2.1: Collection and processing of different parts of test plants after grinding

2.1.1.1. Chemical extraction of the collected materials

There are basically two methods for extracting compounds from plant materials. Which one to choose, depends on whether the aim is to extract the more polar compounds (especially glycosides) that are present in the cell vacuole, or to obtain the less polar aglycones present on the surface of the plant, in aerial parts heartwood or roots. In the present study three solvents were selected to extract. The three solvents were Petroleum ether (PetE), Chloroform (CHCl₃) and Methanol (CH₃OH).

The dried ground plant materials were extracted with sufficient amount of solvents (200g x 600ml x 2 times). The extraction was firstly done by PetE and then with CHCl₃ and CH3OH successively. Separate extracts have been collected by the cool method after 72h of plunging for each of the materials. Extracts, thus obtained were filtered and concentrated on a rotary evaporator at 40˚C and only as residues (extracts) left were collected in a glass vial and kept in a cool place after labeling.

Plate 2.2: Samples plunged in different solvents

Plate 2.3: Plunged samples in conical flasks on a shaker

Plate 2.4: Filtration of extracts **Plate 2.5:** Storage of different extracts in glass vials after labeling

Extraction procedure (using PetE, CHCl³ and CH3OH)

Fig. 2.1: Pathway of extraction before screening

Fig. 2.2: Collection of extracts from the whole plants of *E. nummularius*, *M. piperita, Mi. pudica, P. hysterophorus, Ph. niruri, Po. hydropiper, Pz. zeylanica, S. nodiflora* by different solvents

Fig. 2.3: Collection of extracts of *L. camara* (aerial part and root) by different solvents

Fig. 2.4: Collection of extracts of *Z. zerumbet* (aerial part and rhizome) by different solvents

2.2. Selection of test organisms

To carry on tests for vector control properties of the extractives of the selected plants the eggplant aphid, *Aphis gossypii* (Homoptera: Aphididae); larvae of the mosquito, *Culex quinquefasciatus* (Diptera: Culicidae); red flour beetle, *Tribolium castaneum* (Hbst.) (Coleoptera: Tenebrionidae); and the brine shrimp, *Artemia salina* nauplii were selected as the test organisms. The life histories made these agents popular choice for the evaluation of biological activitiy of certain test materials. They are also easy to collect, hatch out/ culture in large numbers and require no sophisticated equipment for maintenance. Except these test vectors, pathogenic bacteria were also selected to carry out further efficiency tests of the extractives.

2.3. Collection of test organisms

Source of test insects - *A. gossypii* were collected from cultivated field situated in the Fourth Science Building, University of Rajshahi. Eggs (rafts) of *C. quinquefasciatus* were collected from different drains of University of Rajshahi Campus.*T. castaneum,* used in the present investigation were taken from the stock cultures of the Crop Protection and Toxicology Laboratory, Department of Zoology, University of Rajshahi, Bangladesh; and reared as subcultures to be used in the experimentation. Cysts of *A. salina* were collected from (Different aquarium shops) Dhanmondi, Dhaka, Bangladesh. Bacterial strains were collected from the Department of Botany, University of Rajshahi, Bangladesh.

2.3.1. Eggplant aphids, *Aphis gossypii*

Plate 2.6: Eggplants in the net house **Plate 2.7:** Culture of aphid

2.3.1.1. Culture of aphids

Aphids are very soft and tiny creatures. They are highly plant consuming insects. At first some mature aphids were collected from affected plants and then released them on the new fresh eggplants for further production. Aphids are highly reproducing insects. They multiply in a good number within a very short time.

2.3.1.2. Collection of aphids

Aphids were collected from the eggplant leaves with a fine camel hair-brush in a Petridish and used in the experiments.

2.3.2. *Artemia salina* **L.**

2.3.2.1. Culture of *A. salina*

As the *A. salina* is marine crustacean, this is not easy to culture like *T. castaneum* under lab conditions*.* But, they can be reared in a short edition by putting cysts in a beaker of sea-water until the nauplii are hatched out. To carry on toxicity tests of certain materials these nauplii are used.

2.3.2.2. Preparation of environment

Since the lethality test involves the use of brine shrimp nauplii, the nauplii should be grown in seawater. Seawater contains 3.8% of NaCl. Accordingly 3.8% sodium chloride solution was made by dissolving sodium chloride (38g) in distilled water (1000ml) and was filtered off. Brine water was taken in a small tank and *A. salina* cysts (1.5g/L) were added to one side of the perforated divided tank with constant oxygen supply. Constant temperature (37°C) and sufficient light were maintained to give the sufficient aeration.

2.3.2.3. Collection of newly hatched nauplii

After 24h, nauplii were collected and used in the experiments.

2.3.3. Mosquito larvae, *Culex quinquefasciatus*

2.3.3.1. Preparation of environment and culture of mosquito larvae

Mosquito eggs are hatched in stagnant water. They are collected from damp drains with special collecting spoon. Collected mosquito egg-rafts are placed into a new beaker containing pond water and kept it in a dark place inside the lab for hatching.

2.3.3.2 Collection of newly hatched larvae

After 24h, hatched larvae were collected from the hatching tank and used in the experiment.

Plate 2.8: Mosquito eggs-rafts **Plate 2.9:** Culture of *T. castaneum*

2.3.4. *Tribolium castaneum* **Hbst.**

2.3.4.1. Culture of test insect *T. castaneum*

In plastic containers (1200ml) mass cultures were maintained and sub-cultures in beakers (1000ml) with the food medium. The beakers were kept in an incubator at 30° C \pm 0.5°C without light and humidity control. Each container and beaker contained 250g and 150g of food respectively. About 200 adults in each container and 100 adults in each beaker were introduced. The cultures were checked in regular intervals and eggs and larvae were separated to grow up properly. A crumpled filter paper was placed inside each container and beaker for easy movement of the beetles. The containers and beakers were covered with pieces of muslin cloth tightly fixed with the help of rubber bands to avoid possible escape of the beetles.

2.3.4.2. Preparation of food medium

The whole-wheat flour was used as the food medium for the insect species. The flour was sterilized at 60˚C for 24 hours in an oven. A standard mixture of whole wheat flour with powdered dry yeast in a ratio of 19:1 (Park and Frank, 1948; Park., 1962; Zyromska-Rudzka, 1966; Khalequzzaman *et. al*., 1994) was used as food medium throughout the experimental period. Both the flour and the powdered dry yeast were sterilized at 60˚C for six hours in an oven. Food was not used until at least 15 days after sterilization to allow its moisture content to equilibrate with the environment (Khan, 1981).

2.3.4.3. Collection of eggs

About 500 beetles were placed in a 500ml beaker containing food medium. The beaker was covered with a piece of cloth and kept in an incubator at 30° C \pm 5[°]C. In
regular interval the eggs were collected by sieving the food medium by two sieves of 500 and 250mesh separating the adults and eggs respectively following the methods of Khan and Selman (1981). Eggs were then transferred to Petri dish (90mm diam.) and incubated at the same temperature.

2.3.4.4. Collection of newly hatched larvae

After 3-5 days, larvae hatched out in described conditions. Newly hatched larvae were then collected with a fine pointed camel hair brush and then shifted to the fresh food medium for culture. The larvae were yellowish white in color and long cylindrical in shape. It appears 1mm in length after hatching and become 6-7mm at maturation.

2.3.4.5. Collection of mature larvae

Most larvae had six instars as reported by Good (1936). According to Good, the larval instars were determined by counting the number of exuviae (larval skin) deposited in the food medium. Two days old larvae was considered as first instar larva while second, third, fourth, and fifth instar larvae were considered on fourth, seventh, tenth and thirteenth day from hatching respectively. Depending on these days according to larval instar sixteen days old larva have been considered as a mature larva. Larval cultures were maintained in an incubator in the same procedure at 30° C \pm 5°C without light and humidity control. The food medium was replaced by three days interval to a fresh one to avoid conditioning of the larvae (Park, 1934).

2.3.4.6. Collection of adults

A huge number of beetles were thus reared to get a regular supply of the newly formed adults. When sufficient adults produced in the sub-cultures, they were collected from the food medium. For this purpose some pieces of filter paper were kept on the food inside the beaker. Adults crawled upon the paper and then the paper was taken out with the help of forceps. Beetles were then collected in a small beaker (100ml) with the help of a fine camel-hair brush.

2.4. Bioassays for activity of the collected extracts

Crucial to any investigation of plants with biological activities is the availability of suitable bioassays for monitoring the required effects. In order to cope with the number of extracts a high sample throughput is necessary. The test systems should ideally be simple, rapid, reproducible and less-expensive. If active principles are only present at low concentration in the crude extract then bioassay is to be high enough sensitive for

their detection. Another factor of special relevance to plant extracts is the solubility of the sample. Finding a suitable system can pose problems.

For the selection of bioassays to employ in research on plant constituents, the first step is to choose suitable target organisms. The complexity of the bioassay has to be designed as a function of the facilities and resources available. A list of bioassays taken in this investigation is shown in Table 2.1.

2.4.1. Bioassay with residual film/surface film experiments

2.4.1.1. Experiments for surface film test by *T. castaneum* **adults**

Each of the extracts was diluted with the solvent in which it was extracted and the actual amount of extracted matter in a dose was recorded. The application of dose was carried out by residual film method (Busvine, 1971). For each dose 1ml of mixture was dropped on a Petri dish (50mm) in such a way that it made a uniform film over the Petri dish. Then the Petri dishes were air-dried leaving the extract on it. The actual extract present in 1ml mixture was calculated and dividing the value by the area of the Petri dish the dose per square centimeter was calculated. After drying 10 red flour beetles (3-10 days old) were released in each Petri dish with 3 replicates. A control batch was also maintained with the same number of insects after preparing the Petri dish by applying and evaporating the solvent. The treated beetles were placed in the incubator at the same temperature as reared in stock cultures and the mortality of the beetles was counted after 30min and every 12h up to 48h of exposure.

This is also one basic application method for doses of toxic substances to any insect population. The test material has been dissolved in an organic solvent with a certain concentration to apply to a Petri dish of known surface area. After application the volatile solvent evaporates out immediately simply with the atmospheric temperature. Thus the ingredient goes to make film on the surface of the Petri dish. Released

insects within this captivity might have contact with the substance distributed evenly on the floor. However, being covered with the upper lid of the Petri dish there could have a captive environment with the extract distributed even in the air inside and may cause mortality by suffocation. Mortality due to suffocation may cause promptly if there is any volatile bioactive principles in the test material.

2.4.1.2. Preparation of doses with the crude extracts for the surface film test (to be used against *T. castaneum***)**

In this investigation dose-mortality efficiency was evaluated through surface film experiment with series of doses applied on *T. castaneum* adults. All the three extracts (PetE, CHCl3, and CH3OH) of the test plant were applied against *T. castaneum* adults. For each samples, a 'pilot' test was done before final experimentation. A 50mg sample extract was weighed and taken in a small glass vial, and then 1ml of the same chemical (1ml PetE) was added to dissolve it initially to prepare 2.547mg/cm**²** the dose. This process was also maintained during final experimentation. Separate vials were taken for each of the doses. All experiments were done in three replicates. The final doses for surface film application were obtained as follows:

- For *E. nummularius* (wp) in PetE: 1.529, 1.274, 1.019, 0.764, 0.510 and 0.255mg/cm²; CHCl₃: 4.076, 3.567, 3.057, 2.548, 2.038 and 1.529mg/cm²; CH₃OH: 3.567, 3.057, 2.548, 2.038 and 1.529mg/cm²;
- \triangleright For *L. camara* (ap) in PetE: 4.076, 3.567, 3.057, 2.548 and 2.038mg/cm²; CHCl₃: 2.548, 2.038 ,1.529, 1.019 and 0.510mg/cm² ; CH3OH: 2.038 ,1.529, 1.019 , 0.510 and 0.255mg/cm²;
- For *L. camara* (r) in PetE: 3.567, 3.057, 2.548, 2.038 and 1.529mg/cm² ;
- \triangleright For *M. piperita* (wp) in PetE: 1.529, 1.019, 0.510, 0.255 and 0.127mg/cm²; CHCl₃: 2.548, 2.038, 1.529, 1.019 and 0.510mg/cm²; CH₃OH: 1.529, 1.019, 0.510, 0.255 and 0.127 mg/cm²;
- \triangleright For *P. hysterophorus* (wp) in PetE: 3.567, 3.057, 2.548, 2.038 and 1.529mg/cm²; CHCl₃: 4.076, 3.567, 3.057, 2.548 and 2.038mg/cm²; CH₃OH: 2.548, 2.038, 1.529, 1.019 and 0.510 mg/cm²;
- > For *Ph. niruri* (wp) in PetE: 2.037, 1.528, 1.019, 0.509, and 0.255mg/cm²; CHCl₃: 3.565, 3.056, 2.547, 2.037, and 1.528mg/cm²; CH₃OH: 3.565, 3.056, 2.547, 2.037, 1.528, and 1.019mg/cm²;
- For *Po. hydropiper* (wp) in PetE: 2.293, 2.038, 1.783, 1.529, 1.274 and 1.019mg/cm²; CHCl₃: 4.586, 4.076, 3.567, 3.057 and 2.548mg/cm²; CH₃OH: 1.529, 1.019, 0.510, 0.255 and 0.127mg/cm²;
- \triangleright For *Pz. zeylanica* (wp) in PetE: 3.056, 2.547, 2.037, 1.528, and 1.019 mg/cm²; CHCl₃: 3.056, 2.547, 2.037, 1.528, and 1.019mg/cm²;
- > For S. nodiflora (wp) in PetE: 1.529, 1.274, 1.019 and 0.764mg/cm²; CH₃OH: 3.567, 3.057, 2.548, 2.038 and 1.529mg/cm²;
- > For *Z. zerumbet* (ap) in PetE: 2.548, 2.038, 1.529, 1.019 and 0.510mg/cm²; CHCl₃: 3.567, 3.057, 2.548, 2.038 and 1.529mg/cm²; CH₃OH: 2.548, 2.038, 1.529, 1.019 and 0.510mg/cm²;
- ► For *Z. zerumbet* (rh) in PetE: 2.038 ,1.529, 1.019, 0.510 and 0.255mg/cm²; CHCl₃: 3.567, 3.057, 2.548, 2.038 and 1.529mg/cm²; CH₃OH: 2.038, 1.529, 1.019, 0.510 and 0.255 mg/cm²

2.4.1.3. Application of doses in the surface film test

To conduct surface film activity test 50mm Petri dishes were taken for all doses and their replicates, 1ml of each of the doses were poured into the lower part of the Petri dishes and allowed them to dry out. Being volatile the solvent was evaporated out within a few minutes. Ten insects were released in each of the treated Petri dish. A control experiment by applying the only solvent into the Petri dish was also set at the same time under the same condition.

(A) Pilot test (B) Final experiment

Plate 2.10: Bioassay with plant extract on *T. castaneum* adults by surface film methods

2.4.1.4. Observation of mortality in surface film tests

After completing all the arrangements of the experiment treated Petri dishes were placed in a secure place at room temperature. The whole experiment was observed from time to time and mortality was observed by after 30min and every 12h and the data was recorded up to 48h. A simple microscope was used to check each and every beetle by tracing natural movement of its organs. In some cases hot needle was taken closer to the bodies (without movement) to confirm death. Attention was also paid to recovery of the insects (if occurred).

2.4.2. Experiments for surface film test by *A. gossypii*

Fresh eggplant leaves were collected from net house and placed on a Petri dish. Stalks of each of the leaves were wetted with water and cotton to keep fresh. With the help of a permanent marker a round mark (3.6cm diam.) drawn on the leaves surrounding the open edge of the 10ml plastic cup (3.6cm diam. normally used for measuring liquid medicines) and were cut into round shapes by a pair of scissors keeping the stalks attached. Extracts were applied on both the sides of the round shaped pieces of leaves with the help of a 1ml syringe. Three replicates for each of the doses were maintained and allowed to dry out as exposed in the air for 30 to 45min. Then ten insects were released in the middle of each of the leaf circles.

2.4.2.1. Preparation of doses with the crude extracts for the surface film test (to be used against *A. gossypii***)**

In this investigation dose-mortality efficiency was evaluated through surface film experiment with series of doses applied on *A. gossypii*. All the three extracts (PetE, $CHCl₃$, and $CH₃OH$) of the test plant were applied. For each samples, a 'pilot' test was set before final experimentation. About 4mg sample extract was weighed and taken in a small glass vial. Dimethyl sulfoxide (DMSO) was used to make the extracts hydrophilic and 0.1ml of distilled water was added to dissolve initially for preparation of a 0.196mg/cm**²** dose. This process was also maintained in the final experiment. Separate vials were taken for each of the doses, while the doses were maintained in three replicates. The final doses for surface film application were obtained as follows:

 \triangleright For *E. nummularius* (wp) in PetE: 0.196, 0.147, 0.098, 0.049 and 0.025mg/cm²; CHCI₃: 0.196, 0.147, 0.098, 0.049 and 0.025mg/cm²; CH₃OH: 0.196, 0.147, 0.098, 0.049 and 0.025mg/cm $^2_\mathrm{p}$

- \triangleright For *L. camara* (ap) in PetE: 0.196, 0.147, 0.098, 0.049 and 0.025mg/cm²; CHCl₃: 0.196, 0.147, 0.098, 0.049 and 0.025mg/cm²; CH₃OH: 0.246, 0.196, 0.147, 0.098 and 0.049mg/cm $^2_\mathrm{p}$
- \triangleright For *L. camara* (r) in PetE: 0.196, 0.147, 0.098, 0.049 and 0.025mg/cm²; CHCl₃: 0.196, 0.147, 0.098, 0.049 and 0.025mg/cm²; CH₃OH: 0.246, 0.196, 0.147, 0.098 and 0.049mg/cm 2 ;
- \triangleright For *M. piperita* (wp) in PetE: 0.196, 0.147, 0.098, 0.049 and 0.025mg/cm²; CHCl₃: 0.246, 0.196, 0.147, 0.098 and 0.049mg/cm²; CH₃OH: 0.246, 0.196, 0.147, 0.098 and 0.049mg/cm 2 ;
- > For *Mi. pudica* (wp) in PetE: 0.246, 0.196, 0.147, 0.098 and 0.049mg/cm²; CHCl₃: 0.196, 0.147, 0.098, 0.049 and 0.025mg/cm²; CH₃OH: 0.196, 0.147, 0.098, 0.049 and 0.025mg/cm 2 ;
- For *P. hysterophorus* (wp) in PetE: 0.196, 0.147, 0.098, 0.049 and 0.025mg/cm² ; CHCl₃: 0.196, 0.147, 0.098, 0.049 and 0.025mg/cm²; CH₃OH: 0.246, 0.196, 0.147, 0.098 and 0.049mg/cm $^2_\mathrm{p}$
- \triangleright For *Ph. niruri* (wp) in PetE: 0.196, 0.147, 0.098, 0.049 and 0.025mg/cm²; CHCl₃: 0.393, 0.344, 0.295, 0.246 and 0.196mg/cm²; CH₃OH: 0.246, 0.196, 0.147, 0.098 and 0.049mg/cm 2 ;
- \triangleright For *Po. hydropiper* (wp) in PetE: 0.196, 0.147, 0.098, 0.049 and 0.025 mg/cm²; CHCl₃: 0.246, 0.196, 0.147 and 0.098mg/cm²; CH₃OH: 0.246, 0.196, 0.147, 0.098 and 0.049mg/cm 2 ;
- \triangleright For *Pz. zeylanica* (wp) in PetE: 0.196, 0.147, 0.098, 0.049 and 0.025 mg/cm²; CHCI₃: 0.246, 0.196, 0.147, 0.098, 0.049 and 0.025mg/cm²; CH₃OH: 0.295, 0.246, 0.196, 0.147 and 0.098mg/cm 2 ;
- > For S. nodiflora (wp) in PetE: 0.196, 0.147, 0.098, 0.049 and 0.025mg/cm²; CHCl₃: 0.196, 0.147, 0.098, 0.049 and 0.025mg/cm²; CH₃OH: 0.246, 0.196, 0.147, 0.098 and 0.049mg/cm 2 ;
- > For *Z. zerumbet* (ap) in PetE: 0.147, 0.098, 0.049, 0.025 and 0.012mg/cm²; CHCl₃: 0.196, 0.147, 0.098, 0.049 and 0.025mg/cm²; CH₃OH: 0.246, 0.196, 0.147, 0.098 and 0.049mg/cm 2 ;

> For *Z. zerumbet* (rh) in PetE: 0.147, 0.098, 0.049, 0.025 and 0.012mg/cm²; CHCl₃: 0.196, 0.147, 0.098, 0.049 and 0.025mg/cm²; CH₃OH: 0.246, 0.196, 0.147, 0.098 and 0.049 mg/cm²

2.4.2.2. Application of doses in the surface film test against eggplant aphids

An amount of 0.1ml of each of the doses were poured onto both sides of the leaves and allowed them to dry out. Distilled water was considered as the solvent and DMSO (Dimethyl sulfoxide) was used to make the extract hydrophilic. The water was evaporated out within 45 minutes. Ten adult aphids were released on either side of each of the treated round shaped leaves with a fine camel hair-brush. A control experiment by applying the only DMSO in water applied onto either side of the leaf circle was also set at the same time under the same condition as a control.

Plate 2.11: Bioassay through dose-mortality assay against eggplant aphids

2.4.2.3. Observation of mortality in the surface film tests in case of eggplant aphids

After completing all the arrangements the treated leaves were placed in a 50mm Petri dish where stalks of the leaf circles were covered with water soaked cotton to keep it fresh for a long time and placed on a Petri dish at room temperature. The whole experiment was observed from time to time and the mortality was observed every 3h and the data was recorded up to 24h. A magnifying glass was used to check each and every adult by tracing natural movement of their limbs.

2.4.2.4. Statistical analysis

The recorded mortality data of the dose-mortality assay done against *T. castaneum* adults and on *A. gossypii* were corrected by the Abbott's (1925) formula:

$$
P_r = \frac{P_o - P_c}{100 - P_c} \times 100
$$

Where,

 P_r = Corrected mortality (%)

 P_o = Observed mortality (%)

 P_c = Control mortality (%), sometimes called natural mortality (%).

Then mortality percentages were subjected to statistical analysis according to Finney (1947) and Busvine (1971) by using a 'computer software'. The dose-mortality relationship was expressed as a median lethal dose (LD_{50}) .

2.4.3. Experiment for repellent activity of the extracts

The technique of the repellency test used in this investigation was adopted from the method (No.3) of McDonald *et al*., (1970) with some modifications by Talukder and Howse (1993, 1994). No significant difference was detected between the repellency of the only solvent impregnated and untreated filter papers/areas in tests designed to check for any possible influence of different solvents. The average of the counts was converted to per cent repellency (PR) using the formula of Talukder and Howse (1993, 1995):

A general concentration for each of the plant extracts was selected as a stock dose for repellency test against adult beetles of *T. castaneum*, while other successive doses 0.314, 0.157, 0.079, 0.039 and 0.019mg/cm² were made by serial dilution. Similarly 0.393, 0.197, 0.098, 0.049 and 0.025mg/ $cm²$ were made doses for the application against *A. gossypii*.

2.4.3.1. Application of doses in the repellency test

2.4.3.1.1. Application of doses against *T. castaneum* **adults**

Half filter paper discs (Whatman No. 40, 9cm diam.) were prepared and selected doses of all the extracts separately applied onto each of the half-discs and allowed to dry out as exposed in the air for 10 minutes. Each treated half-disc was then attached lengthwise, edge-to-edge, to a control half-disc with adhesive tape and placed in a Petri dish (9cm diam.). The experiments were set in triplicates. Being volatile the solvent was evaporated out within a few minutes.

Plate 2.12: Photographs showing setting up of repellency test with the test extracts against *T. castaneum* adults by filter paper disc method

Then ten insects were released in the middle of each of the filter paper circles. The orientation of the same was changed in the replica to avoid the effects of any external directional stimulus affecting the distribution of the test insects. Each concentration was tested five times. Insects that settled on each non treated half of the filter paper discs were counted after 1h and then at hourly intervals up to 5h.

2.4.3.1.2. Application of doses against eggplant aphids

Fresh eggplant leaves were collected from net house and placed on a Petri dish. Stalks of each of the leaves were wetted with water soaked in cotton to keep the leaf fresh. With the help of a permanent marker a round mark (3.6cm diam.) was drawn on the leaves surrounding the open edge of the 10ml plastic cup (3.6cm diam., normally used for measuring liquid medicines) and were cut into a round shape by a pair of scissors keeping the stalks attached. The permanent marker was again used to demark two halves of the leaf circles. Extracts were applied on one half of the round shaped pieces of leaves with the help of a 1ml syringe. Three replicates for each of the doses were maintained and allowed to dry out as exposed in the air for 30 to 45min. Then ten insects were released in the middle of each of the circles and covered by the plastic cups. A small stone piece was used as load for each of the cup to prevent escaping of the insects from the restricted circle. Insects that settled on each of the non-treated half of the circles were counted after 1h and then at hourly intervals up to 5h.

Plate 2.13: Photographs of repellency test of the plant extracts against *A. gossypii*

2.4.3.2. Observation and analyses of repellency data

Repellency was observed for one-hour interval and up to five successive hours of exposure, just by counting the number of insects from the non-treated part of the filter paper spread on the floor of the 90mm Petri dish (for *T. castaneum*) and non-treated part of the restricted circle (36mm) on the eggplant leaves (for *A. gossypii*). The values in the recorded data were then calculated for percent repellency, which was again developed by arcsine transformation for the calculation of analysis of variance (ANOVA).

2.4.3.3. Statistical analysis

The values in the recorded data were calculated for percent repellency (PR).

$$
PR = (Nc - 5) \times 20
$$

Where, Nc is the average hourly observation of insects on the untreated half of the disc. Positive and negative values expressed for repellent and attractant activity respectively.

PR data were again developed by arcsine transformation for the calculation of ANOVA.

2.4.4. Lethality test against the brine shrimp nauplii

2.4.4.1. Experimental design for lethality test

Brine shrimp cysts were hatched in simulated seawater to get fresh nauplii for the experimentation. Test samples were prepared by the addition of calculated amount of DMSO (Dimethyl sulfoxide) for obtaining desired concentration of test sample. The nauplii were counted by visual inspection and were taken in test-tubes containing 5ml of simulated seawater. Then samples of different concentrations were added to the pre-marked test-tubes using a pipette. The test-tubes were left for 30h and then the dead nauplii were counted to find out cytotoxicity of the test material and compared the results with the control.

Test materials:

- *A. salina* (Brine shrimp) cysts;
- \triangleright Sea salt (Non-ionized NaCl);
- \triangleright Small tank/beaker to hatch the shrimp;
- \triangleright Pipette (1ml and 5ml);
- \triangleright Test tubes (20ml);
- \triangleright Magnifying glass.

2.4.4.2. Preparation of simulated seawater (brine water) and hatching of brine shrimp nauplii

Since the lethality test involves the culture of brine shrimp nauplii, the nauplii should be grown in the seawater. Seawater contains 3.8% of NaCl. Accordingly 3.8% sodium chloride solution was made by dissolving sodium chloride (38g) in normal pond water (1000ml) and was filtered off.

Brine water was taken in a small tank and *A. salina* cysts (1.5g/L) were added. Constant temperature (37°C) and sufficient light were maintained to give the sufficient aeration. After 24h, shrimp nauplii were appeared, which were then collected and used in the experiments.

2.4.4.3. Experimentation of lethality test

All the three (PetE, CHCl $_3$, and CH $_3$ OH) extract samples were applied against brine shrimp nauplii. For each samples, a 'pilot' test was done before final experimentation. 2mg sample extract was weighed and taken in a small glass vial, and then 1-2 drops of pure Dimethyl sulfoxide (DMSO) added to dissolve initially before adding 1ml of pond water to mix up the sample extract with water to prepare a 200ppm dose. When it mixed up completely, 1ml more water added to the test-tube (10ml marked) for giving a concentration of 100ppm.Half of it was used as the Dose A, and in the rest 1ml of water was added to give 2ml in amount with a concentration of 50ppm. This process was also maintained during final experiment. Separate vials were taken for each of the doses. For each and every dose 3 replicates were maintained. The final doses for cytotoxicity test were obtained as follows:

- For *E. nummularius* (wp) in PetE: 100, 50, 25, 12.5, 6.25 and 3.125ppm; CHCl3; 400, 200, 100, 50 and 25ppm; CH3OH: 400, 200, 100, 50, 25 and 12.5ppm;
- ► For *L. camara* (ap) in PetE: 100, 50, 25, 12.5 and 6.25ppm; CHCl₃: 100, 50, 25, 12.5 and 6.25ppm; CH₃OH: 200, 100, 50, 25 and 12.5ppm;
- For *L. camara* (r) in PetE: 100, 50, 25, 12.5 and 6.25ppm; CHCl3: 200, 100, 50, 25 and 12.5ppm; $CH₃OH$: 200, 100, 50, 25 and 12.5ppm;
- **Example 7 For** *M. piperita* (wp) in PetE: 100, 50, 25, 12.5, 6.25 and 3.125 ppm; CHCl₃: 100, 50, 25, 12.5 and 6.25ppm; CH₃OH: 400, 200, 100, 50 and 25ppm;
- For *Mi. pudica* (wp) in PetE: 100, 50, 25, 12.5 and 6.25ppm; CHCl3: 200, 100, 50, 25 and 12.5ppm; CH₃OH: 400, 200, 100, 50 and 25ppm;
- For *P. hysterophorus* (wp) in PetE: 200, 100, 50, 25 and 12.5ppm; CHCl3: 200, 100, 50, 25 and 12.5ppm; CH₃OH: 200, 100, 50, 25 and 12.5ppm;
- ► For *Ph. niruri* (wp) in PetE: 400, 200, 100, 50, 25 and 12.5ppm; CHCl₃: 200, 100, 50, 25, 12.5 and 6.25ppm; CH₃OH: 800, 400, 200, 100, 50 and 25ppm;
- For *Po. hydropiper* (wp) in PetE: 200, 100, 50, 25, 12.5 and 6.25ppm; CHCl3: 100, 50, 25, 12.5, 6.25 and 3.125ppm; CH₃OH: 200, 100, 50, 25 and 12.5ppm;
- For *Pz. zeylanica* (wp) in PetE: 100, 50, 25, 12.5, 6.25 and 3.125ppm; CHCl3: 200, 100, 50, 25 and 12.5ppm; CH3OH: 200, 100, 50, 25 and 12.5ppm;
- For *S. nodiflora* (wp) in PetE: 200, 100, 50, 25, 12.5 and 6.25ppm; CHCl3: 400, 200, 100, 50, 25 and 12.5ppm; CH₃OH: 800, 400, 200, 100 and 50ppm;
- For *Z. zerumbet* (ap) in PetE: 100, 50, 25, 12.5, 6.25 and 3.125ppm; CHCl3: 200, 100, 50, 25 and 12.5ppm; CH₃OH: 400, 200, 100, 50 and 25ppm;
- For *Z. zerumbet* (rh) in PetE: 100, 50, 25,12.5 and 6.25ppm; CHCl3: 100, 50, 25,12.5 and 6.25ppm; CH3OH: 400, 200, 100, 50 and 25ppm

Plate 2.14: Cyst of brine shrimp Nauplii

Plate 2.15: Bioassay with plant extracts on *A. salina* nauplii by brine shrimp lethality test

2.4.5. Larvicidal test against mosquito larvae

2.4.5.1. Experimental design for larvicidal test

Mosquito eggs are hatched in stagnant water. Test samples were prepared by the addition of calculated amount of DMSO (Dimethyl sulfoxide) to obtain desired concentration of the test samples. The larvae were counted by visual inspection and were taken in test-tubes containing 5ml of pond water. Then samples of different concentrations were added up to the pre-marked test-tubes by using a pipette. The test-tubes were left for 30h and then the larvae were counted again after 6h intervals to find out the lethality of the test materials.

Test materials:

- * Mosquito eggs in rafts;
- * Small beaker with pond water to hatch the eggs;
- Pipette (1ml and 10ml); and
- * Test tubes (20ml).

2.4.5.2. Preparation of environment for the hatching of eggs

Collected mosquito egg-rafts were placed in a beaker containing pond water and kept in a dark place in the lab for hatching out of the larvae. After 24 hours, hatched larvae were collected and used in the experiments.

2.4.5.3. Experimentation for larvicidal test

All the three (PetE, CHCl₃ and CH₃OH) of the test plants extracts samples were applied against mosquito larvae. 'Pilot' test was done before final experimentation. For each sample, 2mg extract sample was weighted and taken in a small glass vial, and then 1-2 drops of pure Dimethyl sulfoxide (DMSO) added to dissolve initially.

One ml of pond water was taken into the vial to mix up the sample extract with water to prepare 200ppm dose. When it mixed up completely added to the test-tube (10ml marked) for conducting tests. This process was also maintained during final experiment. Separate vials were taken for each dose. For each dose three replications were made.

The final doses for larvicidal test were

obtained as follows:

Plate 2.16: Bioassay with plant extracts on mosquito larvae by larvicidall activity test

 For *E. nummularius* (wp) in PetE: 200, 100, 50, 25, 12.5 and 6.25ppm; CHCl3; 400, 200, 100, 50, 25 and 12.5ppm; CH₃OH: 800, 400, 200, 100, 50 and 25ppm;

- **► For** *L. camara* (ap) in PetE: 100, 50, 25, 12.5 and 6.25ppm; CHCl₃: 400, 200, 100, 50 and 25ppm; CH₃OH: 200, 100, 50, 25 and 12.5ppm;
- For root part of *L. camara* (r) in PetE: 100, 50, 25, 12.5 and 6.25ppm; CHCl3: 800, 400, 200, 100 and 50ppm; CH3OH: 200, 100, 50, 25 and 12.5ppm;
- For *M. piperita* (wp) in PetE: 200, 100, 50, 25 and 12.5ppm; CHCl3: 400, 200, 100, 50 and 25ppm; CH₃OH: 800, 400, 200, 100 and 50ppm;
- For *Mi. pudica* (wp) in PetE: 400, 200, 100, 50 and 25ppm; CHCl3: 800, 400, 200, 100 and 50ppm;
- For *P. hysterophorus* (wp) in PetE: 200, 100, 50, 25 and 12.5ppm; CHCl3: 800, 400, 200, 100 and 50ppm; CH₃OH: 200, 100, 50, 25 and 12.5ppm;
- For *Ph. niruri* (wp) in PetE: 100, 50, 25,12.5, 6.25 and 3.125ppm; CHCl3: 200, 100, 50, 25, 12.5 and 6.25ppm; CH3OH: 800, 600, 400, 200, 100 and 50ppm;
- For *Po. hydropiper* (wp) in PetE: 100, 50, 25, 12.5 and 6.25ppm; CHCl3: 200, 100, 50, 25, 12.5 and 6.25ppm; CH₃OH: 200, 100, 50, 25, 12.5 and 6.25ppm;
- For *Pz. zeylanica* (wp) in PetE: 200, 100, 50, 25, 12.5 and 6.25ppm; CHCl3: 400, 200, 100, 50, 25 and 12.5ppm;
- For *S. nodiflora* (wp) in PetE: 200, 100, 50, 25, 12.5 and 6.25ppm; CHCl3: 400, 200, 100, 50 and 25ppm; CH₃OH: 400, 200, 100, 50, 25 and 12.5ppm;
- For *Z. zerumbet* (ap) in PetE: 400, 200, 100, 50 and 25ppm; CHCl3: 800, 400, 200, 100 and 50ppm;
- For *Z. zerumbet* (rh) in PetE: 50, 25, 12.5, 6.25 and 3.125ppm; CHCl3: 800, 400, 200, 100 and 50ppm; CH₃OH: 800, 400, 200, 100 and 50ppm

2.4.5.4. Analysis of data

The mortality records of the cytotoxicity test done on the nauplii of *A. salina* and mosquito larvae were corrected by the Abbott's (1925) formula:

$$
P_r = \frac{P_o - P_c}{100 - P_c} \times 100
$$

Where,

 P_r = Corrected mortality (%)

 P_0 = Observed mortality (%)

 P_c = Control mortality (%), sometimes called natural mortality (%).

Then mortality percentages were subjected to statistical analysis according to Finney (1947) and Busvine (1971) by using 'computer software'. The dose-mortality relationship was expressed as a median lethal concentration (LC_{50}).

2.4.6. Antimicrobial tests

The antimicrobial screening of an agent is essential to as certain its spectrum of activity against various types of pathogenic organisms. Antimicrobial activity of all the three (PetE, CHCl $_3$ and CH $_3$ OH) of the test plants extracts samples can be detected by observing the growth response of various microorganisms to the test plants extracts.

2.4.6.1. *In vitro* **antibacterial screening**

In general antimicrobial screening *in vitro* is undertaken in the following two steps:

(i) Primary assay: It is essentially a qualitative or semiqualitative test that indicates the sensitivity or resistance of microorganisms to the compound. However, this technique cannot be used to distinguish between bacteriostatic and bactericidal agents (Reiner, 1980). The primary assay can be done in three ways, such as-

- A. Diffusion method
- B. Dilution method and
- C. Bioautographic method.

Among these methods the disc diffusion method (Bauer *et al.* 1966; Reiner, 1982) is widely acceptable for the preliminary evaluation of antimicrobial activity. It uses different concentrations of the agents absorbed on sterile filter paper discs. There is no standardized method for expressing the results of antimicrobial screening (Ayafar *et al.* 1982). Some investigators use the diameter of the zone of inhibition or the minimum weight of extract that inhibits the growth of a microorganism. Disc diffusion is essentially a qualitative or semiquantitative test indicating the sensitivity or resistance of microorganisms to the test material. No distinction between bacteriostatic and bactericidal activity can be made by this method (Reiner, 1982). However *in vitro* antibacterial activity tests were done by disc diffusion method.

(ii) Secondary assay: It quantifies the relative potency such as minimum inhibitory concentration (MIC). The lowest concentration of an antimicrobial agent required to

inhibit the growth of the microorganisms *in vitro* is referred to as minimum inhibitory concentration (MIC). It is done by serial dilution technique (Reiner, 1980). The MIC measurement was done by dilution technique in this experimentation.

Principle of the diffusion method

Diffusion assay (Barry, 1976) is based on the ability of antibiotics to diffuse from a confined source through the nutrient agar gel and create a concentration gradient. If the agar is seeded or streaked with a sensitive organism, a zone of inhibition will result where the concentration exceeds the minimum inhibitory concentration (MIC) for the particular organism.

In this method, measured amount of the test samples are dissolved in definite volumes of solvent to give solutions of known concentrations (μ g/ml). The sterile (BBL, Cocksville, USA) filter paper (diameter 5mm) disc are impregnated with known amounts of the test substances and dried. These test material discs are placed on plates containing a suitable medium (nutrient agar) seeded with the test organisms. These plates are kept at low temperature $(4^{\circ}C)$ for 24h to allow maximum diffusion. A number of events take place simultaneously which includes-

- i) the dried discs absorb water from the agar medium and the material under test is dissolved,
- ii) the test material diffuses from the discs to the surrounding medium according to the physical law that controls the diffusion of molecules through agar gel,
- iii) there is a gradual change of test material concentration in the agar surrounding each disc.

In this study, sterile 5mm filter paper disks were treated with 200μ solvent only (used as a control), and 200μ l and 400μ l of each test plant extracts. The bacteria were inoculated on full-strength Nutrient Agar (Qualigens Fine Chemicals Prod # 58673) by suspending loops of bacteria in sterile de-ionized water. The bacterial suspension was then smeared on agar plates with a sterile glass-rod to ensure the entire surface of the agar had an even coating of the bacterial suspension. Plates were divided into several areas and one filter paper disk was placed in each area so that each plate had one disk of each treatment. Effects of the extracts on bacterial growth were quantified by measuring the diameter of the zones of inhibition less the size of the treated filter paper disks.

The plates are then kept in an incubator $(37^{\circ}C)$ for 12-18h to allow the growth of the organisms. If the test material has antimicrobial activity, it will inhibit the growth of microorganisms, giving a clear, distinct zone called 'Zone of Inhibition'. Effects of the different plant extracts on bacterial growth were quantified by measuring the diameter of the zones of inhibition in term of mm. The size of the inhibitory zones depends principally on the following factors:

- i) Intrinsic antimicrobial sensitivity of the test sample
- ii) Growth rate of the test microorganisms
- iii) Diffusion rate of the freshly seeded test organisms
- iv) Concentration of the freshly seeded test organisms
- v) Amount of test sample on disc
- vi) Thickness of the test medium in the Petri dishes
- vii) Composition of the culture medium
- viii) Inoculums size
- ix) Incubation time
- x) Temperature of incubation

Test materials used for the study

- i) PetE, CHCl $_3$ and CH $_3$ OH extract of test plant
- ii) Ampicillin (10 μ g/disc) as standard discs

Apparatus and reagents

- i) Blank sterilized filter paper discs (diameter 5mm)
- ii) Petri dishes (diameter 90mm)
- iii) Test tubes
- iv) Inoculating loop
- v) Spirit burner and a match box
- vi) Sterile forceps
- vii) Sterile cotton
- viii) Laminar air flow unit (BIOCRAFT and SCIENTIFIC INDUSTRIES, INDIA)
- ix) Micropipette $(10\mu 100\mu)$
- x) Autoclave (ALP Co. Ltd. KT- 30L, JAPAN)
- xi) Incubator (OSK- 9639A, Japan)
- xii) Refrigerator (ARISTON, ITALY)
- xiii) Punch machine
- xiv) Beaker
- xv) Nutrient agar media (DIFCO)
- xvi) Solvents (PetE, CHCl $_3$ and CH $_3$ OH)
- xvii) Vials
- xviii) Rectified spirit and
- xix) Alcohol (95%)

2.4.6.2. Test organisms used for the antibacterial activity test

Eight pathogenic bacteria were selected for the test, five of which were Gram negative and the remaining were Gram positive. These organisms of pure culture were collected from the Department of Biochemistry, University of Rajshahi. The bacterial strains used for this investigation are listed in Table 2.2.

2.4.6.3. Sterilization procedures

The antibacterial screening was carried out in a laminar airflow unit and all types of precautions were highly maintained to avoid any type of contamination during the test. UV light was switched on for half an hour before working in the laminar hood to avoid any accidental contamination. Petri dishes and other glass-wares were sterilized in the autoclave at 121° C temperatures and a pressure of 15 lbs. /sq. inch for 15 minutes. Micropipette tips, culture media, cotton, forceps, blank discs, etc were also sterilized.

2.4.6.4. Culture media

A number of culture media are available to demonstrate the antibacterial activity. These are:

- i) Nutrient agar medium
- ii) Nutrient broth medium
- iii) Mueller-Hinton medium
- iv) Tryptic Soy broth (TSB) medium
- v) Trypticase Soy agar medium
- vi) Staphylococcus defined medium
- vii) Adams and Roe medium
- **viii)** NTH agar or broth medium.

Table 2.3. Composition of nutrient agar medium (for preparation of 100ml media)

For demonstrating the antibacterial activity and subculture of the test organisms the nutrient agar media (DIFCO) was used.

2.4.6.5. Preparation of the nutrient agar (DIFCO) medium

The instant nutrient agar (DIFCO) medium was weighed and then reconstituted with distilled water in a conical flask according to specification $(2.3\% \text{ w/v})$. It was then heated in a water bath to dissolve the agar until a transparent solution was obtained.

2.4.6.6. Preparation of fresh culture of the pathogenic organisms

The nutrient agar medium was prepared and dispersed in a number of clean test tubes to prepare slants (5ml in each test tube). The test tubes were plugged with cotton and sterilized in an autoclave at 121 \degree C and 15lbs. /sq. inch pressure for 15min. After sterilization, the test tubes were kept in an inclined position for solidification. These were then incubated at 37.5° C to ensure sterilization. The test organisms were transferred to the agar slants from the supplied pure cultures with the help of an inoculating loop in an aseptic condition. Burning the loop after each transfer of microorganism was done to avoid contamination very carefully. The inoculated slants were then incubated at 37.5° C for 24h to assure the growth of test organisms. These fresh cultures were used for the sensitivity test.

2.4.6.7. Preparation of the test plates

The test plates were prepared according to the following procedure:

- (i) The nutrient agar medium prepared in the previous section was poured in 15ml quantity in each in the clean test tubes and plugged with cotton.
- (ii) The test tubes and a number of Petri dishes were sterilized in an autoclave at 121°C and 15lbs/sq. inch pressure for 15min and were transferred into laminar airflow unit and then allowed to cool to about 45° C to 50° C.
- (iii) The test organism was transferred from the fresh subculture to the test tube containing 15ml autoclaved medium with the help of an inoculating loop in an aseptic condition. Then the test tube was shaken by rotation to get a uniform suspension of the organism.
- (iv) The bacterial suspensions were immediately transferred to the sterile Petri dishes in an aseptic area. The petri dishes were rotated several times, first clockwise and then anticlockwise to assure homogenous distribution of the test organisms. The media were poured into petri dishes in such a way as to give a uniform depth of approximately 4mm.
- (v) Finally, after medium was cooled to room temperature in laminar airflow unit, it was stored in a refrigerator $(4^{\circ}C)$.

2.4.6.8. Preparation of discs containing samples

For the preparation of discs containing samples, following procedure was utilized:

(a) Sample discs: Sterilized filter paper discs (5mm in diameter) were taken by the forceps in to the plates. Sample solutions of desired concentrations were applied on the discs with the help of a micropipette in an aseptic condition. These discs were left for a few minutes in aseptic condition for complete removal of the solvent.

(b) Standard discs: These were used to compare the antibacterial activity of the test material. In the present study, Ampicillin discs containing $10\mu g/disc$ of antibiotic Ampicillin were used as standard discs for comparison purpose.

Plate 2.17: Antibacterial activity by disc diffusion method

2.4.6.9. Placement of the discs and incubation

For the placement of the discs, the following procedure was utilized:

- (i) By means of a pair of sterile forceps, the sample impregnated discs were placed gently on the solidified agar plates seeded with the test organisms to ensure contact with the medium.
- (ii) The plates were then kept in a refrigerator at 4° C for 24h in order to provide sufficient time to diffuse the antibiotics into the medium.
- (iii) Finally, the plates were incubated at 37.5° C for 24h in an incubator.

[Precaution: The discs were placed in such a way that they were not closer than 15mm to the edge of the plate and far enough apart to prevent overlapping the zones of inhibition].

2.4.6.10. Measurement of the zones of inhibition

After incubation, the antibacterial activities of the test samples were determined by measuring the diameter of inhibitory zones in term of millimeter (mm) with a transparent scale.

2.5. Chromatographic techniques used in this investigation 2.5.1. Chromatography on TLC plates

Thin layer chromatography is a very convenient technique for finding the separation slurry along with its stationary phase. The mixtures of the compounds were well separated from each other and resolved by preparative thin layer chromatographic technique. This tool is considered to be one of the most helpful methods of the detection of organic compounds, which involves an adsorbent (using silica gel) as stationary phase and a solvent system as a mobile phase. Due to the differential rate of adsorption on the adsorbent, the components in a mixture migrate differentially along with the TLC plate. In other words due to the difference in mobility of the components often depend on their polarity and that of the solvents used.

To select the solvent system for the run of the open column separation was made on the preparative thin layer chromatographic plates. For the normal phase chromatography silica gel G60 F_{254} on Al sheets (Merck) were used. Ten mg/ml of the sample in the solvent extraction offered 100μ l/spot by spotting 10μ l for each of the sample extracts. The chromatograms then developed within a conventional chamber with the following solvent systems: $CH_3OH - CHCl_3(1:1)$, CHCl₃ - EtOAc (ethyl acetate) (15:1) for PetE extract of *E. nummularius* (wp) and $CH_3OH - CHCl_3$ (1:1) for MeOH extract of *Po. hydropiper* (wp). All chromatograms were observed on TLC by Godin revelation and marked with a pencil.

2.5.2. Detection of the compound on TLC by Godin revelation

The properly developed plates were dried and sprayed with Godin reagent (Godin, 1954) to reveal the bands.

Visual detection: The development chromatogram was examined visually to detect the presence of colored compound.

Godin reagent spray: Equal volume of 1% ethanolic solution of vanillin and 3% aqueous solution of percloric acid was mixed sprayed on to the prepared chromatogram and 10% ethanolic solution of H_2 SO₄ was also sprayed afterwards and allowed the plate to dry out at 100 \degree C by using a hair dryer. Revelation was observed in different colors for different compounds (Godin, 1954).

Measurement of *Rf* **values:** The *Rf* values of he separated compounds were calculated on the developed chromatogram using the pre-established solvent system. The *Rf* values were calculated by the following formula:

 Rf = Distance traveled by the compound Distance traveled by the solvent

2.5.3. Open column chromatography

Of the methods in the solid phase category, column chromatography is very popular and used extensively. It can include non-exchange resins, polymeric columns, gelfiltration and chromatography over silica gel or chemically modified silica gel. Open column chromatography has a high load capacity but the separation time is long and the resolution is respectively low.

Column running **Fraction**

The stationary phase for the open column chromatography was silica gel Si60 [200- 400 mesh for PetE extract of *E. nummularius* (wp) and *Po. hydropiper* (wp)] (Merck),

and glass column of different size $(4.8\times36$ cm, 4.0×27 cm etc.) were used. Cotton pads washed with relevant solvent system was used at the base of the gel column. A similar cotton cloud was used at the top of the column (after application of the sample and the solvent) to protect destruction of the sample layer. Selected solvent systems were used as eluents and the elution rate was 1ml/min. Fractions were collected carefully.

2.5.4. Gel filtration

Open column for gel filtration generally used to apply sephadex LH-20 (Pharmacia) for the chromatography of exclusion. For methanol soluble samples MeOH (100%) and for $CHCl₃$ soluble samples $CHCl₃$ -MeOH (1:1) systems were used. The eluent allowed about 0.5ml/min.

2.5.5. Preparative separation techniques

Chromatography is an analytical technique for separating compounds on the basis of differences in affinity for a stationary and mobile phase. The separation of pure constituents from plant materials chromatography is a popular technique. The aim of choice any technique for separation is to have maximum yield with minimum effort to reduce the time and cost of the separation procedure. Preparative separation techniques can be tedious and time consuming, especially when complex mixtures, such as crude plant extracts have to be resolved. In the present study for isolation of pure compounds from PetE extract of *E. nummularius* (wp) and from MeOH extract of *Po. hydropiper* (wp) were done mainly by open column chromatography, while thin layer chromatography (TLC) and gel filtration were used as supporting tools.

2.5.6. Selection of extracts for fractionation

For fractionation of the extraction with a view to isolate biologically active compounds all the extracts were subjected to biological assay. Repetition of the same assay is required until the purification of the target compound, and thus a suitable bioassay technique was selected.

2.5.7. Selection of slurry (solvent system) for respective extracts

Aluminiun backed precoated preparative thin layer chromatographic (TLC) plates $(20 \times 20 \text{cm})$ with silica gel GF₂₅₄ with 0.5mm thickness and active in the usual manner (Merck, Germany) were used. The sample was applied on the activated plates with the help of a gradient micropipette as a narrow band at 1cm above the lower edge of the plate to make sure that the sample was washed away when the plates were placed inside the TLC chamber with the solvent system. The plates were developed in the usual manner.

After development, the chromatograms were air dried and observed sprayed with Godin reagent (Godin, 1954). The distinct bands were expected and by changing the solvent system with increase of either the polar or the apolar one. After having a better separation the selected solvent system was applied on the open column for isolation the compounds by fractionation. Small pieces of aluminum backed TLC plate was taken to spot the target extracts and run with a mixture of a relatively polar and relatively apolar solvent (1:1). For the better separation on the TLC with a known stationary phase the amount of both solvents were increased or decreased and applied accordingly. The combination given a better separation was selected for that extract for fractionation on the open column.

2.6. Isolation of the compounds

2.6.1. Isolation of *E. nummularius* **(wp) compound (ENP) from the PetE extract**

For the first fractionation of the PetE extract of *E. nummularius* (wp) LH_{20} (Pharmacia) was used as the stationary phase and $CHCl₃$ -MeOH (1:1) was the eluent on a glass column of 4.8 \times 36cm for 500mg of the extract. Elution time was adjusted to yield 1ml/min. It gave 71 tubes, which were then spotted on TLC to run and reveal the compounds by reagent spray. Three fractions were made for tubes 1-22 (Fr. I), 23-55 (Fr. II), 56-71 (Fr. III). The 2^{nd} fraction was targeted compound and subjected fractionation on selecting a solvent system by TLC, CHCl₃ and EtOAc (CHCl₃-EtOAc, 15:1) was applied on a glass column of 4.0×27 cm was packed with silica gel (200-400 mesh, 85gm) (Sigma). The elution was kept similar to that of the previous one. This fractionation yielded 151 tubes and TLC was made for all of them to get 6 fractions as Sfr. I (T/ 1-65), Sfr. II (T/ 66-78) and Sfr. III (T/ 79-94) Sfr. IV (T/ 95-103) Sfr. V (T/ 104-117) Sfr. VI (T/ 118-151). The Fr. III was tested on TLC under UV and Godin reagent spray to give Light blue color with a single spot compare to the crude. The compound was named as ENP (Fig 2.5).

Fig 2.5: Isolation pathway of *E. nummularius* (wp/PetE) compounds

Plate 2.19: Revelation of compound 1 (ENP) spots by Godin reagent spray

2.6.2. Isolation of *Po. hydropiper* **(wp) compound (POM) from the MeOH extract**

For the first fractionation of the MeOH extract of the Po . hydropiper (wp) LH_{20} (Pharmacia) was used as the stationary phase and $CHCl₃$ - MeOH (1:1) was the eluent on a glass column of 4.8×36 cm for 500mg of the extract. Elution time was adjusted to yield 1ml/min. It gave 59 tubes, which were then spotted on TLC to run and reveal the compounds by reagent spray. Four fractions were made for test tubes 1-22 (Fr. I), 23- 48 (Fr. II), 49-56 (Fr. III) and 57-59 (Fr. IV). The Fr. III was tested on TLC under Godin reagent spray to give light brownish color with a single spot compare to the crude. The compound was named as POM (Fig 2.6).

Fig 2.6: Isolation pathway of *Po. hydropiper* (wp/MeOH) compounds

Plate 2.20: Revelation of compound 2 (POM) spots by Godin reagent spray

RESULTS

3.1. Bioassay on eggplant aphid, *A. gossypii*

3.1.1. Dose mortality effect of the test extracts against *A. gossypii* **through residual film assay**

All the extracts (PetE, CHCl₃ and CH₃OH) of the *E. nummularius* (wp), aerial and root part of *L. camara*, *M. piperita* (wp), *Mi. pudica* (wp), *P. hysterophorus* (wp), *Ph. niruri* (wp), *Po. hydropiper* (wp), *Pz. zeylanica* (wp), *S. nodiflora* (wp), aerial and rhizome part of *Z. zerumbet* were tested against the eggplant aphid, *A. gossypii* with doses (mentioned in Chapter 2/2.4.2.1.) established through *Ad Hoc* experiments done prior to go through final experimentation. The respective doses for each of the extracts were applied on both the sides of treated leaves (taken as a part round in shape of 3.6cm diam.) and the test insects were released there to observe mortality or to find any sort of abnormality due to the efficacy of extracts compared to the controls (where no extracts were used). The results have been presented in Tables 3.1a - 3.1b, and Appendix Tables I - CCLXXVII.

The highest mortality was observed in case of PetE extracts of *E. nummularius* (wp) [LD₅₀] 0.034mg/cm²] followed by the PetE extracts of *Z. zerumbet* (rh) [LD₅₀ 0.035mg/cm²] after 24h of exposure against the eggplant aphid. The lowest mortality was observed in case of CHCl₃ extract of Ph. niruri (wp) [LD₅₀ 0.298mg/cm²] after 24h of exposure. According to the intensity of activity the extracts could be arranged in a descending order: *E. nummularius* (wp/PetE) *> Z. zerumbet* (rh/PetE) *> Z. zerumbet* (ap/PetE) *> L. camara* (r/PetE) *> E. nummularius* (wp/CH3OH) > *Ph. niruri* (wp/PetE) *> Z. zerumbet* (rh/CHCl3) > *S. nodiflora* (wp/CH3OH) > *Pz. zeylanica* (wp/PetE) > *P. hysterophorus* (wp/PetE) > *Pz. zeylanica* (wp/CHCl3) > *M. piperita* (wp/PetE) > *Z. zerumbet* (ap/CHCl3) *> Mi. pudica* (wp/CHCl3) > *P. hysterophorus* (wp/CHCl3) > *S. nodiflora* (wp/CHCl₃) > *E. nummularius* (wp/CHCl₃) > *Po. hydropiper* (wp /CHCl₃) > *L. camara* (ap/PetE) > *S. nodiflora* (wp/PetE) > *L. camara* (ap/CHCl3) > *Po. hydropiper* (wp/Pet.E) > *L. camara* (r/CHCl3) > *M. piperita* (wp/CHCl3) > *Z. zerumbet* (rh/CH3OH) > *Mi. pudica* (wp/CH3OH) > *M. piperita* (wp/CH3OH) > *L. camara* (ap/CH3OH) > *Po. hydropiper* $(wp/CH_3OH) > L.$ camara $(r/CH_3OH) > Mi.$ pudica $(wp/PetE) > Z.$ zerumbet $(ap/CH_3OH) > P.$ *hysterophorus* (wp/CH3OH) > *Ph. niruri* (wp/CH3OH) > *Pz. zeylanica* (wp/CH3OH) > *Ph. niruri* $(wp/CHCl₃)$.

Table 3.1a. LD50 values established through dose-mortality assay against *A. gossypii*

*** Variance has been adjusted for heterogeneity**

Table 3.1b. LD50 values established through dose mortality assay against *A. gossypii*

*** Variance has been adjusted for heterogeneity**

3.1.2. Repellent activity of the test extracts against *A. gossypii*

The PetE, CHCl₃ and CH₃OH extracts of test plants offered repellent activity against the eggplant aphids even for a concentration ranges between 0.393mg/cm² to as less as 0.025mg/cm² [0.393, 0.197, 0.098, 0.049 and 0.025mg/cm² for $\frac{1}{2}$ of treated area (3.6cm diam.) for all the plant extracts]. The data was read with 1h interval for up to 5h of exposure and was subjected to ANOVA after transforming them into arcsine percentage values which are given in Tables 3.2a - 3.2b and Appendix Tables DCCLII - DCCLXXXVII.

The extracts of *E. nummularius* (wp/CH3OH), *L. camara* (ap/CH3OH), *L. camara* (r/PetE & CH3OH), *M. piperita* (wp/CHCl3,), *Mi. pudica* (wp/CH3OH), *P. hysterophorus* (wp/CHCl3), *Po. hydropiper* (wp/PetE), *Pz. zeylanica* (wp/CH3OH) and *Z. zerumbet* (rh/CHCl³ & CH3OH) were found mildly active (P<0.05) but, the extracts of *M. piperita* (wp/CH3OH) was found moderately active (P<0.01) against the eggplant aphids. The PetE and CHCl₃ extracts of *E. nummularius* (wp), *L. camara* (ap), *Mi. pudica* (wp) and *Pz. zeylanica* (wp); all the solvent extracts of *Ph. niruri* (wp), *S. nodiflora* (wp) and *Z. zerumbet* (ap); PetE extracts of *M. piperita* (wp), *P. hysterophorus* (wp) and *Z. zerumbet* (rh); CHCl³ extracts of *L. camara* (r) and *Po. hydropiper* (wp) and CH3OH extracts of *P. hysterophorus* (wp) and *Po. hydropiper* (wp) gave no repellent activity against the eggplant aphids.

According to the intensity of repellency the result could be arranged in a descending order: *M. piperita* (wp/CH3OH) > *Po. hydropiper* (wp/PetE) > *Pz. zeylanica* (wp/CH3OH) > *L. camara* (r/PetE) > *L. camara* (r/CH3OH) > *P. hysterophorus* (wp/CHCI3) > *L. camara* (ap/CH3OH) > *Mi. pudica* (wp/CH3OH) > *Z. zerumbet* (rh/CH3OH) > *E. nummularius* (wp/CH3OH) > *M. piperita* $wp/CHCl₃$ > *Z. zerumbet* (rh/CHCl₃).

Table 3.2a. ANOVA results of repellency by test extracts against *A. gossypii*

*** = (P <0.05), ** = (P <0.01) and *** = (P <0.001), NS = not significant**

Table 3.2b. ANOVA results of repellency by test extracts against *A. gossypii*

*** = (P <0.05), ** = (P <0.01) and *** = (P <0.001), NS = not significant**
3.2. Bioassay on *A. salina* **nauplii**

3.2.1. Lethal effect of the test extracts against *A. salina* **nauplii through brine shrimp lethality assay**

All the extracts (PetE, CHCl₃ and CH₃OH) of the *E. nummularius* (wp), aerial and root part of *L. camara*, *M. piperita* (wp), *Mi. pudica* (wp), *P. hysterophorus* (wp), *Ph. niruri* (wp), *Po. hydropiper* (wp), *Pz. zeylanica* (wp), *S. nodiflora* (wp), aerial and rhizome part of *Z. zerumbet* were tested against the 1 day aged brine shrimp *A. salina* nauplii through lethality test with doses (mentioned in Chapter 2/2.4.4.3.) established through *Ad Hoc* experiments prior to set final experiments. The doses were applied in test-tubes, where the test organisms were released to observe lethality or any sort of abnormality caused due to efficacy of the extracts compared to the controls (where no extracts were used). Observation of mortality was made after 6h of application with 6h interval up to 30h. The data were then subjected to Probit analysis and the results have been presented in Tables 3.3a - 3.3b, and Appendix Tables CCLXXVIII - CDLV.

The highest lethality has been observed for the CHCl₃ extracts of *Po. hydropiper* (wp) [LC₅₀] 1.590ppm] followed by the PetE extracts of same plant $[LC_{50} 8.901$ ppm] after 30h of exposure against the brine shrimp nauplii. The lowest mortality was observed for the $CH₃OH$ extracts of *S. nodiflora* (wp) [LC₅₀ 197.230ppm] after 30h of exposure.

According to the intensity of activity the extracts could be arranged in a descending order: *Po. hydropiper* (wp/CHCl3) > *Po. hydropiper* (wp/PetE) > *M. piperita* (wp/PetE) > *L. camara* (ap/CHCl3) > *Z. zerumbet* (ap/PetE) > *Z. zerumbet* (rh/CHCl3) > *S. nodiflora* (wp/PetE) > *L. camara* (ap/PetE) > *Pz. zeylanica* (wp/PetE) > *L. camara* (r/PetE) > *M. piperita* (wp/CHCl3) > *E. nummularius* (wp/PetE) > *Z. zerumbet* (rh/PetE) > *L. camara* (r/CHCl3) > *L. camara* (r/CH3OH) > *E. nummularius* (wp/CHCl3) > *Po. hydropiper* (wp/CH3OH) > *Z. zerumbet* (ap/CHCl3) > *Mi. pudica* (wp/PetE) > *Ph. niruri* (wp/CHCl3) > *P. hysterophorus* (wp/PetE) > *P. hysterophorus* (wp/CH3OH) > *Pz. zeylanica* (wp/CH3OH) > *Mi. pudica* (wp/CHCl3) > *P. hysterophorus* (wp/CHCl3) > *Ph. niruri* (wp/PetE) > *L. camara* (ap/CH3OH) > *Pz. zeylanica* (wp/CHCl3) > *S. nodiflora* (wp/CHCl3) > *M. piperita* (wp/CH3OH) > *E. nummularius* (wp/CH3OH) > *Mi. pudica* (wp/CH3OH) > *Z. zerumbet* (ap/CH3OH) > *Z. zerumbet* (rh/CH3OH) > *Ph. niruri* (wp/CH3OH) > *S. nodiflora* (wp/CH3OH).

Table 3.3a. LC50 values of the test extracts established through brine shrimp lethality assay against *A. salina* **nauplii**

*** Variance has been adjusted for heterogeneity**

Table 3.3b. LC50 values of the test extracts established through brine shrimp lethality assay against *A. salina* **nauplii**

*** Variance has been adjusted for heterogeneity**

3.3. Bioassay on mosquito larvae

3.3.1. Effect of test extracts against *C. quinquefasciatus* **larvae through larvicidal assay**

All the extracts (PetE, CHCl₃ and CH₃OH) of the *E. nummularius* (wp), aerial and root part of *L. camara*, *M. piperita* (wp), *Mi. pudica* (wp), *P. hysterophorus* (wp), *Ph. niruri* (wp), *Po. hydropiper* (wp), *Pz. zeylanica* (wp), *S. nodiflora* (wp), aerial and rhizome part of *Z. zerumbet* were tested against the 1 day aged *C. quinquefasciatus* larvae through larvicidal activity test with doses (mentioned in Chapter 2/2.4.5.3.) established through *Ad Hoc* experiments done prior to set final experiments. The doses were applied in test-tubes, where the test organisms were released to observe lethality or any sort of abnormality took place due to efficacy of the extracts compared to the controls (where no extracts were used). Observation of mortality was made after 6h of application with 6h intervals up to 30h. The data were then subjected to Probit analysis and the results have been presented in Tables 3.4a - 3.4b, and Appendix Tables CDLVI - DCXX.

The highest lethality has been observed for the CHCl₃ extracts of *Ph. niruri* (wp) [LC $_{50}$] 3.220ppm] followed by the PetE extracts of same plant $[LC_{50}$ 3.390ppm] after 30h of exposure against the *C. quinquefasciatus* larvae. The lowest mortality was observed for the CH₃OH extracts of *M. piperita* (wp) [LC₅₀ 309.859ppm] after 30h of exposure. Methanol extract of *Mi. pudica* (wp), *Pz. zeylanica* (wp) and *Z. zerumbet* (ap) did not show any larvicidal effects.

According to the intensity of activity the extracts could be arranged in a descending order: *Ph. niruri* (wp/CHCl3) > *Ph. niruri* (wp/PetE) > *Z. zerumbet* (rh/PetE) > *Po. hydropiper* (wp/CH3OH) > *L. camara* (ap/PetE) > *Po. hydropiper* (wp/PetE) > *Pz. zeylanica* (wp/PetE) > *L. camara* (r/PetE) > *L. camara* (ap/CH3OH) > *P. hysterophorus* (wp/PetE) > *P. hysterophorus* (wp/CH3OH) > *L. camara* (r/CH3OH) > *Po. hydropiper* (wp/CHCl3) > *E. nummularius* (wp/PetE) > *M. piperita* (wp/PetE) > *S. nodiflora* (wp/PetE) > *Z. zerumbet* (ap/PetE) > *L. camara* (ap/CHCl3) > *E. nummularius* (wp/CHCl3) > *Pz. zeylanica* (wp/CHCl3) > *Mi. pudica* (wp/PetE) > *M. piperita* (wp/CHCl₃) > *S. nodiflora* (wp/CHCl₃) > *S. nodiflora* (wp/CH₃OH) > *P. hysterophorus* (wp/CHCl₃) > *E. nummularius* (wp/CH₃OH.) > *Z. zerumbet* (ap/CHCl₃) > *Z. zerumbet* (rh/CHCl3) > *Z. zerumbet* (rh/CH3OH) > *L. camara* (r/CHCl3) > *Ph. niruri* (wp/CH3OH) > *Mi. pudica* (wp/CHCl3) > *M. piperita* (wp/CH3OH).

Table 3.4a. LC50 values of the test extracts established through larvicidal assay against *C. quinquefasciatus* **larvae**

Table 3.4b. LC50 values of the test extracts established through larvicidal assay against *C. quinquefasciatus* **larvae**

3.4. Bioassay on *T. castaneum* **adults**

3.4.1. Effect of the test extracts against *T. castaneum* **adults through residual film assay**

All the extracts (PetE, CHCl₃ and CH₃OH) of the *E. nummularius* (wp), aerial and root part of *L. camara*, *M. piperita* (wp), *Mi. pudica* (wp), *P. hysterophorus* (wp), *Ph. niruri* (wp), *Po. hydropiper* (wp), *Pz. zeylanica* (wp), *S. nodiflora* (wp), aerial and rhizome part of *Z. zerumbet* were tested against the *T. castaneum* adults through residual film assay with doses (mentioned in Chapter 2/2.4.1.2) established (through *Ad Hoc* experiment) doses were applied on the inner surface of the petridishes, where the test insects were released to observe mortality or any sort of abnormality due to efficacy of the extracts compared to the controls (where no extracts were used). To trace acute toxicity an observation of mortality was made after 30min of application of the doses and followed by 12h intervals up to 48h. The data were subjected to probit analysis and the results have been presented in Table 3.5a - 3.5b and Appendix Table DCXXI - DCCLI.

The highest mortality has been observed for the CH₃OH extracts of *M. piperita (wp)* [LD₅₀] 0.238mg/cm²] and the lowest mortality was observed for the CHCl₃ extracts of *Po. hydropiper* (wp) $[LD_{50}$ 4.019mg/cm²] after 48h of exposure respectively; while the CHCl₃ and CH₃OH extracts of L. camara (r); all the solvent extracts of *Mi. pudica* (wp); CH₃OH extracts of Pz. zeylanica (wp); CH₃OH and CHCl₃ extracts of *L. camara* (root) and CHCl₃ extract of *S. nodiflora* didn't show any mortality against the adult beetles of *T. castaneum*.

According to the intensity of activity the extracts could be arranged in a descending order: *M. piperita* (wp/CH3OH) > *M. piperita* (wp/PetE) > *Po. hydropiper* (wp/CH3OH) > *Ph. niruri* (wp/PetE) > *Z. zerumbet* (rh/CH3OH) > *L. camara* (ap/CH3OH) > *Pz. zeylanica* (wp/PetE) > *Pz. zeylanica* (wp/CHCl3) > *E. nummularius* (wp/PetE) > *Z. zerumbet* (rh/PetE) > *S. nodiflora* (wp/PetE) > *M. piperita* (wp/CHCl3) > *Z. zerumbet* (ap/CH3OH) > *L. camara* (ap/CHCl3) > *Po. hydropiper* (wp/PetE) > *Z. zerumbet* (ap/PetE) > *P. hysterophorus* (wp/CH3OH) > *Ph. niruri* (wp/CH3OH) > *E. nummularius* (wp/CH3OH) > *E. nummularius* (wp/CHCl3) > *Ph. niruri* (wp/CHCl3) > *Z. zerumbet* (ap/CHCl3) > *Z. zerumbet* (rh/CHCl3) > *P. hysterophorus* (wp/PetE) > *L. camara* (r/PetE) > *S. nodiflora* (wp/CH3OH) > *P. hysterophorus* (wp/CHCl3) > *L. camara* (ap/PetE) > *Po. hydropiper* (wp/CHCl3).

Table 3.5a. LD50 values of the test extracts established through residual film assay against *T. castaneum* **adults**

*** Variance has been adjusted for heterogeneity**

Table 3.5b. LD50 values of the test extracts established through residual film assay against *T. castaneum adults*

*** Variance has been adjusted for heterogeneity**

3.4.2. Repellent effect of the test extracts against *T. castaneum* **adults**

The PetE, CHCl₃ and CH₃OH extracts of the test plants offered repellent activity against *T*. *castaneum* adults for a concentration ranges from 0.314 mg/cm² to as less as 0.025 mg/cm² [0.314, 0.157, 0.079, 0.039 and 0.019mg/cm² for $\frac{1}{2}$ of filter paper (9cm diam.) for all the extracts]. The data was read with 1h interval for up to 5h of exposure and was subjected to ANOVA after transforming them into arcsine percentage values which were presented in Tables 3.6a - 3.6b and the Appendix Tables DCCLXXXVIII - DCCCXXIII.

The extracts of *L. camara* (ap/ PetE); *L. camara* (r/CH₃OH); *M. piperita* (wp/CHCl₃ & CH₃OH); *Mi. pudica* (wp/ CHCl3); *P. hysterophorus* (wp/ PetE); *Ph. niruri* (wp/PetE) and *S. nodiflora* (wp/ PetE & CH₃OH) were weakly active (P<0.05) and extract of *E. nummularius* (wp/ PetE, CHCl₃ & CH3OH); *L. camara* (r/ CHCl3); *P. hysterophorus* (wp/ CHCl3); *Po. hydropiper* (wp/PetE, CHCl3); *Pz. zeylanica* (wp/PetE & CH3OH) and *S. nodiflora* (wp/CHCl3) were found moderately active (P<0.01) but, the extracts of *Po. hydropiper* (wp/CH3OH) were found highly active (P<0.001) against the *T. castaneum* adults. The extracts of *L. camara* (ap/ CHCl₃ & CH₃OH); *L. camara* (r/PetE); *M. piperita* (wp/PetE); *Mi. pudica* (wp/PetE & CH3OH); *P. hysterophorus* (wp/CH₃OH); *Ph. niruri* (wp/CHCl₃ & CH₃OH); *Pz. zeylanica* (wp/CHCl₃) and all solvent extracts of aerial and rhizome part of *Z. zerumbet* gave no repellent activity against the adult beetles of *T. castaneum*.

According to the intensity of repellency the result could be arranged in a descending order:

Po. hydropiper (wp/CH₃OH) > *Po. hydropiper* (wp/PetE) > *L. camara* (r/CHCI₃) > *E. nummularius* (wp/PetE) > *S. nodiflora* (wp/CHCI3) > *Pz. zeylanica* (wp/PetE) > *P. hysterophorus* (wp/CHCI3) > *Po. hydropiper* (wp/CHCI3) > *E. nummularius* (wp/CH3OH) > *Pz. zeylanica* (wp/CH3OH) > *E. nummularius* (wp/CHCI3) > *P. hysterophorus* (wp/PetE) > *Mi. pudica* (wp/CHCI3) > *L. camara* (ap/PetE) > *M. piperita* (wp/CHCI3) > *S. nodiflora* (wp/CH3OH) > *L. camara* (r/CH3OH) > *S. nodiflora* (wp/PetE) > *Ph. niruri* (wp/PetE) > *M. piperita* $(wp/CH₃OH)$

Table 3.6a. ANOVA results of repellency by selected plant extracts against *T. castaneum* **adults**

*** = (P <0.05), ** = (P <0.01) and *** = (P <0.001), NS = not significant**

 $\overline{}$

Table 3.6b. ANOVA results of repellency by selected plant extracts against *T. castaneum*

*** = (P <0.05), ** = (P <0.01) and *** = (P <0.001), NS = not significant**

3.5. Antibacterial activity of the test extracts

The PetE, CHCI $_3$ and CH $_3$ OH extracts of the selected plants were tested against 8 selected bacteria (3 gram-positive bacteria- *Bacillus subtilis*, *Listeria monocytogenes*, *Staphylococcus aureus* and 5 gram-negative bacteria- *Escherichia coli*, *Klebsiella pneumoniae*, *Salmonella enteritidis*, *Shigella flexneri* and *Shigella sonnei*) to evaluate their antibacterial potential at concentrations of 200µg disc⁻¹ and 400µg disc⁻¹ along with a standard antibiotic, Ampicillin 10µg disc-1 .The results obtained are shown in Tables 3.7a - 3.7c.

3.5.1. Antibacterial activity of the test extracts against some selected bacteria 3.5.1.1. Antibacterial activity of *E. nummularius* **(wp) extracts**

Among the collection CHCl3 extracts of *E. nummularius* (wp) showed the highest antibacterial activity. Among the selected test bacteria *B. subtilis*, *L. monocytogenes*, *Sa. enteritidis* and *St. aureus* were responsive. *B. subtilis* was most susceptible against CHCl₃ and CH₃OH extracts and gave inhibition zones 14mm (CHCl₃) and 16mm (CHCl₃) at concentrations of 200µg disc⁻¹ and 400μg disc⁻¹ respectively; 09mm (CH₃OH) at concentrations of 400μg disc⁻¹. While L. *monocytogenes, St. aureus* and *Sa. enteritidis* were responsive against the CHCl₃ extract and gave inhibition zones 09mm, 09mm and 12mm respectively for 400μg disc⁻¹ application.

3.5.1.2. Antibacterial activity of *L. camara* **(aerial part and root) extracts**

Among the *L. camara* extracts the root extract showed the highest antibacterial activity. Among the test bacteria *B. subtilis*, *L. monocytogenes*, *K. pneumoniae*, *Sa. enteritidis* and *Sh. flexneri* were responsive. *B. subtilis* was the most susceptible against the aerial (CHCl3) and root (CHCl₃) extracts, and gave inhibition zones 13mm and 18mm respectively for 200µg disc⁻¹ application and 15mm and 21mm respectively for 400 μ g disc⁻¹ application; and the root (CH₃OH) extract offered inhibition zones 10mm for 400μg disc⁻¹. While L. monocytogenes was susceptible against the aerial part (CHCl₃ and CH₃OH) extracts which gave inhibition zones 10mm and 09mm respectively for 200μg disc⁻¹ application and 12mm in both cases for 400μg disc⁻¹ application; and 09mm for the root (CHCl₃) extract for 400μg disc⁻¹. *K. pneumoniae*, Sa. *enteritidis* and *Sh. flexneri* were susceptible against the aerial part (CH3OH) extract, and gave inhibition zone 16mm,10mm and 14mm respectively for 400µg disc⁻¹ application.

3.5.1.3. Antibacterial activity of *M. piperita* **(wp) extract**

Among the three collection PetE extracts of this test plant showed the highest antibacterial activity, where *St. aureus*, *E.coli*, *K. pneumoniae, Sa. enteritidis* and *Sh. flexneri* were responsive among the selected test bacteria. *St. aureus* was responsive against the PetE extract and gave inhibition zones 12mm and 14mm at concentrations 200µg disc⁻¹ and 400µg disc-1 respectively. *E.coli* was responsive against PetE extract; *K. pneumoniae* was responsive against PetE and CHCl₃ extracts; *Sa. enteritidis* was responsive against PetE and CH₃OH extracts; and *Sh. flexneri* was susceptible against CH₃OH extract and gave inhibition zone 10mm in all the cases at a concentration of $400 \mu g$ disc⁻¹.

3.5.1.4. Antibacterial activity of *Mi. pudica* **(wp) extract**

Only *E. coli* was susceptible among the selected test bacteria against CHCl₃ extract of *Mi.* pudica (wp) and gave inhibition zones 09mm and 11mm at concentrations 200μg disc⁻¹ and 400 µg disc⁻¹ respectively; and against the CH₃OH extracts with the inhibition zone of 12mm at a concentration of 400µg disc⁻¹.

3.5.1.5. Antibacterial activity of *P. hysterophorus* **(wp) extract**

Among the collection CHCl₃ extracts of this test plant showed promising antibacterial activity against *B. subtilis*, *L. monocytogenes*, *E.coli* and *K. pneumoniae*. For *B. subtilis* the inhibition zones were 21mm and 25mm at concentrations of 200μg disc⁻¹ and 400μg disc⁻¹ respectively. While *L. monocytogenes* only susceptible against the CHCl₃ extract and gave inhibition zones 10mm for 400μg disc⁻¹ application; and *E. coli* was susceptible against the CHCl₃ extract that gave inhibition zones 10mm and 13mm for 200µg disc⁻¹ and 400µg disc⁻¹ respectively. While K. pneumoniae was susceptible against the CH₃OH extract by showing the inhibition zone of 09mm for 400μg disc⁻¹ application.

3.5.1.6. Antibacterial activity of *Ph. niruri* **(wp) extract**

Among the selected test bacteria only the *E. coli* was susceptible against the CHCl₃ and $CH₃OH$ extracts that offered inhibition zones 10mm and 12mm at 400 μ g disc⁻¹; and also gave 10mm for CH₃OH extracts at 200 μ g disc⁻¹ application.

3.5.1.7. Antibacterial activity of *Po. hydropiper* **(wp) extract**

Among the collection CHCl₃ extracts was found active that offered the highest antibacterial activity. Only the *B. subtilis*, *L. monocytogenes*, *St. aureus, Sa. enteritidis* and *Sh. flexneri* were responsive among the selected test bacteria. *B. subtilis* was most susceptible against the PetE and CHCl₃ extracts and gave inhibition zones 09mm (PetE) and 11mm (PetE) for 200μg

disc⁻¹ and 400µg disc⁻¹ respectively; 11mm (CHCl₃) and 13mm (CHCl₃) for 200µg disc⁻¹ and 400μg disc-1 respectively. While *L. monocytogenes*, *St. aureus* and *Sa. enteritidis* were susceptible against the CHC I_3 extracts and gave inhibition zone of 10mm in all the cases for 200μg disc⁻¹ and 12mm in all the cases for 400μg disc⁻¹. While Sh. flexneri was found susceptible against the CHCl₃ extract that gave inhibition zones 10mm for 400µg disc⁻¹.

3.5.1.8. Antibacterial activity of *Pz. zeylanica* **(wp) extract**

None of the extracts collected from *Pz. zeylanica* (wp) offered any activity against the 8 selected test bacteria.

3.5.1.9. Antibacterial activity of *S. nodiflora* **(wp) extract**

Among the three collections the PetE extract was the highest in response against some of the selected bacteria, *B. subtilis* and *K. pneumonia*. *B. subtilis* was the most susceptible one against PetE and CHC I_3 extracts and gave inhibition zones of 09mm (PetE) and 11mm (PetE) for 200μg disc⁻¹ and 400μg disc⁻¹ respectively; while the CHCl₃ extract gave inhibition zone of 09mm at 400μg disc-1 . While *K pneumoniae* was susceptible against the PetE extract and gave inhibition zone of 13mm and 17mm respectively for 200 and 400µg disc⁻¹ application.

3.5.1.10. Antibacterial activity of the *Z. zerumbet* **(aerial and rhizome part)**

Among the *Z. zerumbet* extracts the rhizome extract showed the highest antibacterial activity. The bacteria *B. subtilis*, *St. aureus*, *E.coli* and *K. pneumoniae* were responsive among the selected test bacteria. *B. subtilis* was susceptible against the rhizome (CHCl₃) extract and gave inhibition zones 09mm and 11mm respectively for 200µg disc⁻¹ and 400µg disc⁻¹ application. While *St. aureus* was susceptible against the aerial part (PetE) extract that gave 09mm inhibition zone for 400µg disc⁻¹. It also susceptible against the rhizome part (CHCl₃) that gave 10mm of inhibition zone for 200µg disc⁻¹ and 13mm of inhibition zone for 400µg disc⁻¹. While *E.coli* was only susceptible against the aerial part extract (PetE) that gave inhibition zone 10mm for 400μg disc⁻¹ application and *K. pneumoniae* rhizome extract (CHCl₃) gave inhibition zone of 09mm for 400 μ g disc⁻¹.

To compare the efficacy of the extracts against the selected bacteria a standard Ampicillin (10μg disc-1) was applied against the same bacteria through the similar procedure in application. The test bacteria *B. subtilis*, *L. monocytogenes, St. aureus, K. pneumoniae, Sa. enteritidis*, *Sh. flexneri* and *Sh. sonnei* were responsive against the standard provided with the inhibition zones 41, 40, 35, 35, 23, 34 and 35mm respectively; except *E. coli,* where no clear zone traced so far.

Table 3.7a. Antibacterial activity of the extracts and the standard Ampicillin

Table 3.7b. Antibacterial activity of the extracts and the standard Ampicillin

Table 3.7c. Antibacterial activity of the extracts and the standard Ampicillin

3.6. Interpretation of the isolated compounds

Two compounds from the extracts of two plants (ENP = Whole plant of *E. nummularius* PetE extract/COMP. 1 and POM = *Po. hydropiper* CH₃OH extract/COMP. 2) were subjected to NMR spectral analyses for their possible characterization.

The LC-MS analyses seemed they were not so pure, however, to have more information ¹H-NMR was run and the Mass spectra were made (Appendix Table DCCCXXIV & DCCCXXV); and according to the NMR spectra, the ENP (Comp. 1) seemed probably be a fatty acid, and the idea about POM (Comp. 2) remained unclear. The compound ENP might be a palmitic acid considering the chemical shifts and integration of the proton signals in the 1 H-NMR spectrum. Finding similarity with the palmitic acid in 1 H-NMR spectrum the 13 C-NMR of the compound was not run, however, the online information for the same was given in the following manner: Formula- $C_{16}H_{32}O_2$; Molecular weight- 256.4241; IUPAC Standard InChI-1S/ $C_{16}H_{32}O_2$ c1-2-3- 4-5-6- 7-8-9- 10-11- 12-13- 14-15-16 (17)18/h2-15H2,1H3,(H,17,18) and there were only two proton signals at 2.6 (3h, s) and 3.2 (2h, s) in the 1 H-NMR spectrum of POM and it was difficult to establish an idea what type of natural product it should be. The chemical structure of the compound 1 or ENP is given below:

Palmitic acid (Merck index, 1996)

The isolation and purification of the compounds through chromatographic techniques was maintained with the available facilities present there in the IES, University of Rajshahi and in the Crop Protection and Toxicology Lab, Department. of Zoology, University of Rajshahi using the solvents marketed under the brand Merck, Germany; however it was difficult to avoid impurities. The attempt of chromatographic fractionation was not an obligation in terms of the title of the current research, however introduction of the procedure of isolation of compound(s) would be an essential tool for further approach if the bioactive compound(s) are targeted for future endeavors; and this is why it was attempted so far.

3.7. Summary of the experimentation

3.7.1. Summary of the biological activities

For the detection of vector control potentials in PetE, CHCl₃, CH₃OH extracts of the test plants insecticidal, insect repellency, larvicidal and brine shrimp lethality tests were carried out. For further know the activity of the test plant the antimicrobial (antibacterial) test were done. The total outcomes of the bioassays done are represented in Table 3.8a-3.8d.

Types of extract		Test agents (tests)					
		A. gossypii		A. salina	C. quinquefasciatus	T. castaneum	
		mortality Dose	Repellency	lethality shrimp Brine	Larval lethality	mortality Dose	Repellency
E. nummularius (wp)	PetE	$\ddot{}$	L.	$\ddot{}$	$\ddot{}$	$\ddot{}$	$\ddot{}$
	CHCl ₃	$\ddot{}$	$\frac{1}{2}$	$\ddot{}$	$\ddot{}$	+	$\ddot{}$
	CH ₃ OH	$\ddot{}$	+	$\ddot{}$	$\ddot{}$	$\ddot{}$	$\ddot{}$
L. camara (ap)	PetE	$\ddot{}$	ä,	$\ddot{}$	$\ddot{}$	+	$\ddot{}$
	CHCl ₃	$\ddot{}$	$\overline{}$	$\ddot{}$	$\ddot{}$	+	$\overline{}$
	CH ₃ OH	$\ddot{}$	$\ddot{}$	$\ddot{}$	$\ddot{}$	$\ddot{}$	\blacksquare
L. camara (r)	PetE	$\ddot{}$	$\ddot{}$	$\ddot{}$	$\ddot{}$	+	$\overline{}$
	CHCl ₃	$\ddot{}$	$\overline{}$	$\ddot{}$	$\ddot{}$	\blacksquare	$\ddot{}$
	CH ₃ OH	$\ddot{}$	$\ddot{}$	$\ddot{}$	$\ddot{}$	\overline{a}	$\ddot{}$
M. piperita (wp)	PetE	$\ddot{}$	÷,	$\ddot{}$	$\ddot{}$	$\ddot{}$	\blacksquare
	CHCl ₃	$\ddot{}$	$\ddot{}$	$\ddot{}$	$\ddot{}$	$\ddot{}$	$\ddot{}$
	CH ₃ OH	$\ddot{}$	$\ddot{}$	$\ddot{}$	$\ddot{}$	$\ddot{}$	$\ddot{}$
Mi. pudica (wp)	PetE	$\ddot{}$	÷,	$\ddot{}$	$\ddot{}$	$\overline{}$	$\overline{}$
	CHCl ₃	$\ddot{}$	$\qquad \qquad \blacksquare$	$\ddot{}$	$\ddot{}$	$\qquad \qquad \blacksquare$	$\ddot{}$
	CH ₃ OH	$\ddot{}$	$\ddot{}$	$\ddot{}$	$\overline{}$	$\frac{1}{2}$	$\overline{}$

Table 3.8a. Summary of biological activities of the selected plant extracts

(+ = active, - = not active)

Table 3.8b. Summary of biological activities of the selected plant extracts

(+ = active, - = not active)

Table 3.8c. Summary of antibacterial activities of the selected plant extracts

(+ = active, - = not active)

Table 3.8d. Summary of antibacterial activities of the selected plant extracts

(+ = active, - = not active)

3.7.2. Summary of isolation, purification and characterization

Two compounds ENP (from *E. nummularius* PetE extract) and POM (from *Po. hydropiper* CH3OH extract) were subjected to NMR spectral analyses, while the ENP was determined as a palmitic acid and the POM was remained unknown.

DISCUSSION

Petroleum ether, chloroform and methanol extracts of *E. nummularius*, *M. piperita*, *Mi. pudica*, *P. hysterophorus*, *Ph. niruri*, *Po. hydropiper*, *Pz. zeylanica,* and *S. nodiflora* (whole plant)*, L. camara* (aerial part and roots) and *Z. zerumbet* (aerial part and rhizome) were screened against four vectors namely eggplant aphid, *A. gossypii* (plant disease vector); larvae of the mosquito, *C. quinquefasciatus* (human disease vector); rust-red flour beetle, *T. castaneum* (animal disease vector) and brine shrimp nauplii, *A. salina* (shrimp disease vector) under laboratory conditions to yield their efficacy through different assays with much success. Antimicrobial tests were also done to know the antibacterial activities of the extractives collected from the test plants.

In the present investigation the effects of extracts of the test plants on mortality of the eggplant aphid were found in the following order: E . *nummularius* (wp/PetE; LD₅₀ 0.034 mg/cm²) > Z. zerumbet (rh/PetE; LD₅₀ 0.035mg/cm²) > Z. zerumbet (ap/PetE; LD₅₀ 0.043 mg/cm²) > L. camara (r/PetE; LD₅₀ 0.053mg/cm²) > Ph. niruri (wp/PetE; LD_{50} 0.060mg/cm²) > S. nodiflora (wp/CH₃OH; LD₅₀ 0.065mg/cm²) > Pz. zeylanica $wp/PetE$; LD_{50} 0.067mg/cm²) > P. hysterophorus (wp/PetE; LD_{50} 0.068mg/cm²) > M. $piperita$ (wp/PetE; LD_{50} 0.070mg/cm²) > Mi. pudica (wp/CHCl₃; LD_{50} 0.071mg/cm²) > Po. hydropiper (wp/CHCl₃, LD₅₀ 0.079mg/cm²) > L. camara (ap/PetE; LD₅₀ 0.080mg/cm²)*.*

The results receive supports from many previous findings. Moawad and Barty (2011) found that ethanol and water extracts of five medicinal and ornamental plant species: *Aerva lanata*, *Ruta chalepensis*, *Fagonia arabica*, *Malva parviflora*, and *Calotropis procera*; were active against pomegranate aphid, *Aphis punicae* Passerini under laboratory conditions, while the ethanol extract of *R. chalepensis* (wp) showed the highest mortality against aphid. Neem extract generally recorded low aphid populations and an average fewer plants infected with symptoms of viral infestation (Sing and Korpraditskul, 1999). Karanja extract treated plants had minimal rate of incidence of viral infestation, with maximum plant height, flower production, fruit formation and highest yield (Bhyan *et al.*, 2007). According to Das *et al*., 2008 aphidicidal activity of hot and cold water extracts of some indigenous plants were tested against the bean aphid, *Aphis craccivora* Koch. Hot water extract of *Po. hydropiper* and *A. indica* were found to be the most effective (87.6 - 94.5 and 80.47 - 89.6% mortality respectively, P < 0.01) among all the extracts.

In case of larvicidal activity the results could be arranged in a descending order as follows: *Ph. niruri* (wp/CHCl₃; LC₅₀ 3.220ppm) $> Z$. zerumbet (rh/PetE; LC₅₀ 5.389ppm) > Po. hydropiper (wp/CH₃OH; LC₅₀ 21.432ppm) > L. camara (ap/PetE; LC₅₀ 22.950ppm) *> Pz. zeylanica* (wp/PetE; LC_{50} 24.226ppm) *> L. camara* (r/PetE; LC_{50} 28.532ppm) *> P. hysterophorus* (wp/PetE; LC⁵⁰ 37.858ppm) *> E. nummularius* (wp/PetE; LC⁵⁰ 45.345ppm) *> M. piperita* (wp/PetE; LC⁵⁰ 46.065ppm) *> S. nodiflora* (wp/PetE; LC⁵⁰ 50.361ppm) *> Z. zerumbet* (ap/PetE; LC⁵⁰ 55.681ppm) *> Mi. pudica* (wp/PetE; LC⁵⁰ 88.187ppm)*.* Methanol extract of *Mi. pudica* (wp), *Pz. zeylanica* (wp) and *Z. zerumbet* (ap) did not show any larvicidal effects.

Essential oil from the leaves of *L. camara* was reported to possess adulticidal activity against *Aedes aegypti*, *C. quinquefasciatus*, *Anopheles culicifacies*, *An*. *fluvialitis* and *An. stephensi* (mosquitoes) with LD_{50} values 0.06, 0.05, 0.05, 0.05 and 0.06mg/cm² (Dua, *et al*., 2010). Mosquito larvicidal assays through methanol and ethanol extracts of leaves and flowers of *L. camara* exhibited significant activity against 3rd and 4th instar larvae of *Ae. aegypti* and *C. quinquefasciatus* (Kumar and Maneemegalai, 2008). Bosly (2013) showed that the essential oils of peppermint and lavender have a control potential against *M. domestica*, which supports larvicidal activity of the aerial part of *L. camara* and *M. piperita* extracts in the present investigation. According to Bucker *et al*. (2013) CH3OH extracts showed higher activity against *A. nuneztovari* larvae than against *A. aegypti* larvae, suggesting that the extracts have speciesspecific activity. Cent percent mortality of *Anopheles stephensi* was observed in case of four plants, namely- *Albizia amara*, *Areca catechu*, *Leucas aspera* and *Ocimum* sanctum after 24h of exposures (Vinayagam *et al.*, 2008). The CHCl₃ extract of the fruit and the root of *Cleome viscosa* showed the highest and the second highest toxicity $(LC_{50}$ values were 185.390 and 272.910ppm after 30h of exposure respectively) against the larvae of *Culex* sp. (Islam *et al*., 2014). Kamaraj *et al*. (2011) reported that the ethyl acetate and methanol extract of stem bark of *Annona squamosa*, leaf extract of *Chrysanthemum indicum* and acetone and ethyl acetate extracts of leaf of *Tridax procumbens* have the potentials to be used as an eco-friendly approach for the control of the *Anopheles subpictus*, and *Culex tritaeniorhynchus*.

According to Batabyal *et al.* (2007) CH3OH extract of *Azadirachta indica* exhibited most potential larvicidal activity against *Anopheles stephensi* with LC₅₀ 15.25 and 12.70ppm after 24 and 48h of exposure. Murugan *et al.* (2013) reported on larval mortality of neem products against malarial vector *Anopheles stephensi* was dose-

dependent. The LC_{50} and LC_{90} values of Azadirachtin were 0.299% and 1.061%, respectively. To establish the larvicidal activities seeds of *Rauvolfia serpentina* were extracted with five solvents graded according to the polarity [viz. PetE, benzene, ethyl acetate, acetone and absolute alcohol] where mortality rate with PetE extract was significantly higher than other extracts (Das and Chandra, 2012). Citrus limonoids, nomilin and limonin, were used for larvicidal assay against *Aedes albopictus*. Results exhibited that citrus nomilin was more toxic than limonin against mosquito larvae (Hafeez *et al.*, 2011). The ethanol extract of unripe fruits of *Evodia rutaecarpa* Hook F. et Thomas (Rutaceae) was found to possess larvicidal activity against the mosquitoes larvae of the Culicidae mosquito *Aedes albopictus* (Liu *et al.*, 2012). Neem oil as larvicide against the main African malaria vector, *Anopheles gambiae* were observed by Okumu *et al.*, 2007. The steam distilled oils of 3 species of marigold, *Tagetes patula*, *T. erecta* and *T. minuta* were tested for larvicidal activity toward third instar larvae of *Aedes aegypti*; however activity at 10ppm was demonstrated only for *T. minuta* (Green *et al.*, 1991). The acetone, chloroform, ethyl acetate, hexane, and methanol extracts of leaf and flower of *Ocimum sanctum* were studied against fourth instar larvae of *Aedes aegypti* and *Culex quinquefasciatus*, where highest larval mortality was found in leaf extract of *O*. *sanctum* against the larvae of *A. aegypti* and *C. quinquefasciatus* (Anees, 2008). According to Kamaraj *et al.* (2010) the larval mortality was found in the hexane extract of *Zingiber zerumbet* (wp) against *Culex* gelidus (LC₅₀ 26.48) and against *Culex quinquefasciatus* (LC₅₀ 69.18ppm) after 24h of exposures. Kundu *et al.* (2013) reported that crude and ethyl acetate extracts of matured seed-coat of *Cassia sophera* can be used for the control of *Culex quinquefaciatus* larvae. The CH3OH extract of leaf of *Cocculus hirsutus* L. and *Tagetes erecta* L. have the potentials to be used as an ideal eco-friendly means for the control of *Anopheles subpictus* Grassi (Elango *et al.*, 2011). Larval mortality was found in case of acetone, chloroform, methanol, and PetE extract of leaf of *Canna indica* L. against second and fourth instar larvae of *C*. *quinquefasciatus* (Rahuman *et al.*, 2009). Mosquito larval mortality up to 93.33% and reduction of egg hatchability was observed in case of *Acacia nilotica* extract by Zaitoun in 2012. The leaf extract of *Ageratina adenophora* is more toxic to both *A. aegypti* and *C. quinquefasciatus* (Mohan and Ramaswamy, 2007).

The present investigation depicts the effects of extracts of the test plants through cytotoxicity test against *A. salina* were found in the following order: *Po. hydropiper* (wp/ CHCl3; LC⁵⁰ 1.590ppm) > *M. piperita* (wp/PetE; LC⁵⁰ 9.573ppm) > *L. camara*

(ap/CHCl3; LC⁵⁰ 10.481ppm) > *Z. zerumbet* (ap/PetE; LC⁵⁰ 10.938ppm) > *Z. zerumbet* (rh/CHCl3; LC⁵⁰ 11.172ppm) > *S. nodiflora* (wp/PetE; LC⁵⁰ 11.380ppm) > *Pz. zeylanica* (wp/PetE; LC⁵⁰ 11.648ppm) > *L. camara* (r/PetE; LC⁵⁰ 12.497ppm) > *E. nummularius* (wp/PetE; LC₅₀ 13.601ppm) > *Mi. pudica* (wp/PetE; LC₅₀ 23.815ppm) > *Ph. niruri* $(wp/CHCl₃; LC₅₀ 24.331ppm).$

According to Saha and Paul (2012a) ethanol extract of *Pouzolzia zeylanica* (L.) Benn posseses significant cytotoxic activity with the LC_{50} value of 6.1µg/ml and the LC_{90} value of 12.2μg/ml. Chloroform extracts of the fruit shell, leaves, root bark, root wood, seeds, stem bark and stem wood of *Derris indica* Bennet were found to be effective against the brine shrimp, A*rtemia salina* nauplii (Mondal *et al.*, 2012b). Rhizome essential oils of red and black varieties of *Cyperus articulatus* were tested for bioactivity using brine shrimp lethality test that showed significant activity with LC_{50} of 2.84g/ml and 3.34g/ml for red and black varieties respectively (Ameen *et al.*, 2011). Different concentrations 10, 100 and 1000ug/ml of the methanolic extract of medicinal herb *Ajuga parviflora* were used by Rahman *et al.*, 2013 for brine shrimp lethality bioassay and significant results were obtained. Chloroform extracts of the leaf, stem bark, stem wood and roots of *Glycosmis pentaphylla* (Retz.) were tested against the brine shrimp, *Artemia salina* nauplii for cytotoxic activity in which the dose-mortality assay revealed LC_{50} values of 28.579, 28.659, 57.213 and 84.111ppm respectively, for the plant parts efficacy of which could be arranged in the order of leaf > stem bark > stem wood > root (Pramanik *et al.*, 2009). The cytotoxic effect of *Cleome viscosa* extracts (PetE, CHCl₃ and CH₃OH) against the brine shrimp (A. salina) nauplii were found promising where PetE extract of the root and CHCl₃ extract of the fruit showed the highest and the second highest toxicity (LC_{50} values were 21.905 and 26.675ppm after 30h of exposure respectively) against the nauplii (Islam *et al.*, 2014). The cytotoxic activities of crude extract (CH3OH/fruits) of *Hibiscus sabdariffa* (MEHS) was determined using brine shrimp lethality assay and LC_{50} values of standard vincristine sulphate as a positive 50 control and the crude extract was found to be 0.21±0.19μg/ml and 5.082±0.12μg/ml respectively (Al-Mamun *et al.*, 2011). The methanol extracts of *Lantana camara* (Root, Stem, Leaf, Flower and Fruit) were tested for *In Vivo* brine shrimp lethality assay. All the tested extracts exhibited very low toxicity on brine shrimp nauplii. The results showed that the root extract was the most toxic part of *L. camara* (Badakhshan *et al.*, 2009). Crude ethanol extracts of the rhizome of *Z. zerumbet* showed the highest cytotoxicity (LC₅₀ was 1.24μg/ml) against brine shrimp nauplii as compared with that of *Curcuma zedoaria* Rosc. (LC₅₀ was

33.593μg/ml) after 24h of exposure (Hossain *et al.*, 2012); and it supports the findings of this investigation. Good cytotoxic activity (66.66%) was shown by the *n*-hexane fraction of crude methanolic extract of the aerial parts of *Myrsine africana* plant at 1000μg/ml (Ahmad *et al.*, 2011a). Preliminary cytotoxicity tests were done with the methanolic, hexane, chloroform, ethyl acetate and butanol fractions of *Vitex agnuscastus* using the nauplii of the brine shrimp, *Artemia salina*. These fractions were, however found to be relatively nontoxic (Azizuddin and Choudhary, 2011). In cytotoxicity determination, LC⁵⁰ of *Candida albicans* compound against brine shrimp nauplii was 13.25μg/ml (Khan *et al.*, 2008). Moderate level of cytotoxicity (LD₅₀ value 9.23μg/ml) against brine shrimp larvae was found with the methanolic extract of *Calendula arvensis* (Ullah *et al.*, 2012).

The mortality of *T. castaneum* adults through dose-mortality experiments by residual film method was done and the results have been analyzed and tabulated; While, no activity was traced for whole plant of *Mi. pudica*. While, no activity was traced for whole plant of *Mi. pudica*. The methanol extract of *Pz. zeylanica* (wp); methanol and CHCl₃ extracts of *L. camara* (r) and CHCl3 extract of *S. nodiflora* (wp) didn't show any mortality against the adult beetles of *T. castaneum*.

Mortality in LD₅₀ values for the rest extracts were found active in the following order: *M.* piperita (wp/CH₃OH; LD₅₀ 0.238mg/cm²) > Po. hydropiper (wp/CH₃OH; LD₅₀ 0.342mg/cm²) > Ph. niruri (wp/PetE; LD₅₀ 0.508mg/cm²) > Z. zerumbet (rh/CH₃OH; LD_{50} 0.793mg/cm²) > L. camara (ap/CH₃OH); LD_{50} 0.797mg/cm²) > Pz. zeylanica $wp/PetE$; LD_{50} 0.799mg/cm²) > E. nummularius (wp/PetE; LD_{50} 1.031mg/cm²) > S. nodiflora (wp/PetE; LD_{50} 1.248mg/cm²) > Z. zerumbet (ap/CH₃OH; LD_{50} 1.316mg/cm²) > P. hysterophorus (wp/CH₃OH; LD₅₀ 1.631mg/cm²) > L. camara (r/PetE; LD₅₀ 2.672 mg/cm²).

Insecticidal activity of the methanolic extract of *Synedrela nodiflora* leaves against the stored product pest *Sitophilus oryzae* using surface film method was found active and dose dependent that showed 95%, 96% and 98% activity against the doses of 20, 40 and 50mg/ml respectively after 12h of exposures (Haque *et al.*, 2012). Methanol extract of *Polygonum persicaria* (ap) and *Polygonum plebejum* (wp) showed significant insecticidal activities against *Tribolium castaneum* (Hussain *et al.*, 2010). According to Islam *et al.* (2014) CHCl₃ and CH₃OH extracts of *Cleome viscosa* (ap) showed the highest and the second highest mortality (LD_{50} values were 0.170 and 0.248mg/cm² respectively) against *T. castaneum* adults and CHCl₃ extract of the root part of that

plant was found to show no mortality against the adult beetles of *T. castaneum*. Ethanol extract of *Phyllanthus amarus* (root) possessed significant insecticidal activity against *T. castaneum* (Khanna *et al.*, 2003). Padin *et al.* (2000) reported that the essential oil of *Rosemary* (*Rosmarinus officinalis*) killed *Tribolium* adults. The crude methanolic extract and various fractions of *Zizyphus jujuba* except n-hexane showed low activity (20%) against *T. castaneum* (Ahmad *et al.*, 2011b). According to Rajashekar *et al.* (2012) the methanol extract collected from leaves of *Lantana camara* had fumigant and contact toxicity against *Sitophilus oryzae*, *Callosobruchus chinesis* and *Tribolium castaneum*. Petroleum spirit extract of custard apple, *Annona squamosa* L. seeds offered highest toxicity (LD₅₀ 58.697µg/cm²) on CTC-12 strain of the red flour beetle, *Tribolium castaneum* (Herbst) adults (Khalequzzaman and Sultana, 2006).The LD₅₀ results revealed that methanol extract of *Synedrella nodiflora* Gaertn is the most toxic to *Spodoptera litura* followed by benzene and chloroform, PetE and water extracts (Martin Rathi and Gopalakrishnan, 2006). According to Mamun *et al.* (2011) Methanol (85%) extract of fruits of *Hibiscus sabdariffa* showed the significant activity with 100% mortality of *Tribolium castaneum* at a dose of 50mg/ml with 12 hours of exposures. Low insecticidal activity (20%) was shown by chloroform (CHCl₃) and aqueous fractions of aerial parts of *Myrsine africana* against *Tribolium castaneum* (Ahmad *et al.*, 2011a). Methanolic extract and its fractions of *Vitex agnus-castus* did not show any insecticidal activity against *Tribolium castaneum* (Azizuddin and Choudhary, 2011). Chloroform and ethanol extracts of Bishkatali, *Polygonum hydropiper* were moderately toxic to *Tribolium castaneum*. The toxicity of ethanol extract was higher than chloroform extract after 24 and 72h of exposures (Kundu *et al.*, 2007).

Against the eggplant aphids CH3OH extracts of *E. nummularius* (wp), *L. camara* (ap), *Mi. pudica* (wp) and *Pz. zeylanica* (wp) offered repellent activity, while the PetE and CHCl₃ extracts of the same didn't show repellency; however PetE, CHCl₃ and CH₃OH the extracts of *Ph. niruri* (wp), *S. nodiflora* (wp) and *Z. zerumbet* (ap); PetE extracts of *M. piperita* (wp), P. hysterophorus (wp) and Z. zerumbet (rh); CHCl₃ extracts of L. *camara* (r) and *Po. hydropiper* (wp) and CH3OH extracts of *P. hysterophorus* (wp) and *Po. hydropiper* (wp) offered no repellent activity at all.

According to intensity of repellency the result could be arranged in a descending order: *M. piperita* (wp/CH3OH) > *Po. hydropiper* (wp/PetE) > *Pz. zeylanica* (wp/CH3OH) > *L. camara* (r/PetE) > *L. camara* (r/CH3OH) > *P. hysterophorus* (wp/CHCI3) > *L. camara*

(ap/CH3OH) > *Mi. pudica* (wp/CH3OH) > *Z. zerumbet* (rh/CH3OH) > *E. nummularius* (wp/CH3OH) > *M. piperita* (wp/CHCI3) > *Z. zerumbet* (rh/CHCI3).

For the repellency against *T. castaneum* adults CH₃OH extracts of *Po. hydropiper* (wp) offered the most promising activity, however except *L. camara* (ap/CHCl₃ and CH₃OH); *L. camara* (r/PetE); *M. piperita* (wp/PetE); *Mi. pudica* (wp/PetE and CH3OH); *P. hysterophorus* (wp/CH3OH); *Ph. niruri* (wp/CHCl3 and CH3OH); *Pz. zeylanica* (wp/CHCl₃) showed repellent activity of different degree; and aerial and rhizome part of *Z. zerumbet* extracts of all the three solvents showed no repellent activity.

According to intensity of repellency the result could be arranged in a descending order: *Po. hydropiper* (wp/CH₃OH) > *Po. hydropiper* (wp/PetE) > *L. camara* (r/CHCl₃) > *E. nummularius* (wp/PetE) > *S. nodiflora* (wp/CHCI3) > *Pz. zeylanica* (wp/PetE) > *P. hysterophorus* (wp/CHCI3) > *Po. hydropiper* (wp/CHCI3) > *E. nummularius* (wp/CH3OH) > *Pz. zeylanica* (wp/CH3OH) > *E. nummularius* (wp/CHCI3) > *P. hysterophorus* (wp/PetE) > *Mi. pudica* (wp/CHCI3) > *L. camara* (ap/PetE) > *M. piperita* (wp/CHCI3) > *S. nodiflora* (wp/CH3OH) > *L. camara* (r/CH3OH) > *S. nodiflora* (wp/PetE) > *Ph. niruri* (wp/PetE) > *M. piperita* (wp/CH3OH).

Gallardo *et al.* (2011) showed that essential oils (EOs) from *Tagetes lucida*, *Lepechinia betonicifolia*, *Lippia alba*, *Cananga odorata*, and *Rosmarinus* were repellent, followed a dose-response relationship and EOs from *C. odorata* and *L. alba* were the most active repellents against *T. castaneum*. Pramanik *et al.* (2009) observed that the F values of the arcsine transformed data were 60.983, 14.177, 19.437, 15.429 and 1.082 respectively for the repellency against *T. castaneum* adults for CHCI₃ extracts of leaf, stem bark, stem wood, root bark and root wood extracts of *Glycosmis pentaphylla*. Except for the root wood extract, strong repellent activity was observed for the rest of the extracts (P<0.001). *Abroma augusta* extracts can be used as a reduced risk repellent compound in the grain and cereal stores to manage the population of *T. castaneum* (Mondal, 2012a). Chloroform and ethanol extract of Bishkatali showed strong repellency against *T. castaneum* (like that of the mortality results) where chloroform extract was better than ethyl alcohol extract (Kundu *et al.*, 2007), and this finding supports the present study. Three plant species, leaf of *Polygonum hydropiper* Linn. (Bishkatali), *Vitex negundo* Linn. (Nishinda) and *Aphanamixis polystachya* (Pithraj) extracted with water and acetone were evaluated for their repellent and feeding deterrent activity against adult red flour beetle, *Tribolium castaneum* (Herbst). It was observed that Biskatali (leaf/water extract) have shown strong repellent and

feeding deterrent effect followed by Nishinda and Pithraj (Islam *et al.*, 2000). Petroleum and methanol extracts of *Cyperus articulatus* (rh) were evaluated against the red flour beetle, *Tribolium castaneum* (Hbst.) using standard techniques and both the extracts were observed to have similar repellant actions (Abubakar, 2000). Islam *et al*, 2014 reported that the CH3OH extract of *Cleome viscosa* (ap) showed the highest repellency between dose interval at 1% level of significance (P<0.01) against the adult beetles of *T. castaneum*. The PetE extract of the aerial part and the CHCl₃ extract of the root of that plant showed repellency at 5% level of significance (P<0.05) while the other parts (fruit) did not show significant repellency against the adult flour beetles*.*

The anitbacterial activity of the PetE, $CHCl₃$ and $CH₃OH$ extracts of the selected plants were tested against 8 selected pathogenic bacteria (3 gram-positive bacteria- *Bacillus subtilis*, *Listeria monocytogenes*, *Staphylococcus aureus* and 5 gram-negative bacteria- *Escherichia coli*, *Klebsella pneumoniae*, *Salmonela enteritidis*, *Shigella* flexneri and Shigella sonnei) at concentrations of 200μg disc⁻¹ and 400μg disc⁻¹ along with a standard antibiotic, Ampicillin 10 μ g disc⁻¹. The chloroform extract was found most effective in comparison to the PetE and CH₃OH extracts.

Pavithra *et al*. (2009) reported that CH3OH extract of *E. nummularius* has antibacterial activity against *Escherichia coli* (MIC, 12.50mg/ml) and *Bacillus subtilus* (MIC, 3.125mg/ml) and the most resistant strains were *Staphylococcus aureus*, *Klebsiella pneumoniae* and *Pseudomonas aeruginosa*. Barreto *et al.* (2010) showed that ethanolic extracts of leaves and roots of *Lantana camara* and *Lantana montevidensis* exhibited antibacterial activity against clinically relevant pathogens (gram positive and gram negative bacteria). *L. camara* leaf extract was active against *Proteus vulgaris* and *Víbrio cholerae* (MIC 128 μg/ml for both the strains); in addition to the root extract which was effective against *P. vulgaris* and *Pseudomonas aeruginosa* (64 and 128μg/ml respectively). The leaves and roots of *L. montevidensis* extracts were active against *P. vulgaris* and *P. aeruginosa* (MIC 8μg/ml) and two strains of *E. coli* (MIC 16μg/ml for the multi-resistant strain). Methanolic extracts of different parts of *L. camara* were screened for antimicrobial activity against 10 bacteria by disk diffusion method. The leaf extract of *L. camara* showed highest activity against the Gram positive bacteria, *Bacillus cereus* and Gram negative bacteria, *Salmonella typhi* (Badakhshan *et al.*, 2009). The ethanol extract of *Pouzolzia zeylanica* showed highest antibacterial activity against *Staphylococcus aureus*, *E. coli* compared with the standard drug amoxicillin (Saha *et al.*, 2012b). The antimicrobial activity of CH₃OH

extracts of different plant parts of *Phyllanthus niruri* were tested against 5 bacterial strains (*E. cloacae*, *S. aureus*, *P. aeruginosa*, *E. coli* and *S. viridians*). Individually against *E. coli* maximum IZ was observed in extract of leaves, which was as par with that of seeds (12mm) and minimum was in roots. In case of *S. aureus* maximum IZ was observed in seeds (16mm) and minimum in stem (7mm) reported by Mathur *et al.* (2012). Hasan *et al.* (2009) observed that chloroform extract of *Polygonum hydropiper* (L.) had significant antibacterial activities against the four gram-positive bacteria, *Bacillus subtilis*, *Bacillus megaterium*, *Stapphylococcus aureus* and *Enterobacter aerogenes*; and four gram-negative bacteria, *Escherichia coli*, *Pseudomonas aeruginosa*, *Salmonella typhi* and *Shigella sonnei* and the minimum inhibitory concentration (MIC) values against these bacteria ranged from 16 to 64μg/ml. Antibacterial activity of aqueous crude extracts of *Phyllanthus niruri* was effective against *Lactobacillus* strain (Kanthimathi and Soranam, 2013) and the methanol crude extract of *S. nodiflora* exhibited 14.0mm IZ against *Bacillus cereus* (Chowdhury *et al.*, 2013). The antibacterial potential of six organic solvent (ethanol, methanol, ethyl acetate, chloroform, hexane and PetE) extracts from leaf, stem and root of *Mentha piperita* against pathogenic bacteria, such as *Bacillus subtilis*, *Streptococcus pneumonia*, *Staphylococcus aureus*, *Escherichia coli*, *Proteus vulgaris and Klebsiella pneumonia* were evaluated by agar diffusion method. The leaf extract activity was found comparatively higher on *Bacillus subtilis*, *Staphylo-coccus aureus* and *Proteus vulgaris* than *Escherichia coli*, *Streptococcus pneumonia* and *Klebsiella pneumonia* (Sujana *et al.*, 2013). According to Bhogaonkar *et al.* (2011) PetE extracts of *Synedrella nodiflora* (leaves) showed maximum activity against all the test microbes (Strains used are *Bacillu*s *subtilis* NCIM 2063, *Staphylococcus aureus* NCIM 2079, *Escherichia coli* NCIM 2065, *Candida albicans* NCIM 3100 and *Aspergillus flavus* NCIM 519) tested except *E. coli*. Kumar *et al.* (2013) found that the aqueous and ethanolic extracts of leaf were able to inhibit *Bacillus cereus* MTCC 1272 with IZ 11±1.0mm. CH3OH extracts of *Mimosa pudica* displayed considerable bacteriostatic activity against all six bacterial strains (MIC range $= 0.625$ to 2.50mg/ml) including *Bacillus cereus*, *B. subtilis*, *Escherichia coli*, ampicillin-resistant *Escherichia coli*, *Staphylococcus aureus* and *Pseudomonas aeruginosa* (Genest *et al.*, 2008). The antibacterial activity of the compound Zederone which was isolated from the crude ethanolic extract of the rhizomes of *Zingiber zerumbet* (L.) Smith was determined against a number of multi-drug resistant and methicillin-resistant *Staphylococcus aureus* strains (SA1199B, ATCC25923, XU212, RN4220 and EMRSA15) and

minimum inhibitory concentration (MIC) values were found to be in the range of 64- 128ug/ml (Kader *et al.*, 2010). Ethyl acetate fraction of *Zizyphus oxyphylla* showed good antibacterial activity against *Bacillus subtilis* (IZ 16mm) and *Staphylococcus* aureus (IZ 18mm) (Nisar *et al.*, 2010). The antibacterial activity of the CH₃OH (85%) extract of fruits of *Hibiscus sabdariffa* (MEHS) were done against six Gram-positive and eight Gram-negative bacteria using disc diffusion method. The extract showed the highest activity against *Sarcina lutea* with an IZ 13±0.21mm followed by *Shigella dysenteriae* (12±0.07mm), *Escherichia coli* (12±0.11mm) and *Shigella boydii (*12±0.13mm) and inactivity against *Bacillus subtilis*, *B. megaterium*, *B. anthracis*, *B. cereus* and *P. aeruginosa* were also recorded (Mamun *et al.*, 2011). Hot aqueous extract of *Cassia senna* had the strongest inhibition effect on *Staphylococcus aureus* sub sp. *aureus*, *Salmonella typhimurium* and *Bacillus stearothermophilus*. *Acacia nilotica* showed only complete inhibition against *Staphylococcus aureus* sub sp. *aureus*, *Salmonella typhimurium* (Zaitoun *et al.*, 2012). The anti-listerial effect of water extract of garlic shoot juice (GSJ) was investigated against the four strains of *Listeria monocytogenes* ATCC 19116, 19118, 19166 and 15313. Various concentrations of (1%, 2.5% and 5%) were used and showed the strongest anti-listerial effect (Kim and Kang, 2007). Tamilarasi and Ananthi (2012) reported that the ethanolic extracts of *Mimosa pudica* leaves showed antimicrobial activity against *Bacillus subtilis*, *Pseudomonas aeruginosa*, *Klebsiella pneumonia* may be due to the presence of active constituents like alkaloids or tannins.

Finding potential activity the PetE extract of *E. nummularius* (wp) and CH3OH extract of *Po. hydropiper* (wp) were attempted for chromatographic fractionation to isolate bioactive compound(s) and as a result two compounds named ENP and POM were isolated, and only the ENP was determined as palmitic acid. However, isolation of compound(s) from each of the test plants was rather impossible due to limitations in the laboratory supports, obligations in my job and shortage of funds.

The data achieved in these experiments clearly depicted the presence of bioactive properties or vector control potentials in the test plants- *E. nummularius*, *L. camara*, *M. piperita*, *Mi. pudica*, *P. hysterophorus*, *Ph. niruri*, *Po. hydropiper*, *Pz. zeylanica*, *S. nodiflora* and *Z. zerumbet.* Since the test organisms were considered as vectors (because, they play role as vectors either for plant or animal diseases) the biological activities incorporated here as the vector control potentials, and everyone knows that the mosquito, *C. quinquefasciatus* and the egg-plant aphid, *A. gossypii* is are vectors,

however a very few people know that the flour beetle, *T. castaneum*, and the brine shrimp, *A. salina* are also vectors. The focal point of this investigation was to determine biological activities as control potentials of the test plants which were selected through survey of literature considering the published and the online information on them; and the magnitude of their activities were also established along with their comparative evaluation. It was rather interesting that some of them have been found promisingly active. Thus, comprehensive phytochemical analyses of the test plants for their insecticidal, insect repellent, cytotoxic and larvicidal components, as well as the physiological studies of the active ingredients are very much to be solicited for their effective use in the future vector control and pharmaceutical endeavors.
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APPENDICES

Regression equation : $Y = 3.097 + 1.041X$ Chi-squared is 3.957 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.828 mg/cm² LD_{50} is 0.673 mg/cm² 95% confidence limits are 0.080 to 5.631mg/ $cm²$

Regression equation : $Y = 2.639 + 1.933X$ Chi-squared is 2.767 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.221 mg/cm² LD_{50} is 0.167mg/cm² 95% confidence limits are 0.117 to 0.236 mq/cm²

Appendix Table III: Dose mortality effect of *E. nummularius* (wp/PetE) extracts against *A. gossypii* after 9h of exposure

95% confidence limits are 0.092 to 0.156mg/ cm^2

Appendix Table IV: Dose mortality effect of *E. nummularius* (wp/PetE) extracts against *A. gossypii* after 12h of exposure

Dose	Ldos $(+2)$					#U Kl %Kill Cr% E Pr Ex Pr Wk Pro		Weaht F Pro
0.196 0.147 0.098 0.049 0.025	1.292 1.167 0.991 0.690 0.398	30 30 30 30	30 22 73.333 19 63.333 16 53.333 9 30.000 5 16.667	73 5.61 63 5.33 53 5.08 30 4.48 17 4.05	5.581 5.363 5.056 4.530 4.020	5.584 5.075 4.460	5.318 18.48 5.347 19.11 17.43 4.037 13.17	17.43 5.566 5.039 4.513 4.002
Regression equation : Y = 3.306 + 1.749X								

Chi-squared is 0.111 with 3 degrees of freedom No significant heterogeneity Log LD_{50} IS 0.969mg/cm² LD_{50} is 0.093mg/cm² 95% confidence limits are 0.070 to 0.123 mq/cm²

Regression equation: $Y = 3.553 + 1.720X$ Chi-squared is 0.618 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.841 mg/cm² LD_{50} is 0.069mg/cm² 95% confidence limits are 0.052 to 0.093 mq/cm²

Appendix Table VI: Dose mortality effect of *E. nummularius* (wp/PetE) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 3.861 + 1.776X$ Chi-squared is 0.546 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.641 mg/cm² LD_{50} is 0.044 mg/cm² 95% confidence limits are 0.0309 to 0.062 mg/cm²

Appendix Table VII: Dose mortality effect of *E. nummularius* (wp/PetE) extracts against *A. gossypii* after 21h of exposure

Regression equation: $Y = 4.008 + 1.810X$ Chi-squared is 0.237 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.548 mg/cm² LD_{50} is 0.035mg/cm² 95% confidence limits are 0.024 to 0.052 mg/cm²

Appendix Table VIII: Dose mortality effect of *E. nummularius* (wp/PetE) extracts against *A. gossypii* after 24h of exposure

Regression equation: $Y = 3.999 + 1.884X$ Chi-squared is 1.646 with 3 degrees of freedom No significant heterogeneity Log LD_{50} IS 0.531mg/cm² LD_{50} is 0.034 mg/cm² 95% confidence limits are 0.023 to 0.050Smg/cm²

Regression equation: $Y = -0.835 + 4.240X$ Chi-squared is 1.003 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 1.376mg/cm² LD_{50} is 0.238 mg/cm² 95% confidence limits are 0.171 to 0.331 mq/cm²

Appendix Table X: Dose mortality effect of *E. nummularius* (wp/CHCl₃) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = -1.543 + 5.006X$ Chi-squared is 0.454 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 1.307 mg/cm² LD_{50} is 0.203mg/cm² 95% confidence limits are 0.166 to 0.248mg/cm²

Appendix Table XI: Dose mortality effect of *E. nummularius* (wp/CHCl₃) extracts against *A. gossypii* after 9h of exposure

Regression equation: $Y = -1.482 + 5.142X$ Chi-squared is 0.597 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 1.261 mg/cm² LD_{50} is 0.182 mg/cm² 95% confidence limits are 0.156 to 0.213 mg/cm²

Appendix Table XII: Dose mortality effect of *E. nummularius* (wp/CHCl3) extracts against *A. gossypii* after 12h of exposure

Regression equation: $Y = -1.561 + 5.451X$ Chi-squared is 0.057 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 1.203mg/cm² LD_{50} is 0.160 mg/cm² 95% confidence limits are 0.141 to 0.181mg/cm²

Regression equation: $Y = 0.810 + 3.695X$ Chi-squared is 8.790 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.134 mg/cm² LD_{50} is 0.136mq/cm² 95% confidence limits are 0.103 to 0.179mg/cm²

Appendix Table XIV: Dose mortality effect of *E. nummularius* (wp/CHCl3) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 1.976 + 2.899X$ Chi-squared is 12.927 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.043mg/cm² LD_{50} is 0.110 mq/cm² 95% confidence limits are 0.075 to 0.163 mq/cm²

Appendix Table XV: Dose mortality effect of *E. nummularius* (wp/CHCl3) extracts against *A. gossypii* after 21h of exposure

Regression equation: $Y = 2.076 + 3.027X$ Chi-squared is 5.485 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.966 mg/cm² LD_{50} is 0.093 mg/cm² 95% confidence limits are 0.077 to 0.111mg/cm²

Appendix Table XVI: Dose mortality effect of *E. nummularius* (wp/CHCl3) extracts against *A. gossypii* after 24h of exposure

Regression equation: $Y = 2.338 + 2.984X$ Chi-squared is 5.788 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.892 mg/cm² LD_{50} is 0.078 mg/cm² 95% confidence limits are 0.065 to 0.094mg/cm²

Regression equation: $Y = 2.817 + 1.313X$ Chi-squared is 2.639 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.663mg/cm² LD_{50} is 0.461mq/cm² 95% confidence limits are 0.158 to 1.340mq/cm²

Appendix Table XVIII: Dose mortality effect of *E. nummularius* (wp/CH₃OH) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 2.850 + 1.513X$ Chi-squared is 2.313 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.421 mg/cm² LD_{50} is 0.264mg/cm² 95% confidence limits are 0.142 to 0.491 mq/cm²

Appendix Table XIX: Dose mortality effect of *E. nummularius* (wp/CH₃OH) extracts against *A. gossypii* after 9h of exposure

Regression equation: $Y = 2.853 + 1.713X$ Chi-squared is 3.859 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.253mg/cm² LD_{50} is 0.179 mg/cm² 95% confidence limits are 0.120 to 0.268mg/cm²

Appendix Table XX: Dose mortality effect of *E. nummularius* (wp/CH₃OH) extracts against *A. gossypii* after 12h of exposure

Regression equation: $Y = 2.913 + 1.768X$ Chi-squared is 2.140 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.180 mg/cm² LD_{50} is 0.151 mg/cm² 95% confidence limits are 0.107 to 0.214mg/cm²

Regression equation: $Y = 2.988 + 1.890X$ Chi-squared is 0.619 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.064mg/cm² LD_{50} is 0.116mq/cm² 95% confidence limits are 0.088 to 0.153mg/cm²

Appendix Table XXII: Dose mortality effect of *E. nummularius* (wp/CH3OH) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 3.182 + 1.935X$ Chi-squared is 2.015 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.940 mg/cm² LD_{50} is 0.087mq/cm² 95% confidence limits are 0.067 to 0.112mg/cm²

Appendix Table XXIII: Dose mortality effect of *E. nummularius* (wp/CH₃OH) extracts against *A. gossypii* after 21h of exposure

Regression equation: $Y = 3.272 + 2.045X$ Chi-squared is 1.593 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.845 mg/cm² LD_{50} is 0.070mg/cm² 95% confidence limits are 0.054 to 0.090mg/cm²

Appendix Table XXIV: Dose mortality effect of *E. nummularius* (wp/CH₃OH) extracts against *A. gossypii* after 24h of exposure

Regression equation: $Y = 3.397 + 2.164X$ Chi-squared is 2.507 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.741mg/cm² LD_{50} is 0.055mg/cm² 95% confidence limits are 0.042 to 0.071mg/cm²

Regression equation: $Y = 1.771 + 2.563X$ Chi-squared is 6.286 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.260mg/cm² LD_{50} is 0.182 mg/cm² 95% confidence limits are 0.137 to 0.242 mg/cm²

Appendix Table XXVI: Dose mortality effect of *L. camara* (ap/PetE) extracts against *A. gossypii* after 12h of exposure

Regression equation: $Y = 2.223 + 2.335X$ Chi-squared is 7.946 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.190mg/cm² LD_{50} is 0.155 mg/cm² 95% confidence limits are 0.100 to 0.239 mg/cm²

Appendix Table XXVII: Dose mortality effect of *L. camara* (ap/PetE) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 2.564 + 2.154X$ Chi-squared is 6.031 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.131 mg/cm² LD_{50} is 0.135mg/cm² 95% confidence limits are 0.104 to 0.176 mg/cm²

Appendix Table XXVIII: Dose mortality effect of *L. camara* (ap/PetE) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 3.093 + 1.960X$ Chi-squared is 10.268 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 0.973 mg/cm² LD_{50} is 0.094mg/cm² 95% confidence limits are 0.058 to 0.151mg/cm²

Appendices VII IES, RU

Regression equation: $Y = 3.245 + 1.911X$ Chi-squared is 10.239 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 0.919 mg/cm² LD_{50} is 0.083mg/cm² 95% confidence limits are 0.051 to 0.134 mg/cm²

Appendix Table XXXI: Dose mortality effect of *L. camara* (ap/CHCl₃) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = 1.505 + 2.712X$ Chi-squared is 0.393 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 1.289 mg/cm² LD_{50} is 0.194 mg/cm² 95% confidence limits are 0.139 to 0.271 mg/cm²

Appendix Table XXX: Dose mortality effect of *L. camara* (ap/PetE) extracts against *A. gossypii* after 24h of exposure

Regression equation: $Y = 3.833 + 1.295X$ Chi-squared is 1.256 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.901mg/cm² LD_{50} is 0.080 mg/cm² 95% confidence limits are 0.052 to $0.122mg/cm²$

Appendix Table XXXII: Dose mortality effect of *L. camara* (ap/CHCl₃) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 1.350 + 3.065X$ Chi-squared is 0.005 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 1.191 mg/cm² LD_{50} is 0.155 mg/cm² 95% confidence limits are 0.126 to 0.191mg/cm²

Appendices VIII IES, RU

Regression equation: $Y = 1.638 + 2.917X$ Chi-squared is 1.021 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.153 mg/cm² LD_{50} is 0.142 mg/cm² 95% confidence limits are 0.116 to 0.174 mg/cm²

Appendix Table XXXIV: Dose mortality effect of *L. camara* (ap/CHCl₃) extracts against *A. gossypii* after 12h of exposure

Regression equation: $Y = 1.824 + 2.904X$ Chi-squared is 0.255 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.094 mg/cm² LD_{50} is 0.124 mg/cm² 95% confidence limits are 0.102 to 0.150mg/cm²

Appendix Table XXXV: Dose mortality effect of *L. camara* (ap/CHCl3) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 1.753 + 3.112X$ Chi-squared is 0.846 with 3 degrees of freedom No significant heterogeneity log LD_{50} is 1.043mg/cm² LD_{50} is 0.110 mq/cm² 95% confidence limits are 0.093 to 0.132mg/cm²

Appendix Table XXXVI: Dose mortality effect of *L. camara* (ap/CHCl₃) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 2.459 + 2.556X$ Chi-squared is 0.825 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.994mg/cm² LD_{50} is 0.099mg/cm² 95% confidence limits are 0.080 to 0.121mg/cm²

Regression equation: $Y = 2.467 + 2.702X$ Chi-squared is 2.351 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.937 mg/cm² LD_{50} is 0.087mg/cm² 95% confidence limits are 0.071 to 0.105mg/cm²

Appendix Table XXXIX: Dose mortality effect of *L. camara* (ap/CH₃OH) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = -2.597 + 5.348X$ Chi-squared is 2.679 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.421 mg/cm² LD_{50} is 0.263mg/cm² 95% confidence limits are 0.217 to 0.319mg/cm²

Regression equation: $Y = 3.017 + 2.141X$ Chi-squared is 0.021 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.926 mg/cm² LD_{50} is 0.084 mg/cm² 95% confidence limits are 0.064 to 0.111 mg/cm²

Appendix Table XL: Dose mortality effect of *L. camara* (ap/CH₃OH) extracts against *A. gossypii* after 6h of exposure

Dose	Ldos $(+2)$				#U Kl %Kill Cr% E Pr Ex Pr Wk Pro Weght F Pro		
					0.246 1.391 30 16 53.333 53 5.08 4.932 5.065 19.02 4.907		
					0.196 1.292 30 6 20.000 20 4.16 4.380 4.170 15.96 4.368		
0.147	1.167 30 2 6.667 7 3.52 3.680					3.529 9.06 3.685	
0.098					0.991 30 1 3.333 3 3.12 2.693 3.568 1.86 2.723		

Regression equation: $Y = -2.693 + 5.464X$ Chi-squared is 2.649 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.408mg/cm² LD_{50} is 0.256mg/cm² 95% confidence limits are 0.214 to 0.306mg/cm²

Appendices X IES, RU

Appendix Table XLIII: Dose mortality effect of *L. camara* (ap/CH₃OH) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 0.024 + 3.938X$ Chi-squared is 1.712 with 2 degrees of freedom No significant heterogeneity log LD_{50} is 1.264mg/cm² LD_{50} is 0.183 mg/cm² 95% confidence limits are 0.159 to $0.212mg/cm²$

Regression equation: $Y = -0.484 + 4.203X$ Chi-squared is 1.246 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.305 mg/cm² LD_{50} is 0.202 mg/cm² 95% confidence limits are 0.174 to 0.234 mg/cm²

Appendix Table XLIV: Dose mortality effect of *L. camara* (ap/CH₃OH) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 0.830 + 3.518X$ Chi-squared is 5.707 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.186 mg/cm² LD_{50} is 0.153 mg/cm² 95% confidence limits are 0.132 to $0.178mg/cm²$

Regression equation: $Y = 0.903 + 3.621X$ Chi-squared is 8.777 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.131 mg/cm² LD_{50} is 0.135mg/cm² 95% confidence limits are 0.106 to 0.173 mq/cm²

Appendix Table XLVII: Dose mortality effect of *L. camara* (r/PetE) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = 2.100 + 2.158X$ Chi-squared is 0.829 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.344mg/cm² LD_{50} is 0.221 mg/cm² 95% confidence limits are 0.143 to 0.340 mg/cm²

Regression equation: $Y = 1.984 + 2.719X$ Chi-squared is 0.099 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.110 mg/cm² LD_{50} is 0.129 mg/cm² 95% confidence limits are 0.104 to 0.159 mg/cm²

Appendix Table XLVIII: Dose mortality effect of *L. camara* (r/PetE) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 2.106 + 2.362X$ Chi-squared is 0.033 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.225 mg/cm² LD_{50} is 0.168 mq/cm² 95% confidence limits are 0.126 to 0.224 mg/cm²

Appendices XII IES, RU

Appendix Table XLIX: Dose mortality effect of *L. camara* (r/PetE) extracts against *A. gossypii* after 9h of exposure

Regression equation: $Y = 3.071 + 1.632X$ Chi-squared is 1.493 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.182 mg/cm² LD_{50} is 0.152 mg/cm² 95% confidence limits are 0.105 to 0.221 mq/cm²

Appendix Table LI: Dose mortality effect of *L. camara* (r/PetE) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 2.996 + 2.026X$ Chi-squared is 2.157 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.989 mg/cm² LD_{50} is 0.098mg/cm² 95% confidence limits are 0.076 to 0.125 mg/cm²

Appendix Table L: Dose mortality effect of *L. camara* (r/PetE) extracts against *A. gossypii* after 12h of exposure

Regression equation: $Y = 3.042 + 1.815X$ Chi-squared is 0.894 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.079 mg/cm² LD_{50} is 0.120mg/cm² 95% confidence limits are 0.089 to 0.161 mg/cm²

Appendix Table LII: Dose mortality effect of *L. camara* (r/PetE) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 3.059 + 2.228X$ Chi-squared is 3.642 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.871 mg/cm² LD_{50} is 0.074mg/cm² 95% confidence limits are 0.059 to 0.094mg/cm²

Appendices XIII IES, RU

Appendix Table LIII: Dose mortality effect of *L. camara* (r/PetE) extracts against *A. gossypii* after 21h of exposure

Regression equation: $Y = 3.187 + 2.245X$ Chi-squared is 6.271 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.807 mg/cm² LD_{50} is 0.064mg/cm² 95% confidence limits are 0.051 to 0.081 mq/cm²

Appendix Table LV: Dose mortality effect of *L. camara* (r/CHCl3) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = 1.468 + 2.928X$ Chi-squared is 0.067 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 1.206mg/cm² LD_{50} is 0.161 mg/cm² 95% confidence limits are 0.128 to 0.202 mg/cm²

Appendix Table LIV: Dose mortality effect of *L. camara* (r/PetE) extracts against *A. gossypii* after 24h of exposure

Regression equation: $Y = 3.783 + 1.689X$ Chi-squared is 1.362 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.720 mg/cm² LD_{50} is 0.053 mg/cm² 95% confidence limits are 0.038 to 0.073mg/cm²

Appendix Table LVI: Dose mortality effect of *L. camara* (r/CHCl3) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 0.975 + 3.419X$ Chi-squared is 1.454 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.177 mg/cm² LD_{50} is 0.150 mg/cm² 95% confidence limits are 0.125 to 0.180 mg/cm²

Appendix Table LVII: Dose mortality effect of *L. camara* (r/CHCl3) extracts against *A. gossypii* after 9h of exposure

Regression equation: $Y = 1.750 + 2.780X$ Chi-squared is 1.719 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.169 mg/cm² LD_{50} is 0.148 mg/cm² 95% confidence limits are 0.119 to 0.184 mg/cm²

Appendix Table LIX: Dose mortality effect of *L. camara* (r/CHCl₃) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 2.225 + 2.574X$ Chi-squared is 4.133 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.078 mg/cm² LD_{50} is 0.120 mq/cm² 95% confidence limits are 0.097 to 0.148mg/cm²

Appendix Table LVIII: Dose mortality effect of *L. camara* (r/CHCl₃) extracts against *A. gossypii* after 12h of exposure

Regression equation: $Y = 2.155 + 2.575X$ Chi-squared is 4.600 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.105 mg/cm² LD_{50} is 0.127 mg/cm² 95% confidence limits are 0.103 to 0.158mg/ $cm²$

Appendix Table LX: Dose mortality effect of *L. camara* (r/CHCl₃) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 2.795 + 2.217X$ Chi-squared is 2.810 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.995 mg/cm² LD_{50} is 0.099mg/cm² 95% confidence limits are 0.079 to 0.124 mg/cm²

Appendices XV IES, RU

Appendix Table LXI: Dose mortality effect of *L. camara* (r/CHCl₃) extracts against *A. gossypii* after 21h of exposure

Regression equation: $Y = 2.481 + 2.675X$ Chi-squared is 9.499 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 0.941 mg/cm² LD_{50} is 0.087mg/cm² 95% confidence limits are 0.062 to 0.124 mg/cm²

Appendix Table LXIII: Dose mortality effect of *L. camara* (r/CH₃OH) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = 2.813 + 3.948X$ Chi-squared is 0.150 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.554mg/cm² LD_{50} is 0.358 mg/cm² 95% confidence limits are 0.200 to 0.641 mg/cm²

Appendix Table LXII: Dose mortality effect of *L. camara* (r/CHCl₃) extracts against *A. gossypii* after 24h of exposure

Regression equation: $Y = 3.101 + 1.966X$ Chi-squared is 1.226 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.966mg/cm² LD_{50} is 0.092mg/cm² 95% confidence limits are 0.068 to 0.125mg/cm²

Appendix Table LXIV: Dose mortality effect of *L. camara* (r/CH₃OH) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 2.976 + 4.229X$ Chi-squared is 0.093 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.478 mg/cm² LD_{50} is 0.301mg/cm² 95% confidence limits are 0.208 to 0.435 mg/cm²

Appendix Table LXV: Dose mortality effect of *L. camara* (r/CH₃OH) extracts against *A. gossypii* after 9h of exposure

Regression equation: $Y = 1.635 + 2.257X$ Chi-squared is 4.544 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.491 mg/cm² LD_{50} is 0.310mg/cm² 95% confidence limits are 0.199 to $0.482mg/cm²$

Appendix Table LXVII: Dose mortality effect of *L. camara* (r/CH₃OH) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 1.707 + 2.541X$ Chi-squared is 5.564 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.296mg/cm² LD_{50} is 0.198 mg/cm² 95% confidence limits are 0.157 to 0.250mg/cm²

Appendix Table LXVI: Dose mortality effect of *L. camara* (r/CH₃OH) extracts against *A. gossypii* after 12h of exposure

Regression equation: $Y = 1.428 + 2.630X$ Chi-squared is 6.954 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.358 mg/cm² LD_{50} is 0.228 mg/cm² 95% confidence limits are 0.175 to 0.296 mg/cm²

Appendix Table LXVIII: Dose mortality effect of *L. camara* (r/CH₃OH) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 2.136 + 2.384X$ Chi-squared is 6.204 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.201 mg/cm² LD_{50} is 0.159mg/cm² 95% confidence limits are 0.129 to 0.197 mg/cm²

Appendices XVII IES, RU

Appendix Table LXIX: Dose mortality effect of *L. camara* (r/CH₃OH) extracts against *A. gossypii* after 21h of exposure

Regression equation: $Y = 1.898 + 2.744X$ Chi-squared is 8.418 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.130 mg/cm² LD_{50} is 0.135mg/cm² 95% confidence limits are 0.100 to 0.183 mq/cm²

Appendix Table LXXI: Dose mortality effect of *M. piperita* (wp/PetE) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = 1.395 + 2.628X$ Chi-squared is 0.110 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.372mg/cm² LD_{50} is 0.235mg/cm² 95% confidence limits are 0.157 to $0.352mg/cm^{2}$

Appendix Table LXX: Dose mortality effect of *L. camara* (r/CH3OH) extracts against *A. gossypii* after 24h of exposure

Regression equation: $Y = 2.991 + 1.796X$ Chi-squared is 2.049 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.118 mg/cm² LD_{50} is 0.131 mg/cm² 95% confidence limits are 0.096 to 0.179mg/cm²

Appendix Table LXXII: Dose mortality effect of *M. piperita* (wp/PetE) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 1.490 + 2.748X$ Chi-squared is 0.637 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.277 mg/cm² LD_{50} is 0.189 mg/cm² 95% confidence limits are 0.142 to 0.252 mg/cm²

Regression equation: $Y = 1.855 + 2.625X$ Chi-squared is 0.132 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.198mg/cm² LD_{50} is 0.158 mg/cm² 95% confidence limits are 0.123 to 0.202 mg/cm²

Appendix Table LXXIV: Dose mortality effect of *M. piperita* (wp/PetE) extracts against *A. gossypii* after 12h of exposure

Regression equation: $Y = 2.217 + 2.456X$ Chi-squared is 0.907 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.133 mg/cm² LD_{50} is 0.136mg/cm² 95% confidence limits are 0.107 to 0.172 mg/cm²

Appendix Table LXXV: Dose mortality effect of *M. piperita* (wp/PetE) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 2.545 + 2.346X$ Chi-squared is 0.362 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.047 mg/cm² LD_{50} is 0.111 mq/cm² 95% confidence limits are 0.089 to 0.139mg/cm²

Appendix Table LXXVI: Dose mortality effect of *M. piperita* (wp/PetE) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 2.738 + 2.338X$ Chi-squared is 2.122 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.968 mg/cm² LD_{50} is 0.093mg/cm² 95% confidence limits are 0.075 to 0.115mg/cm²

Regression equation: $Y = 2.610 + 2.596X$ Chi-squared is 2.351 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.920 mg/cm² LD_{50} is 0.083mg/cm² 95% confidence limits are 0.068 to 0.102 mg/cm²

Appendix Table LXXIX: Dose mortality effect of *M. piperita* (wp/CHCl₃) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = 3.433 + 3.900X$ Chi-squared is 0.186 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.402 mg/cm² LD_{50} is 0.252 mg/cm² 95% confidence limits are 0.196 to 0.325 mg/cm²

Appendix Table LXXVIII: Dose mortality effect of *M. piperita* (wp/PetE) extracts against *A. gossypii* after 24h of exposure

Regression equation: $Y = 3.169 + 2.173X$ Chi-squared is 0.835 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.843 mg/cm² LD_{50} is 0.070mg/cm² 95% confidence limits are 0.054 to 0.090 mg/cm²

Appendix Table LXXX: Dose mortality effect of *M. piperita* (wp/CHCl₃) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = -3.749 + 6.751X$ Chi-squared is 1.669 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.296mg/cm² LD_{50} is 0.198 mg/cm² 95% confidence limits are 0.180 to 0.217mg/cm²

Appendices XX IES, RU

Regression equation: $Y = 1.262 + 2.929X$ Chi-squared is 8.881 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.276mg/cm² LD_{50} is 0.189mg/cm² 95% confidence limits are 0.135 to 0.264 mg/cm²

Appendix Table LXXXIII: Dose mortality effect of *M. piperita* (wp/CHCl₃) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 2.194 + 2.517X$ Chi-squared is 6.290 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.115 mg/cm² LD_{50} is 0.130 mq/cm² 95% confidence limits are 0.107 to 0.159mg/cm²

Appendix Table LXXXII: Dose mortality effect of *M. piperita* (wp/CHCl₃) extracts against *A. gossypii* after 12h of exposure

Regression equation: $Y = 2.058 + 2.466X$ Chi-squared is 8.146 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.193 mg/cm² LD_{50} is 0.156mg/cm² 95% confidence limits are 0.111 to 0.219mg/cm²

Appendix Table LXXXIV: Dose mortality effect of *M. piperita* (wp/CHCl₃) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 2.344 + 2.576X$ Chi-squared is 6.629 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.031 mg/cm² LD_{50} is 0.107mg/cm² 95% confidence limits are 0.088 to $0.132mg/cm²$

Appendices XXI IES, RU

Regression equation: $Y = 2.469 + 2.500X$ Chi-squared is 2.975 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.013mg/cm² LD_{50} is 0.103 mg/cm² 95% confidence limits are 0.083 to 0.128mg/cm²

Appendix Table LXXXVII: Dose mortality effect of *M. piperita* (wp/CH₃OH) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = 2.851 + 3.495X$ Chi-squared is 0.398 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.615 mg/cm² LD_{50} is 0.412 mg/cm² 95% confidence limits are 0.178 to 0.953 mg/cm²

Appendix Table LXXXVI: Dose mortality effect of *M. piperita* (wp/CHCl₃) extracts against *A. gossypii* after 24h of exposure

Regression equation: $Y = 2.511 + 2.541X$ Chi-squared is 1.213 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.980mg/cm² LD_{50} is 0.095mg/cm² 95% confidence limits are 0.076 to 0.120mg/cm²

Appendix Table LXXXVIII: Dose mortality effect of *M. piperita* (wp/CH₃OH) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 2.715 + 4.389X$ Chi-squared is 0.273 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.521 mg/cm² LD_{50} is 0.332mg/cm² 95% confidence limits are 0.211 to 0.521 mq/cm²

Regression equation: $Y = 0.983 + 2.612X$ Chi-squared is 2.170 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.539mg/cm² LD_{50} is 0.345mg/cm² 95% confidence limits are 0.216 to 0.551 mq/cm²

Appendix Table XCI: Dose mortality effect of *M. piperita* (wp/CH₃OH) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 1.678 + 2.355X$ Chi-squared is 0.496 with 3 degrees of freedom No significant heterogeneity Log LD_{50} IS 1.411 mg/cm² LD_{50} is 0.258mg/cm² 95% confidence limits are 0.184 to 0.360mg/cm²

Appendix Table XC: Dose mortality effect of *M. piperita* (wp/CH₃OH) extracts against *A. gossypii* after12h of exposure

Regression equation: $Y = 1.344 + 2.411X$ Chi-squared is 0.253 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.516 mg/cm² LD_{50} is 0.328 mg/cm² 95% confidence limits are 0.209 to 0.515 mq/cm²

Appendix Table XCII: Dose mortality effect of *M. piperita* (wp/CH₃OH) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 2.025 + 2.258X$ Chi-squared is 1.123 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.318 mg/cm² LD_{50} is 0.208mg/cm² 95% confidence limits are 0.158 to 0.273 mg/cm²

Appendices XXIII IES, RU

Regression equation: $Y = 1.671 + 2.790X$ Chi-squared is 4.089 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.193 mg/cm² LD_{50} IS 0.156 mg/cm² 95% confidence limits are 0.130 to 0.187mq/cm²

Appendix Table XCIV: Dose mortality effect of *M. piperita* (wp/CH₃OH) extracts against *A. gossypii* after 24h of exposure

Regression equation: $Y = 1.760 + 2.940X$ Chi-squared is 7.945 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.102 mg/cm² LD_{50} is 0.126 mg/cm² 95% confidence limits are 0.095 to 0.168 mq/cm²

Appendix Table XCV: Dose mortality effect of *Mi. pudica* (wp/PetE) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = 3.526 + 2.701X$ Chi-squared is 0.519 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.546 mg/cm² LD_{50} is 0.351 mq/cm² 95% confidence limits are 0.168 to 0.735 mg/cm²

Appendix Table XCVI: Dose mortality effect of *Mi. pudica* (wp/PetE) extracts against A. gossypii after 6h of exposure

Regression equation: $Y = 3.527 + 2.796X$ Chi-squared is 0.148 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.527 mg/cm² LD_{50} is 0.336mg/cm² 95% confidence limits are 0.176 to $0.644mg/cm²$

Regression equation: $Y = 0.632 + 3.058X$ Chi-squared is 1.754 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.428 mg/cm² LD_{50} is 0.268mg/cm² 95% confidence limits are 0.200 to 0.359mg/cm²

Appendix Table XCIX: Dose mortality effect of *Mi. pudica* (wp/PetE) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 0.915 + 3.163X$ Chi-squared is 4.535 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.291 mg/cm² LD_{50} is 0.196mq/cm² 95% confidence limits are 0.162 to 0.236 mg/cm²

Appendix Table XCVIII: Dose mortality effect of *Mi. pudica* (wp/PetE) extracts against *A. gossypii* after 12h of exposure

Regression equation: $Y = 1.390 + 2.626X$ Chi-squared is 3.200 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.375 mg/cm² LD_{50} is 0.237mg/cm² 95% confidence limits are 0.180 to 0.313mg/ $cm²$

Appendix Table C: Dose mortality effect of *Mi. pudica* (wp/PetE) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 0.825 + 3.486X$ Chi-squared is 2.534 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.198 mg/cm² LD_{50} is 0.158 mg/cm² 95% confidence limits are 0.136 to 0.183 mg/cm²

Appendices XXV IES, RU

Appendix Table CI: Dose mortality effect of *Mi. pudica* (wp/PetE) extracts against *A. gossypii* after 21h of exposure ──────────────────────────────────────────────────────────────────────

Regression equation: $Y = 0.147 + 4.212X$ Chi-squared is 7.715 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.152 mg/cm² LD_{50} is 0.142 mg/cm² 95% confidence limits are 0.125 to $0.162mg/cm^2$

Appendix Table CIII: Dose mortality effect of *Mi. pudica* (wp/CHCl₃) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = 0.769 + 2.832X$ Chi-squared is 0.147 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 1.494mg/cm² LD_{50} is 0.312 mg/cm² 95% confidence limits are 0.148 to 0.655 mg/cm²

Appendix Table CII: Dose mortality effect of *Mi. pudica* (wp/PetE) extracts against *A. gossypii* after 24h of exposure

Regression equation: $Y = 1.764 + 2.873X$ Chi-squared is 0.869 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.126 mg/cm² LD_{50} is 0.134 mg/cm² 95% confidence limits are 0.110 to 0.163 mg/cm²

Appendix Table CIV: Dose mortality effect of *Mi. pudica* (wp/CHCl₃) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 2.353 + 1.730X$ Chi-squared is 1.413 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.530mg/cm² LD_{50} is 0.339mg/cm² 95% confidence limits are 0.146 to 0.784 mq/cm²

Regression equation: $Y = 2.044 + 2.240X$ Chi-squared is 0.888 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.320mg/cm² LD_{50} is 0.209mg/cm² 95% confidence limits are 0.142 to 0.308 mq/cm²

Appendix Table CVI: Dose mortality effect of *Mi. pudica* (wp/CHCl₃) extracts against *A. gossypii* after 12h of exposure

Regression equation: $Y = 2.296 + 2.225X$ Chi-squared is 2.121 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.215 mg/cm² LD_{50} is 0.164mg/cm² 95% confidence limits are 0.122 to 0.220mg/cm²

Appendix Table CVII: Dose mortality effect of *Mi. pudica* (wp/CHCl₃) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 2.286 + 2.431X$ Chi-squared is 1.944 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.116mg/cm² LD_{50} is 0.131 mg/cm² 95% confidence limits are 0.104 to 0.165mg/cm²

Appendix Table CVIII: Dose mortality effect of *Mi. pudica* (wp/CHCl₃) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 2.712 + 2.323X$ Chi-squared is 1.130 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.985mg/cm² LD_{50} is 0.097mg/cm² 95% confidence limits are 0.077 to 0.121mg/cm²

Regression equation: $Y = 2.464 + 2.816X$ Chi-squared is 1.975 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.901 mg/cm² LD_{50} is 0.080mg/cm² 95% confidence limits are 0.066 to 0.096 mg/cm²

Appendix Table CXI: Dose mortality effect of *Mi. pudica* (wp/CH₃OH) extracts against *A. gossypii* after 9h of exposure

Regression equation: $Y = -0.876 + 4.529X$ Chi-squared is 5.719 with 1 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.297 mg/cm² LD_{50} is 0.198 mg/cm² 95% confidence limits are 0.120 to 0.329 mg/cm²

Appendix Table CX: Dose mortality effect of *Mi. pudica* (wp/CHCl3) extracts against *A. gossypii* after 24h of exposure

Regression equation: $Y = 2.922 + 2.437X$ Chi-squared is 0.298 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.853 mg/cm² LD_{50} is 0.071mg/cm² 95% confidence limits are 0.056 to 0.090mg/cm²

Appendix Table CXII: Dose mortality effect of Mi. pudica (wp/CH₃OH) extracts against *A. gossypii* after 12h of exposure

Regression equation: $Y = 2.169 + 2.070X$ Chi-squared is 6.508 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.368mg/cm² LD_{50} is 0.233 mg/cm² 95% confidence limits are 0.151 to $0.361mg/cm²$

Appendices XXVIII IES, RU

Regression equation: $Y = 2.183 + 2.228X$ Chi-squared is 8.290 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.265mg/cm² LD_{50} is 0.184 mg/cm² 95% confidence limits are 0.107 to 0.315mg/cm²

Appendix Table CXIV: Dose mortality effect of *Mi. pudica* (wp/CH₃OH) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 1.818 + 2.847X$ Chi-squared is 7.662 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.118 mg/cm² LD_{50} is 0.131 mg/cm² 95% confidence limits are 0.107 to 0.160mg/cm²

Appendix Table CXV: Dose mortality effect of *Mi. pudica* (wp/CH₃OH) extracts against *A. gossypii* after 21h of exposure

Regression equation: $Y = 2.616 + 2.363X$ Chi-squared is 12.530 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.009mg/cm² LD_{50} is 0.102 mg/cm² 95% confidence limits are 0.065 to 0.159 mg/cm²

Appendix Table CXVI: Dose mortality effect of Mi. pudica (wp/CH₃OH) extracts against *A. gossypii* after 24h of exposure

Regression equation: $Y = 3.155 + 1.839X$ Chi-squared is 2.946 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.003mg/cm² LD_{50} is 0.101 mg/cm² 95% confidence limits are 0.072 to 0.142mg/cm²

Chi-squared is 1.714 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 1.334 mg/cm² LD_{50} is 0.216 mg/cm² 95% confidence limits are 0.168 to 0.277 mg/cm²

Appendix Table CXIX: Dose mortality effect of *P. hysterophorus* (wp/PetE) extracts against *A. gossypii* after 9h of exposure

Regression equation: $Y = 2.041 + 2.336$ X Chi-squared is 1.877 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.267mg/cm² LD_{50} is 0.185mg/cm² 95% confidence limits are 0.135 to $0.252mg/cm²$

Appendix Table CXVIII: Dose mortality effect of *P. hysterophorus* (wp/PetE) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 0.381 + 3.538X$ Chi-squared is 1.887 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 1.305 mg/cm² LD_{50} is 0.202 mg/cm² 95% confidence limits are 0.153 to 0.267 mg/cm²

Appendix Table CXX: Dose mortality effect of *P. hysterophorus* (wp/PetE) extracts against *A. gossypii* after 12h of exposure

Regression equation: $Y = 2.260 + 2.309X$ Chi-squared is 2.238 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.187 mg/cm² LD_{50} is 0.154 mg/cm² 95% confidence limits are 0.117 to 0.201 mg/cm²

Regression equation: $Y = 2.847 + 1.936X$ Chi-squared is 2.656 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is $1.112mg/cm^2$ LD_{50} is 0.130mg/cm² 95% confidence limits are 0.097 to 0.173 mq/cm²

Appendix Table CXXIII: Dose mortality effect of *P. hysterophorus* (wp/PetE) extracts against *A. gossypii* after 21h of exposure

Regression equation: $Y = 2.857 + 2.421$ X Chi-squared is 8.455 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 0.885 mg/cm² LD_{50} is 0.077mq/cm² 95% confidence limits are 0.054 to 0.110mg/cm²

Appendix Table CXXII: Dose mortality effect of *P. hysterophorus* (wp/PetE) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 2.460 + 2.614X$ Chi-squared is 5.653 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.972 mg/cm² LD_{50} is 0.094mg/cm² 95% confidence limits are 0.077 to 0.115mg/cm²

Appendix Table CXXIV: Dose mortality effect of *P. hysterophorus* (wp/PetE) extracts against *A. gossypii* after 24h of exposure

Regression equation: $Y = 3.565 + 1.723X$ Chi-squared is 0.900 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.833 mg/cm² LD_{50} is 0.068mg/cm² 95% confidence limits are 0.050 to $0.093mg/cm²$

Appendices XXXI IES, RU

Regression equation: $Y = 1.547 + 2.293X$ Chi-squared is 0.134 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.506mg/cm² LD_{50} is 0.321 mg/cm² 95% confidence limits are 0.166 to 0.618 mq/cm²

Regression equation: $Y = 2.213 + 1.839X$ Chi-squared is 0.228 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.516mg/cm² LD_{50} is 0.328mg/cm² 95% confidence limits are 0.152 to 0.708mg/cm²

Appendix Table CXXVII: Dose mortality effect of *P. hysterophorus* (wp/CHCl3) extracts against *A. gossypii* after 9h of exposure

Regression equation: $Y = 2.054 + 2.191X$ Chi-squared is 0.111 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.345mg/cm² LD_{50} is 0.221 mg/cm² 95% confidence limits are 0.144 to 0.339mg/cm²

Regression equation: $Y = 2.349 + 2.249X$ Chi-squared is 0.684 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.179 mg/cm² LD_{50} is 0.151 mg/cm² 95% confidence limits are 0.115 to $0.198mg/cm²$

Regression equation: $Y = 2.663 + 2.160X$ Chi-squared is 0.111 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.082 mg/cm² LD_{50} is 0.121 mg/cm² 95% confidence limits are 0.094 to 0.155mg/cm²

Appendix Table CXXXI: Dose mortality effect of *P. hysterophorus* (wp/CHCl3) extracts against *A. gossypii* after 21h of exposure

Regression equation: $Y = 3.061 + 2.103X$ Chi-squared is 3.498 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.922 mg/cm² LD_{50} is 0.084 mg/cm² 95% confidence limits are 0.066 to 0.106mg/cm²

Regression equation: $Y = 2.935 + 2.019X$ Chi-squared is 1.038 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.023 mg/cm² LD_{50} is 0.105mg/cm² 95% confidence limits are 0.082 to 0.136 mg/cm²

Appendix Table CXXXII: Dose mortality effect of *P. hysterophorus* (wp/CHCl3) extracts against *A. gossypii* after 24h of exposure

Regression equation: $Y = 3.180 + 2.092X$ Chi-squared is 0.959 with 2 degrees of freedom No significant heterogeneity LOG LD_{50} is 0.870 mg/cm² LD_{50} is 0.074mg/cm² 95% confidence limits are 0.057 to $0.097mg/cm²$

Appendices XXXIII IES, RU

Regression equation: $Y = -0.888 + 3.802X$ CHI-SQUARED IS 1.876 WITH 2 DEGREES OF FREEDOM NO SIG HETEROGENEITY LOG LD_{50} IS 1.549mg/cm² LD_{50} IS 0.354mg/cm² 95% CONF LIMITS ARE 0.223 to 0.561mg/cm²

Regression equation: $Y = 1.068 + 2.458X$ Chi-squared is 4.983 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.600 mg/cm² LD_{50} is 0.398mg/cm² 95% confidence limits are 0.222 to 0.713 mg/cm²

Appendix Table CXXXV: Dose mortality effect of *P. hysterophorus* (wp/CH₃OH) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 0.928 + 2.911X$ Chi-squared is 1.630 with 3 degrees of freedom No significant heterogeneity Log LD_{50} IS 1.399mg/cm² LD_{50} is 0.250mg/cm² 95% confidence limits are 0.191 to 0.329mg/cm²

Appendix Table CXXXVI: Dose mortality effect of *P. hysterophorus* (wp/CH₃OH) extracts against *A. gossypii* after 21h of exposure

Regression equation: $Y = 1.457 + 2.841X$ Chi-squared is 5.130 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is $1.247mg/cm²$ LD_{50} is 0.177 mg/cm² 95% confidence limits are 0.146 to 0.214 mg/cm²

Regression equation: $Y = 1.167 + 3.311X$ Chi-squared is 6.853 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.158 mg/cm² LD_{50} is 0.144 mg/cm² 95% confidence limits are 0.123 to 0.168mg/cm²

Appendix Table CXXXVIII: Dose mortality effect of *Ph. niruri* (wp/PetE) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = 1.805 + 2.069X$ Chi-squared is 4.907 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.544 mg/cm² LD_{50} is 0.350mg/cm² 95% confidence limits are 0.176 to 0.697 mq/cm²

Appendix Table CXXXIX: Dose mortality effect of *Ph. niruri* (wp/PetE) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 2.268 + 1.876X$ Chi-squared is 2.647 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.457mg/cm² LD_{50} is 0.286 mg/cm² 95% confidence limits are 0.161 to 0.509mg/cm²

Appendix Table CXL: Dose mortality effect of *Ph. niruri* (wp/PetE) extracts against *A. gossypii* after 9h of exposure

Regression equation: $Y = 2.841 + 1.578X$ Chi-squared is 3.324 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.368mg/cm² LD_{50} is 0.233 mg/cm² 95% confidence limits are 0.136 to 0.401mg/cm²

Appendix Table CXLI: Dose mortality effect of *Ph. niruri* (wp/PetE) extracts against *A. gossypii* after 12h of exposure

Regression equation: $Y = 2.692 + 1.907X$ Chi-squared is 2.183 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.210 mg/cm² LD_{50} is 0.162 mg/cm² 95% confidence limits are 0.116 to 0.227 mg/cm²

Appendix Table CXLII: Dose mortality effect of *Ph. niruri* (wp/PetE) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 3.030 + 1.767X$ Chi-squared is 2.314 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.115mg/cm² LD_{50} is 0.130 mg/cm² 95% confidence limits are 0.095 to 0.178mg/cm²

Appendix Table CXLIII: Dose mortality effect of *Ph. niruri* (wp/PetE) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 3.183 + 1.858X$ Chi-squared is 2.636 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.978 mg/cm² LD_{50} is 0.095mg/cm² 95% confidence limits are 0.073 to 0.124mg/cm²

Appendix Table CXLIV: Dose mortality effect of *Ph. niruri* (wp/PetE) extracts against against *A. gossypii* after 21h of exposure

Regression equation: $Y = 3.211 + 2.088X$ Chi-squared is 4.286 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.857 mg/cm² LD_{50} is 0.072 mg/cm² 95% confidence limits are 0.056 to 0.092mg/cm²

Regression equation: $Y = 3.460 + 1.981X$ Chi-squared is 1.258 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.778mg/cm² LD_{50} is 0.060mg/cm² 95% confidence limits are 0.045 to 0.079 mq/cm²

Appendix Table CXLVI: Dose mortality effect of *Ph. niruri* (wp/CHCl₃) extracts against *A. gossypii* after 9h of exposure

Regression equation: $Y = -1.847 + 10.824X$ Chi-squared is 0.217 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.633 mg/cm² LD_{50} is 0.429 mg/cm² 95% confidence limits are 0.373 to 0.494mg/cm²

Appendix Table CXLVII: Dose mortality effect of *Ph. niruri* (wp/CHCl₃) extracts against *A. gossypii* after 12h of exposure

Regression equation: $Y = 0.646 + 6.958X$ Chi-squared is 0.616 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.626 mg/cm² LD_{50} is 0.422 mg/cm² 95% confidence limits are 0.360 to 0.496mg/cm²

Appendix Table CXLVIII: Dose mortality effect of *Ph. niruri* (wp/CHCl₃) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 0.347 + 7.857X$ Chi-squared is 0.623 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.592 mg/cm² LD_{50} is 0.391 mg/cm² 95% confidence limits are 0.351 to 0.436mg/cm²

Regression equation: $Y = 0.906 + 7.309X$ Chi-squared is 0.430 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.560 mg/cm² LD_{50} is 0.363mg/cm² 95% confidence limits are 0.332 to 0.398mg/cm²

Appendix Table CLI: Dose mortality effect of *Ph. niruri* (wp/CHCl₃) extracts against *A. gossypii* after 24h of exposure

Regression equation: $Y = 2.480 + 5.318X$ Chi-squared is 3.356 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.474 mg/cm² LD_{50} is 0.298 mg/cm² 95% confidence limits are 0.272 to 0.327mg/cm²

Appendix Table CL: Dose mortality effect of *Ph. niruri* (wp/CHCl₃) extracts against *A. gossypii* after 21h of exposure

Regression equation: $Y = 1.900 + 5.828X$ Chi-squared is 0.762 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.532 mg/cm² LD_{50} is 0.340 mg/cm² 95% confidence limits are 0.308 to 0.376 mg/cm²

Appendix Table CLII: Dose mortality effect of *Ph. niruri* (wp/CH₃OH) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 3.330 + 2.374X$ Chi-squared is 0.003 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.703mg/cm² LD_{50} is 0.505 mg/cm² 95% confidence limits are 0.121 to 2.116mg/cm2

Appendices XXXVIII IES, RU

Regression equation: $Y = -0.244 + 3.451X$ Chi-squared is 0.669 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.520mg/cm² LD_{50} is 0.331mg/cm² 95% confidence limits are 0.217 to 0.505 mq/cm²

Regression equation: $Y = -1.438 + 4.593X$ Chi-squared is 2.098 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.402 mg/cm² LD_{50} is 0.252mg/cm² 95% confidence limits are 0.207 to 0.308mg/cm²

Appendix Table CLV: Dose mortality effect of *Ph. niruri* (wp/CH₃OH) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 1.108 + 2.841X$ Chi-squared is 5.444 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.370 mg/cm² LD_{50} is 0.234 mg/cm² 95% confidence limits are 0.182 to $0.302mg/cm²$

Appendix Table CLVI: Dose mortality effect of *Ph. niruri* (wp/CH₃OH) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 1.370 + 2.843X$ Chi-squared is 6.138 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.277 mg/cm² LD_{50} is 0.189 mg/cm² 95% confidence limits are 0.155 to 0.231mg/cm²

Regression equation: $Y = 1.331 + 2.958X$ Chi-squared is 4.954 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.240 mg/cm² LD_{50} is 0.174 mg/cm² 95% confidence limits are 0.145 to 0.209 mg/cm²

Appendix Table CLIX: Dose mortality effect of *Po. hydropiper* (wp/PetE) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = 1.111 + 2.598X$ Chi-squared is 0.848 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.497mg/cm² LD_{50} is 0.314 mg/cm² 95% confidence limits are 0.168 to 0.585mg/cm²

Regression equation: $Y = 1.291 + 3.191X$ Chi-squared is 6.750 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.163 mg/cm² LD_{50} is 0.145 mg/cm² 95% confidence limits are 0.124 to 0.171 mg/cm²

Appendix Table CLX: Dose mortality effect of *Po. hydropiper* (wp/PetE) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 0.361 + 3.565X$ Chi-squared is 1.107 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.301 mg/cm² LD_{50} is 0.200 mg/cm² 95% confidence limits are 0.156 mg/cm² to 0.256 mg/cm²

Appendices XL IES, RU

Regression equation: $Y = 0.505 + 3.720X$ Chi-squared is 0.800 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.208mg/cm² LD_{50} is 0.162 mg/cm² 95% confidence limits are 0.135 to 0.193 mq/cm²

Appendix Table CLXIII: Dose mortality effect of *Po. hydropiper* (wp/PetE) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 0.755 + 3.867X$ Chi-squared is 8.777 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.098 mg/cm² LD_{50} is 0.125 mg/cm² 95% confidence limits are 0.097 to $0.162mg/cm^2$

Appendix Table CLXII: Dose mortality effect of *Po. hydropiper* (wp/PetE) extracts against *A. gossypii* after 12h of exposure

Regression equation: $Y = 0.713 + 3.656X$ Chi-squared is 1.058 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is $1.172mg/cm^2$ LD_{50} is 0.149 mg/cm² 95% confidence limits are 0.125 to 0.176 mq/cm²

Appendix Table CLXIV: Dose mortality effect of *Po. hydropiper* (wp/PetE) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 1.424 + 3.478X$ Chi-squared is 3.287 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.028 mg/cm² LD_{50} is 0.107 mg/cm² 95% confidence limits are 0.091 to 0.125mg/cm²

Regression equation: $Y = 2.460 + 2.437X$ Chi-squared is 2.344 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.042 mg/cm² LD_{50} is 0.110 mg/cm² 95% confidence limits are 0.084 to 0.145 mq/cm²

Appendix Table CLXVI: Dose mortality effect of *Po. hydropiper* (wp/PetE) extracts against *A. gossypii* after 24h of exposure

Regression equation: $Y = 2.555 + 2.560X$ Chi-squared is 3.713 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.955mg/cm² LD_{50} is 0.090mg/cm² 95% confidence limits are 0.071 to 0.114 mg/cm²

Appendix Table CLXVII: Dose mortality effect of *Po. hydropiper* (wp/CHCl3) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = 0.230 + 3.139X$ Chi-squared is 2.405 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.520 mg/cm² LD_{50} is 0.331 mg/cm² 95% Confidence limits are 0.211 to 0.519mg/cm²

Appendix Table CLXVIII: Dose mortality effect of *Po. hydropiper* (wp/CHCl₃) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 0.297 + 3.449X$ Chi-squared is 1.019 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.363mg/cm² LD_{50} is 0.231 mg/cm² 95% confidence limits are 0.185 to 0.288mg/cm²

Regression equation: $Y = 0.706 + 3.392X$ Chi-squared is 1.948 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.266mg/cm² LD_{50} is 0.184 mg/cm² 95% confidence limits are 0.156 to 0.218 mq/cm²

Appendix Table CLXXI: Dose mortality effect of *Po. hydropiper* (wp/CHCl3) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 1.147 + 3.704X$ Chi-squared is 1.219 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.040 mg/cm² LD_{50} is 0.110 mg/cm² 95% confidence limits are 0.088 to 0.136mg/cm²

Appendix Table CLXX: Dose mortality effect of *Po. hydropiper* (wp/CHCl₃) extracts against *A. gossypii* after 12h of exposure

Regression equation: $Y = 1.392 + 3.190X$ Chi-squared is 0.791 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.131 mg/cm² LD_{50} is 0.135mg/cm² 95% confidence limits are 0.112 to 0.164 mq/cm²

Appendix Table CLXXII: Dose mortality effect of *Po. hydropiper* (wp/CHCl₃) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 1.768 + 3.393X$ Chi-squared is 0.768 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.952 mg/cm² LD_{50} is 0.090mg/cm² 95% confidence limits are 0.065 to 0.123mg/cm²

Regression equation: $Y = 2.214 + 3.056X$ Chi-squared is 1.057 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is $0.912mg/cm^2$ LD_{50} is 0.082mg/cm² 95% confidence limits are 0.055 to 0.122 mg/cm²

Appendix Table LXXV: Dose mortality effect of *Po. hydropiper* (wp/CH₃OH) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = 2.851 + 3.495X$ Chi-squared is 0.398 with 1 degrees of freedom No significant heterogeneity Log LD_{50} IS 0.615mg/cm² LD_{50} is 0.412 mg/cm² 95% confidence limits are 0.178 to 0.953 mq/cm²

Appendix Table CLXXIV: Dose mortality effect of *Po. hydropiper* (wp/CHCl₃) extracts against *A. gossypii* after 24h of exposure

Regression equation: $Y = 2.279 + 3.024X$ Chi-squared is 0.002 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.900mg/cm² LD_{50} is 0.079 mg/cm² 95% confidence limits are 0.052 to 0.121mg/cm²

Appendix Table CLXXVI: Dose mortality effect of *Po. hydropiper* (wp/CH₃OH) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 2.802 + 4.514X$ Chi-squared is 0.054 with 1 degrees of freedom No significant heterogeneity Log LD_{50} IS 0.487mg/cm² LD_{50} is 0.307 mg/cm² 95% confidence limits are 0.212 to 0.444 mg/cm²

Regression equation: $Y = 0.900 + 2.722X$ Chi-squared is 2.726 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.507mg/cm² LD_{50} is 0.321mq/cm² 95% confidence limits are 0.212 to 0.486 mg/cm²

Appendix Table CLXXIX: Dose mortality effect of *Po. hydropiper* (wp/CH₃OH) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 0.420 + 3.496X$ Chi-squared is 3.202 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.310mg/cm² LD_{50} IS 0.204 mg/cm² 95% confidence limits are 0.171 to 0.244 mg/cm²

Appendix Table CLXXVIII: Dose mortality effect of *Po. hydropiper* (wp/CH₃OH) extracts against *A. gossypii* after 12h of exposure

Regression equation: $Y = 0.606 + 3.139X$ Chi-squared is 3.224 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.400 mg/cm² LD_{50} is 0.251 mq/cm² 95% confidence limits are 0.194 to 0.325 mg/cm²

Appendix Table CLXXX: Dose mortality effect of *Po. hydropiper* (wp/CH₃OH) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 0.210 + 3.824X$ Chi-squared is 1.433 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.253 mg/cm² LD_{50} is 0.179 mg/cm² 95% confidence limits are 0.155 to 0.207 mg/cm²

Appendices XLV IES, RU

Regression equation: $Y = 0.497 + 3.775X$ Chi-squared is 4.502 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.193 mg/cm² LD_{50} is 0.156mq/cm² 95% confidence limits are 0.135 to 0.179mg/cm²

Appendix Table CLXXXIII: Dose mortality effect of *Pz. zeylanica* (wp/PetE) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = 3.060 + 4.858X$

Chi-squared is -0.00003 with 0 degrees of freedom No significant heterogeneity

Log LD_{50} is 0.399 mg/cm²

- LD_{50} is 0.251 mg/cm²
- 95% confidence limits are 0.154 to $0.407mg/cm²$

Regression equation: $Y = 1.139 + 3.461X$ Chi-squared is 4.159 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.115 mg/cm² LD_{50} is 0.130 mg/cm² 95% confidence limits are 0.112 to 0.152 mg/cm²

Appendix Table CLXXXIV: Dose mortality effect of *Pz. zeylanica* (wp/PetE) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 1.487 + 2.573X$ Chi-squared is 1.910 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.365mg/cm² LD_{50} is 0.232 mg/cm² 95% confidence limits are 0.156 to 0.345 mg/cm²

Appendices XLVI IES, RU

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Regression equation: $Y = 1.934 + 2.436X$ Chi-squared is 2.127 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.258mg/cm² LD_{50} is 0.181 mg/cm² 95% confidence limits are 0.135 to 0.243 mq/cm²

Appendix Table CLXXXVII: Dose mortality effect of *Pz. zeylanica* (wp/PetE) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 2.311 + 2.499X$ Chi-squared is 2.477 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.076 mg/cm² LD_{50} is 0.119 mg/cm² 95% confidence limits are 0.096 to 0.148mg/cm²

Appendix Table CLXXXVI: Dose mortality effect of *Pz. zeylanica* (wp/PetE) extracts against *A. gossypii* after 12h of exposure

Regression equation: $Y = 1.748 + 2.785X$ Chi-squared is 1.950 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is $1.167mg/cm^2$ LD_{50} is 0.147 mg/cm² 95% confidence limits are 0.118 to 0.183 mq/cm²

Appendix Table CLXXXVIII: Dose mortality effect of *Pz. zeylanica* (wp/PetE) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 2.544 + 2.514X$ Chi-squared is 2.170 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.977 mg/cm² LD_{50} is 0.095mg/cm² 95% confidence limits are 0.077 to $0.117mg/cm²$

Appendices XLVII IES, RU

Regression equation: $Y = 2.817 + 2.441X$ Chi-squared is 4.620 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.894 mg/cm² LD_{50} is 0.078 mg/cm² 95% confidence limits are 0.063 to 0.097mg/cm²

Appendix Table CXCI: Dose mortality effect of *Pz. zeylanica* (wp/CHCl₃) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = 3.179 + 4.064X$ Chi-squared is 1.189 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.448 mg/cm² LD_{50} is 0.281 mg/cm² 95% confidence limits are 0.204 to $0.387mg/cm²$

Appendix Table CXC: Dose mortality effect of *Pz. zeylanica* (wp/PetE) extracts against *A. gossypii* after 24h of exposure

Regression equation: $Y = 2.766 + 2.715X$ Chi-squared is 4.736 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.823 mg/cm² LD_{50} is 0.067mg/cm² 95% confidence limits are 0.054 to 0.081mg/ $cm²$

Appendix Table CXCII: Dose mortality effect of *Pz. zeylanica* (wp/CHCl₃) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 1.569 + 2.565X$ Chi-squared is 4.877 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.338mg/cm² LD_{50} is 0.218 mg/cm² 95% confidence limits are 0.169 to 0.281mg/cm²

Appendices XLVIII IES, RU

Appendix Table CXCIII: Dose mortality effect of *Pz. zeylanica* (wp/CHCl₃) extracts against *A. gossypii* after 9h of exposure

Regression equation: $Y = 1.136 + 3.004X$ Chi-squared is 14.083 with 4 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.286mg/cm² LD_{50} is 0.193mg/cm² 95% confidence limits are 0.135 to 0.278 mg/cm²

Appendix Table CXCIV: Dose mortality effect of *Pz. zeylanica* (wp/CHCl3) extracts against *A. gossypii* after 12h of exposure

Regression equation: $Y = 2.302 + 2.194X$ Chi-squared is 16.490 with 4 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.230mg/cm² LD_{50} is 0.170 mg/cm² 95% confidence limits are 0.104 to 0.276 mg/cm²

Appendix Table CXCV: Dose mortality effect of *Pz. zeylanica* (wp/CHCl₃) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 2.481 + 2.176X$ Chi-squared is 17.776 with 4 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.158 mg/cm² LD_{50} is 0.144 mg/cm² 95% confidence limits are 0.090 to 0.230mg/cm²

Appendix Table CXCVI: Dose mortality effect of *Pz. zeylanica* (wp/CHCl₃) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 2.641 + 2.2681X$ Chi-squared is 18.797 with 4 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.040 mg/cm² LD_{50} is 0.110 mg/cm² 95% confidence limits are 0.070 to 0.172 mg/cm²

Regression equation: $Y = 3.074 + 2.066X$ Chi-squared is 23.100 with 4 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is $0.932mg/cm^2$ LD_{50} is 0.085mg/cm² 95% confidence limits are 0.049 to $0.149mg/cm²$

Appendix Table CXCIX: Dose mortality effect of *Pz. zeylanica* (wp/CH3OH) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = -3.961 + 18.102X$ Chi-squared is -0.0001 with 0 degrees of freedom

No significant heterogeneity

Log LD_{50} is 0.495 mg/cm²

 LD_{50} is 0.313 mg/cm²

95% confidence limits are 0.287 to 0.341 mq/cm²

Regression equation: $Y = 3.444 + 1.871X$ Chi-squared is 12.236 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is $0.832mg/cm^2$ LD_{50} is 0.068mg/cm² 95% confidence limits are 0.039 to 0.117 mg/cm²

Appendix Table CC: Dose mortality effect of *Pz. zeylanica* (wp/CH₃OH) extracts against *A. gossypii* after 9h of exposure

Regression equation: $Y = 0.471 + 9.095X$ Chi-squared is 0.003 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.498 mg/cm²

 LD_{50} is 0.315mg/cm²

95% confidence limits are 0.275 to $0.361mg/cm²$

Appendices L IES, RU

Regression equation: $Y = 1.598 + 7.257X$ Chi-squared is 0.966 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.469mg/cm² LD_{50} is 0.294 mg/cm² 95% confidence limits are 0.260 to 0.332 mg/cm²

Appendix Table CCII: Dose mortality effect of *Pz. zeylanica* (wp/CH₃OH) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 2.440 + 5.981X$ Chi-squared is 1.425 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.428 mg/cm² LD_{50} is 0.268mg/cm² 95% confidence limits are 0.238 to 0.301mg/cm²

Appendix Table CCIII: Dose mortality effect of *Pz. zeylanica* (wp/CH₃OH) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 3.122 + 4.980X$ Chi-squared is 0.970 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.377mg/cm² LD_{50} is 0.238 mg/cm² 95% confidence limits are 0.212 to $0.267mg/cm²$

Appendix Table CCIV: Dose mortality effect of *Pz. zeylanica* (wp/CH₃OH) extracts against *A. gossypii* after 21h of exposure

Regression equation: $Y = 3.722 + 3.917X$ Chi-squared is 1.665 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.326 mg/cm² LD_{50} is $0.212mg/cm^{2}$ 95% confidence limits are 0.185 to 0.243 mg/cm²

Appendix Table CCV: Dose mortality effect of *Pz. zeylanica* (wp/CH₃OH) extracts

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Regression equation: $Y = -1.323 + 4.971X$ Chi-squared is 1.174 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.272 mg/cm² LD_{50} is 0.187mq/cm² 95% confidence limits are 0.169 to 0.208 mq/cm²

Appendix Table CCVII: Dose mortality effect of *S. nodiflora* (wp/PetE) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = -0.518 + 4.506X$ Chi-squared is 3.390 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.225mg/cm² LD_{50} is 0.168 mg/cm² 95% confidence limits are 0.144 to 0.196mg/cm²

Regression equation: $Y = -0.278 + 4.058X$ Chi-squared is 5.716 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.301mg/cm² LD_{50} is 0.200mg/cm² 95% confidence limits are 0.159 to 0.250 mg/cm²

Appendix Table CCVIII: Dose mortality effect of *S. nodiflora* (wp/PetE) extracts against *A. gossypii* after 9h of exposure

Regression equation: $Y = -0.768 + 4.827X$ Chi-squared is 3.094 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.195mg/cm² LD_{50} is 0.157 mg/cm² 95% confidence limits are 0.137 to 0.180mg/cm²

Regression equation: $Y = 0.160 + 4.267X$ Chi-squared is 5.538 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.134 mg/cm² LD_{50} is 0.136mg/cm² 95% confidence limits are 0.118 to 0.157 mq/cm²

Appendix Table CCX: Dose mortality effect of *S. nodiflora* (wp/PetE) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 0.307 + 4.213X$ Chi-squared is 4.846 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.114 mg/cm² LD_{50} is 0.130 mg/cm² 95% confidence limits are 0.113 to 0.150mg/cm²

Appendix Table CCXI: Dose mortality effect of *S. nodiflora* (wp/PetE) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 1.352 + 3.571X$ Chi-squared is 4.999 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.021 mg/cm² LD_{50} is 0.105 mg/cm² 95% confidence limits are 0.089 to 0.124 mg/cm²

Appendix Table CCXII: Dose mortality effect of *S. nodiflora* (wp/PetE) extracts against *A. gossypii* after 21h of exposure

Regression equation: $Y = 1.874 + 3.331X$ Chi-squared is 8.733 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 0.939 mg/cm² LD_{50} is 0.087mg/cm² 95% confidence limits are 0.065 to 0.116 mg/cm²

Regression equation: $Y = 2.147 + 3.112X$ Chi-squared is 9.771 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 0.917 mg/cm² LD_{50} is 0.083mg/cm² 95% confidence limits are 0.060 to 0.114 mg/cm²

Appendix Table CCXIV: Dose mortality effect of *S. nodiflora* (wp/CHCl3) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = 2.462 + 7.516X$

Chi-squared is -0.00007 with 0 degrees of freedom No significant heterogeneity Log LD_{50} is 0.338 mg/cm² LD_{50} is 0.218 mg/cm² 95% confidence limits are 0.177 to 0.267mg/cm²

Appendix Table CCXV: Dose mortality effect of *S. nodiflora* (wp/CHCl₃) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 0.732 + 3.139X$ Chi-squared is 2.741 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.360mg/cm² LD_{50} is 0.229 mg/cm² 95% confidence limits are 0.163 to 0.321 mg/cm²

Appendix Table CCXVI: Dose mortality effect of *S. nodiflora* (wp/CHCl₃) extracts against *A. gossypii* after 9h of exposure

Regression equation: $Y = 1.651 + 2.613X$ Chi-squared is 7.168 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.282mg/cm² LD_{50} is 0.191 mg/cm² 95% confidence limits are 0.142 to 0.257 mg/cm²

Regression equation: $Y = 1.984 + 2.481X$ Chi-squared is 4.562 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.215mg/cm² LD_{50} is 0.164 mg/cm² 95% confidence limits are 0.126 to 0.214 mg/cm²

Appendix Table CCXIX: Dose mortality effect of *S. nodiflora* (wp/CHCl3) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 2.222 + 2.706X$ Chi-squared is 5.270 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.027 mg/cm² LD_{50} is 0.106 mg/cm² 95% confidence limits are 0.087 to 0.130mg/cm²

Appendix Table CCXVIII: Dose mortality effect of *S. nodiflora* (wp/CHCl3) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 2.012 + 2.783X$ Chi-squared is 3.653 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.073 mg/cm² LD_{50} is 0.118 mg/cm² 95% confidence limits are 0.097 to 0.144 mg/cm²

Appendices LV IES, RU

Appendix Table CCXX: Dose mortality effect of *S. nodiflora* (wp/CHCl₃) extracts against *A. gossypii* after 21h of exposure

Dose	Ldos $(+2)$					#U Kl %Kill Cr% E Pr Ex Pr Wk Pro Weght F Pro		
0.196 0.147 0.098 0.049 0.025	1.292 1.167 0.991 0.690 0.398	30 30 30 30	18 $7\overline{ }$	30 28 93.333 93 6.48 5.883 60.000 60 5.25 5.535 14 46.667 47 4.92 5.044 23.333 23 4.26 4.204 2 6.667 7 3.52 3.389		5.220 4.925 3.572	6.242 15.09 5.863 17.43 19.11 4.252 15.09 6.24 3.408	5.520 5.037 4.210

Regression equation: $Y = 2.315 + 2.746X$ Chi-squared is 4.169 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.978 mg/cm² LD_{50} is 0.095mg/cm² 95% confidence limits are 0.078 to 0.115mg/cm²

Regression equation: $Y = 2.722 + 2.594X$ Chi-squared is 4.206 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.878 mg/cm² LD_{50} is 0.076mq/cm² 95% confidence limits are 0.062 to 0.093 mq/cm²

Appendix Table CCXXIII: Dose mortality effect of *S. nodiflora* (wp/CH₃OH) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 3.796 + 3.442X$ Chi-squared is 0.051 with 1 degrees of freedom No significant heterogeneity Log LD_{50} IS 0.350mg/cm² LD_{50} is 0.224 mg/cm² 95% confidence limits are 0.181 to 0.277 mg/cm²

Appendix Table CCXXII: Dose mortality effect of *S. nodiflora* (wp/CH₃OH) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = 3.159 + 4.418X$ Chi-squared is 1.885 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is $0.417mg/cm^2$ LD_{50} is 0.261mq/cm² 95% confidence limits are 0.204 to 0.334mg/cm²

Appendix Table CCXXIV: Dose mortality effect of *S. nodiflora* (wp/CH3OH) extracts against *A. gossypii* after 9h of exposure

Regression equation: $Y = 1.953 + 2.525X$ Chi-squared is 0.790 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.207mg/cm² LD_{50} is 0.161 mg/cm² 95% confidence limits are 0.131 to 0.197mg/cm²

Regression equation: $Y = 1.818 + 2.822X$ Chi-squared is 0.681 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.128 mg/cm² LD_{50} is 0.134 mg/cm² 95% confidence limits are 0.112 to 0.160 mq/cm²

Appendix Table CCXXVII: Dose mortality effect of *S. nodiflora* (wp/CH3OH) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 2.821 + 2.272X$ Chi-squared is 0.955 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.959mg/cm² LD_{50} is 0.091 mg/cm² 95% confidence limits are 0.071 to 0.117mg/cm²

Appendix Table CCXXVI: Dose mortality effect of *S. nodiflora* (wp/CH₃OH) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 1.956 + 2.921X$ Chi-squared is 2.580 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.042 mg/cm² LD_{50} is 0.110 mg/cm² 95% confidence limits are 0.092 to 0.132 mg/cm²

Appendix Table CCXXVIII: Dose mortality effect of *S. nodiflora* (wp/CH₃OH) extracts against *A. gossypii* after 21h of exposure

Regression equation: $Y = 2.002 + 3.360X$ Chi-squared is 2.871 with 2 degrees of freedom No significant heterogeneity Log LD_{50} IS 0.892mg/cm² LD_{50} is 0.078 mg/cm² 95% confidence limits are 0.064 to 0.095mg/cm²

Appendices LVII IES, RU

Regression equation: $Y = 2.434 + 3.160X$ Chi-squared is 2.010 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is $0.812mg/cm^2$ LD_{50} is 0.065mg/cm² 95% confidence limits are 0.051 to 0.082 mq/cm²

Appendix Table CCXXXI: Dose mortality effect of *Z. zerumbet* (ap/PetE) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 2.841 + 1.679X$ Chi-squared is 2.339 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.286mg/cm² LD_{50} is 0.193mg/cm² 95% confidence limits are 0.103 to 0.364 mg/cm²

Appendix Table CCXXX: Dose mortality effect of *Z. zerumbet* (ap/PetE) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = 2.642 + 1.732X$ Chi-squared is 1.529 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.361mg/cm² LD_{50} is 0.230mg/cm² 95% confidence limits are 0.112 to 0.472 mq/cm²

 Appendix Table CCXXXII: Dose mortality effect of *Z. zerumbet* (ap/PetE) extracts against *A. gossypii* after 9h of exposure

Regression equation: $Y = 3.107 + 1.661X$ Chi-squared is 2.300 with 2 degrees of freedom No significant heterogeneity Log LD_{50} IS 1.140 mg/cm² LD_{50} is 0.138 mg/cm² 95% confidence limits are 0.086 to $0.222mg/cm²$

Regression equation: $Y = 3.288 + 1.711X$ Chi-squared is 3.766 with 3 degrees of freedom No significant heterogeneity Log LD_{50} IS 1.000 mg/cm² LD_{50} is 0.100mg/cm² 95% confidence limits are 0.070 to 0.144 mg/cm²

Appendix Table CCXXXV: Dose mortality effect of *Z. zerumbet* (ap/PetE) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 3.631 + 1.699X$ Chi-squared is 2.716 with 3 degrees of freedom No significant heterogeneity Log LD_{50} IS 0.806mg/cm² LD_{50} is 0.064mq/cm² 95% confidence limits are 0.047 to 0.087mg/cm²

Appendix Table CCXXXIV: Dose mortality effect of *Z. zerumbet* (ap/PetE) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 3.538 + 1.600X$ Chi-squared is 1.534 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.914 mg/cm² LD_{50} is 0.082 mg/cm² 95% confidence limits are 0.058 to 0.116mg/ $cm²$

Appendix Table CCXXXVI: Dose mortality effect of *Z. zerumbet* (ap/PetE) extracts against *A. gossypii* after 21h of exposure

Regression equation: $Y = 3.612 + 1.970X$ Chi-squared is 2.067 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.705 mg/cm² LD_{50} IS 0.051 mg/cm² 95% confidence limits are 0.039 to 0.066 mg/cm²

Appendices LIX IES, RU

Regression equation: $Y = 3.624 + 2.155X$ Chi-squared is 4.878 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.638 mg/cm² LD_{50} is 0.043mg/cm² 95% confidence limits are 0.034 to 0.055mg/cm²

Appendix Table CCXXXVIII: Dose mortality effect of *Z. zerumbet* (ap/CHCl3) against *A. gossypii* after 3h of exposure

Regression equation: $Y = -0.456 + 3.669X$ Chi-squared is 0.057 with 1 degree of freedom No significant heterogeneity Log LD_{50} is 1.489mg/cm² LD_{50} is 0.309mg/cm² 95% confidence limits are 0.162 to 0.588 mg/cm²

Appendix Table CCXXXIX: Dose mortality effect of *Z. zerumbet*(ap/CHCl3) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 1.656 + 2.175X$ Chi-squared is 0.083 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.537mg/cm² LD_{50} is 0.344mg/cm² 95% confidence limits are 0.163 to 0.728 mq/cm²

Appendix Table CCXL: Dose mortality effect of *Z. zerumbet* (ap/CHCl₃) extracts against *A. gossypii* after 9h of exposure

Regression equation: $Y = 2.441 + 1.780X$ Chi-squared is 0.096 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.438 mg/cm² LD_{50} is 0.274mg/cm² 95% confidence limits are 0.155 to 0.485mg/cm²

Regression equation: $Y = 2.205 + 2.223X$ Chi-squared is 0.024 with 3 degrees of freedom No significant heterogeneity Log LD_{50} IS 1.258 mg/cm² LD_{50} is 0.181 mg/cm² 95% confidence limits are 0.131 to 0.249 mg/cm²

Appendix Table CCXLII: Dose mortality effect of *Z. zerumbet* (ap/CHCl3) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 2.263 + 2.513X$ Chi-squared is 1.739 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.089mg/cm² LD_{50} is 0.123 mg/cm² 95% confidence limits are 0.099 to 0.153mg/cm²

Appendix Table CCXLIII: Dose mortality effect of *Z. zerumbet* (ap/CHCl3) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 2.922 + 2.168X$ Chi-squared is 5.340 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.958 mg/cm² LD_{50} is 0.091 mq/cm² 95% confidence limits are 0.072 to 0.115 mq/cm²

Appendix Table CCXLIV: Dose mortality effect of *Z. zerumbet* (ap/CHCl3) extracts against *A. gossypii* after 21h of exposure

Regression equation: $Y = 2.753 + 2.554X$ Chi-squared is 5.794 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.880 mg/cm² LD_{50} is 0.076mq/cm² 95% confidence limits are 0.062 to 0.093mg/cm²

Regression equation: $Y = 3.002 + 2.350X$ Chi-squared is 1.337 with 2 degrees of freedom No significant heterogeneity Log LD_{50} IS 0.851mg/cm² LD_{50} is 0.071mg/cm² 95% confidence limits are 0.056 to 0.090 mg/cm²

Appendix Table CCXLVII: Dose mortality effect of *Z. zerumbet* (ap/CH₃OH) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 0.598 + 3.130X$ Chi-squared is 2.344 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.406mg/cm² LD_{50} is 0.255mg/cm² 95% confidence limits are 0.170 to 0.383mg/cm²

Appendix Table CCXLVI: Dose mortality effect of *Z. zerumbet* (ap/CH₃OH) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = 1.869 + 7.080X$ Chi-squared is 0.079 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.442 mg/cm² LD_{50} is 0.277 mg/cm² 95% confidence limits are 0.227 to 0.337mg/cm²

Appendix Table CCXLVIII: Dose mortality effect of *Z. zerumbet* (ap/CH₃OH) extracts against *A. gossypii* after 9h of exposure

Regression equation: $Y = 0.126 + 3.398X$ Chi-squared is 10.844 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.434 mg/cm² LD_{50} is 0.272mg/cm² 95% confidence limits are 0.161 to 0.460mg/cm²

Regression equation: $Y = 0.047 + 3.613X$ Chi-squared is 9.894 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.371 mg/cm² LD_{50} is 0.235mg/cm² 95% confidence limits are 0.161 to 0.343 mg/cm²

Appendix Table CCLI: Dose mortality effect of *Z. zerumbet* (ap/CH₃OH) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 0.710 + 3.499X$ Chi-squared is 9.968 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.226 mg/cm² LD_{50} is 0.168 mg/cm² 95% confidence limits are 0.127 to 0.223 mg/cm²

Appendix Table CCL: Dose mortality effect of *Z. zerumbet* (ap/CH₃OH) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = -0.402 + 4.201X$ Chi-squared is 9.785 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.286 mg/cm² LD_{50} is 0.193mg/cm² 95% confidence limits are 0.150 to 0.249 mg/cm²

Appendix Table CCLII: Dose mortality effect of *Z. zerumbet* (ap/CH₃OH) extracts against *A. gossypii* after 21h of exposure

Regression equation: $Y = 0.339 + 4.037X$ Chi-squared is 10.840 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.155mg/cm² LD_{50} is 0.143mg/cm² 95% confidence limits are 0.111 to $0.184mg/cm²$

Appendices LXIII IES, RU

Regression equation: $Y = 2.290 + 2.367X$ Chi-squared is 2.328 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.145mg/cm² LD_{50} is 0.140 mg/cm² 95% confidence limits are 0.109 to 0.179 mg/cm²

Appendix Table CCLV: Dose mortality effect of *Z. zerumbet* (rh/PetE) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 2.261 + 2.259X$ Chi-squared is 0.074 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is $1.212mg/cm^2$ LD_{50} is 0.163mg/cm² 95% confidence limits are 0.108 to 0.247mg/cm²

Appendix Table CCLIV: Dose mortality effect of *Z. zerumbet* (rh/PetE) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = 2.512 + 1.872X$ Chi-squared is 0.207 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.329 mg/cm² LD_{50} is 0.213 mg/cm² 95% confidence limits are 0.112 to 0.405mg/cm²

Appendix Table CCLVI: Dose mortality effect of *Z. zerumbet* (rh/PetE) extracts against *A. gossypii* after 9h of exposure

Regression equation: $Y = 3.184 + 1.597X$ Chi-squared is 2.140 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.137 mg/cm² LD_{50} is 0.137 mg/cm² 95% confidence limits are 0.086 to 0.220 mg/cm²

Regression equation: $Y = 3.525 + 1.465X$ Chi-squared is 0.931 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.007 mg/cm² LD_{50} is 0.102 mg/cm² 95% confidence limits are 0.066 to 0.155 mg/cm²

Appendix Table CCLVIII: Dose mortality effect of *Z. zerumbet* (rh/PetE) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 3.646 + 1.601X$ Chi-squared is 0.386 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.845 mg/cm² LD_{50} is 0.070mg/cm² 95% confidence limits are 0.050 to 0.097mg/cm²

Appendix Table CCLIX: Dose mortality effect of *Z. zerumbet* (rh/PetE) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 3.844 + 1.553X$ Chi-squared is 0.821 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.744 mg/cm² LD_{50} is 0.055mg/cm² 95% confidence limits are 0.040 to 0.076mg/cm²

Appendix Table CCLX: Dose mortality effect of *Z. zerumbet* (rh/PetE) extracts against *A. gossypii* after 21h of exposure

Regression equation: $Y = 3.939 + 1.743X$ Chi-squared is 0.969 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.609mg/cm² LD_{50} is 0.041 mq/cm² 95% confidence limits are 0.030 to 0.054mg/cm²

Regression equation: $Y = 4.143 + 1.591X$ Chi-squared is 0.069 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.539 mg/cm² LD_{50} is 0.035mg/cm² 95% confidence limits are 0.025 to 0.049 mg/cm²

Appendix Table CCLXII: Dose mortality effect of *Z. zerumbet* (rh/CHCl3) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = 2.132 + 2.124X$ Chi-squared is 1.019 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.350 mg/cm² LD_{50} is 0.224 mg/cm² 95% confidence limits are 0.144 to 0.349 mg/cm²

Appendix Table CCLXIII: Dose mortality effect of *Z. zerumbet* (rh/CHCl3) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 2.251 + 2.129X$ Chi-squared is 2.737 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.291mg/cm² LD_{50} is 0.195mg/cm² 95% confidence limits are 0.134 to 0.285 mg/cm²

Appendix Table CCLXIV: Dose mortality effect of *Z. zerumbet* (rh/CHCl₃) extracts against *A. gossypii* after 9h of exposure

Regression equation: $Y = 2.993 + 1.669X$ Chi-squared is 3.230 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.202 mg/cm² LD_{50} is 0.159mg/cm² 95% confidence limits are 0.108 to 0.235mg/cm²

Regression equation: $Y = 3.285 + 1.566X$ Chi-squared is 6.463 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.095mg/cm² LD_{50} is 0.124 mg/cm² 95% confidence limits are 0.088 to 0.176mq/cm²

Appendix Table CCLXVII: Dose mortality effect of *Z. zerumbet* (rh/CHCl3) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 3.105 + 2.081X$ Chi-squared is 4.596 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.911 mg/cm² LD_{50} is 0.081 mg/cm² 95% confidence limits are 0.064 to 0.104mg/cm²

Appendix Table CCLXVI: Dose mortality effect of *Z. zerumbet* (rh/CHCl₃) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 3.289 + 1.683X$ Chi-squared is 4.991 with 3 degrees of freedom No significant heterogeneity LOG LD_{50} is 1.017 mg/cm² LD_{50} is 0.104mg/cm² 95% confidence limits are 0.077 to 0.140 mq/cm²

Appendix Table CCLXVIII: Dose mortality effect of *Z. zerumbet* (rh/CHCl₃) extracts against *A. gossypii* after 21h of exposure

Regression equation: $Y = 3.529 + 1.692X$ Chi-squared is 0.279 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.869 mg/cm² LD_{50} is 0.074mg/cm² 95% confidence limits are 0.054 to $0.102mg/cm^2$

Appendices LXVII IES, RU

Regression equation: $Y = 3.386 + 2.038X$ Chi-squared is 0.047 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.792mg/cm² LD_{50} is 0.062mg/cm² 95% confidence limits are 0.047 to 0.081mq/cm²

Appendix Table CCLXXI: Dose mortality effect of *Z. zerumbet* (rh/CH3OH) extracts against *A. gossypii* after 6h of exposure

Regression equation: $Y = 1.264 + 2.553X$ Chi-squared is 0.563 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.463mg/cm² LD_{50} is 0.290mg/cm² 95% confidence limits are 0.190 to 0.444 mg/cm²

Appendix Table CCLXX: Dose mortality effect of *Z. zerumbet* (rh/CH₃OH) extracts against *A. gossypii* after 3h of exposure

Regression equation: $Y = -0.486 + 3.669X$ Chi-squared is 0.299 with 2 degrees of freedom No significant heterogeneity Log LD_{50} IS 1.495mg/cm² LD_{50} is 0.313mg/cm² 95% confidence limits are 0.217 to 0.450mg/cm²

Appendix Table CCLXXII: Dose mortality effect of *Z. zerumbet* (rh/CH₃OH) extracts against *A. gossypii* after 9h of exposure

Regression equation: $Y = 1.763 + 2.369X$ Chi-squared is 0.607 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.366mg/cm² LD_{50} is 0.232mg/cm² 95% confidence limits are 0.173 to $0.312mg/cm²$

Regression equation: $Y = 1.693 + 2.601X$ Chi-squared is 0.113 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.271 mg/cm² LD_{50} is 0.187mg/cm² 95% confidence limits are 0.150 to 0.232 mg/cm²

Appendix Table CCLXXV: Dose mortality effect of *Z. zerumbet* (rh/CH₃OH) extracts against *A. gossypii* after 18h of exposure

Regression equation: $Y = 2.250 + 2.469X$ Chi-squared is 1.504 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.114 mg/cm² LD_{50} is 0.130 mg/cm² 95% confidence limits are 0.106 to 0.159mg/cm²

Appendix Table CCLXXIV: Dose mortality effect of *Z. zerumbet* (rh/CH₃OH) extracts against *A. gossypii* after 15h of exposure

Regression equation: $Y = 1.983 + 2.477X$ Chi-squared is 0.415 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.218 mg/cm² LD_{50} is 0.165mg/cm² 95% confidence limits are 0.134 to 0.204 mg/cm²

Appendix Table CCLXXVI: Dose mortality effect of *Z. zerumbet* (rh/CH₃OH) extracts against *A. gossypii* after 21h of exposure

Regression equation: $Y = 1.987 + 2.858X$ Chi-squared is 1.901 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.054 mg/cm² LD_{50} is 0.113 mg/cm² 95% confidence limits are 0.094 to 0.136 mg/cm²

Appendices LXIX IES, RU

Regression equation: $Y = 2.043 + 2.990X$ Chi-squared is 1.716 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.989mg/cm² LD_{50} is 0.097mg/cm² 95% confidence limits are 0.080 to 0.118 mq/cm²

Appendix Table CCLXXIX: Cytotoxicity effect of *E. nummularius* (wp/PetE) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 3.918 + 0.390X$ Chi-squared is 3.922 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.775 ppm LC_{50} is 594.987ppm 95% confidence limits are 16.143 to 21929.030ppm

Appendix Table CCLXXVIII: Cytotoxicity effect of *E. nummularius* (wp/PetE) extracts against *A. salina* after 6h of exposure

Regression equation: $Y = 3.855 + 0.256X$ Chi-squared is 2.878 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 4.477ppm LC_{50} is 29991.140ppm 95% confidence limits are 0.170 to 5.306E+09ppm

Appendix Table CCLXXX: Cytotoxicity effect of *E. nummularius* (wp/PetE) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 4.210 + 0.341X$ Chi-squared is 2.095 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.318 ppm LC_{50} is 207.889ppm 95% confidence limits are 11.231 to 3847.972ppm

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Regression equation: $Y = 4.120 + 0.583X$ Chi-squared is 1.542 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.510 ppm LC_{50} is 32.328ppm 95% confidence limits are 14.223 to 73.476ppm

Appendix Table CCLXXXIII: Cytotoxicity effect of *E. nummularius* (wp/CHCl3) extracts against *A. salina* after 6h of exposure

Regression equation: $Y = 3.272 + 0.579X$ Chi-squared is 0.372 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.983ppm LC_{50} is 961.694ppm 95% confidence limits are 116.728 to 7923.193ppm

Appendix Table CCLXXXII: Cytotoxicity effect of *E. nummularius* (wp/PetE) extracts against *A. salina* after 30h of exposure

Regression equation: $Y = 4.475 + 0.463X$ Chi-squared is 4.275 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.134 ppm LC_{50} is 13.601ppm 95% confidence limits are 5.286 to 34.991ppm

Appendix Table CCLXXXIV: Cytotoxicity effect of *E. nummularius* (wp/CHCl3) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 3.317 + 0.683X$ Chi-squared is 1.293 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.465ppm LC_{50} is 291.519ppm 95% confidence limits are 106.257 to 799.790ppm

\blacksquare **LXXI** IES, RU

Regression equation: $Y = 3.334 + 0.818X$ Chi-squared is 2.629 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.038ppm LC_{50} is 109.120ppm 95% confidence limits are 61.112 to 194.839ppm

Appendix Table CCLXXXVII: Cytotoxicity effect of *E. nummularius* (wp/CHCl3) extracts against *A. salina* after 30h of exposure

Regression equation: $Y = 3.689 + 1.000X$ Chi-squared is 0.710 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.311ppm LC_{50} is 20.485ppm 95% confidence limits are 8.226 to 51.011ppm

Regression equation: $Y = 3.497 + 1.038X$ Chi-squared is 0.115 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.448 ppm LC_{50} is 28.035ppm 95% confidence limits are 13.327 to 58.977ppm

Appendix TableCCLXXXVIII: Cytotoxicity effect of *E. nummularius* (wp/CH₃OH) extracts against *A. salina* after 6h of exposure

Regression equation: $Y = 2.127 + 0.782X$ Chi-squared is 0.144 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 3.674ppm LC_{50} is 4716.815ppm 95% confidence limits are 169.616 to 131168.900ppm

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Regression equation: $Y = 2.509 + 0.827X$ Chi-squared is 1.731 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 3.013 ppm LC_{50} is 1029.678 ppm 95% confidence limits are 246.416 to 4302.630ppm

Appendix Table CCXC: Cytotoxicity effect of *E. nummularius* (wp/CH3OH) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 2.455 + 1.093X$ Chi-squared is 2.031 with 4 degrees of freedom No significant heterogeneity Log LC_{50} IS 2.328ppm LC_{50} is 213.030ppm 95% confidence limits are 124.660 to 364.046ppm

\blacksquare **LXXIII** IES, RU

95% confidence limits are 86.722 to 248.623ppm

Appendix Table CCXCII: Cytotoxicity effect of *E. nummularius* (wp/CH₃OH) extracts against *A. salina* after 30h of exposure

Regression equation: $Y = 2.604 + 1.258X$ Chi-squared is 5.681 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.905ppm LC_{50} is 80.432ppm 95% confidence limits are 55.841 to 115.854ppm

Regression equation: $Y = 2.245 + 1.097X$ Chi-squared is 0.162 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.512ppm LC_{50} is 324.814ppm 95% confidence limits are 74.339 to 1419.231ppm

Appendix Table CCXCV: Cytotoxicity effect of *L. camara* (ap/PetE) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 2.639 + 1.624X$ Chi-squared is 2.138 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.454ppm LC_{50} is 28.415ppm 95% confidence limits are 20.804 to 38.810ppm

Appendix Table CCXCIV: Cytotoxicity effect of *L. camara* (ap/PetE) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 2.311 + 1.619X$ Chi-squared is 2.448 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.661ppm LC_{50} is 45.819 ppm 95% confidence limits are 32.377 to 64.841ppm

Appendix Table CCXCVI: Cytotoxicity effect of *L. camara* (ap/PetE) extracts against *A. salina* after 24h of exposure

Regression equation: $Y = 3.130 + 1.643X$ Chi-squared is 1.262 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.138ppm LC50 is 13.748ppm 95% confidence limits are 9.740 to 19.404ppm

Regression equation: $Y = 3.120 + 1.772X$ Chi-squared is 1.076 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.061ppm LC_{50} is 11.515ppm 95% confidence limits are 8.159 to 16.251ppm

Appendix Table CCXCVIII: Cytotoxicity effect of *L. camara* (ap/CHCl₃) extracts against *A. salina* after 6h of exposure

────────────────────────────────────────────────────────────────────── Regression equation: $Y = 3.005 + 0.737X$ Chi-squared is 2.099 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.707 ppm LC_{50} is 508.798ppm 95% confidence limits are 48.258 to 5364.418ppm

Appendix Table CCXCIX: Cytotoxicity effect of *L. camara* (ap/CHCl₃) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 3.036 + 1.178X$ Chi-squared is 3.537 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.667ppm LC_{50} is 46.466ppm 95% confidence limits are 28.994 to 74.466ppm

Appendix Table CCC: Cytotoxicity effect of *L. camara* (ap/CHCl₃) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 2.969 + 1.509X$ Chi-squared is 2.380 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.346ppm LC_{50} is 22.178ppm 95% confidence limits are 15.884 to 30.967ppm

Appendix Table CCCI: Cytotoxicity effect of *L. camara* (ap/CHCl3) extracts against *A.*

salina after 24h of exposure

Regression equation: $Y = 3.248 + 1.537X$ Chi-squared is 1.818 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.140ppm LC_{50} is 13.802ppm 95% confidence limits are 9.562 to 19.924ppm

Appendix Table CCCIII: Cytotoxicity effect of *L. camara* (ap/CH₃OH) extracts against *A. salina* after 6h of exposure

Regression equation: $Y = 2.755 + 0.852X$ Chi-squared is 0.345 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.636ppm LC_{50} is 432.490 ppm 95% confidence limits are 103.267 to 1811.301ppm

Appendix Table CCCII: Cytotoxicity effect of *L. camara* (ap/CHCl₃) extracts against *A. salina* after 30h of exposure

Regression equation: $Y = 3.166 + 1.797X$ Chi-squared is 0.333 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.020 ppm LC_{50} is 10.481ppm 95% confidence limits are 7.316 to 15.016ppm

Appendix Table CCCIV: Cytotoxicity effect of *L. camara* (ap/CH3OH) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 2.672 + 0.979X$ Chi-squared is 0.465 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.378 ppm LC_{50} is 238.550ppm 95% confidence limits are 94.570 to 601.736ppm

25.000 1.398 30 8 26.667 27 4.39 4.352 4.394 15.96 4.340 12.500 1.097 30 6 20.000 20 4.16 4.082 4.160 13.17 4.065

Appendix Table CCCV: Cytotoxicity effect of *L. camara* (ap/CH3OH) extracts against

────────────────────────────────────────────────────────────────────── Regression equation: $Y = 2.771 + 1.153X$ Chi-squared is 3.576 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.933ppm LC_{50} is 85.608ppm 95% confidence limits are 53.667 to 136.561ppm

Appendix Table CCCVII: Cytotoxicity effect of *L. camara* (ap/CH₃OH) extracts against *A. salina* after 30h of exposure

Regression equation: $Y = 2.871 + 1.215X$ Chi-squared is 2.661 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.752ppm LC_{50} is 56.523ppm 95% confidence limits are 37.722 to 84.694ppm

Appendix Table CCCVIII: Cytotoxicity effect of *L. camara* (r/PetE) extracts against *A. salina* after 6h of exposure

Regression equation: $Y = 2.717 + 0.713X$ Chi-squared is 0.045 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 3.203ppm LC_{50} is 1595.095ppm 95% confidence limits are 14.936 to 170344.700ppm

Appendix Table CCCIX: Cytotoxicity effect of *L. camara* (r/PetE) extracts against *A. salina* after 12h of exposure

────────────────────────────────────────────────────────────────────── Regression equation: $Y = 3.578 + 0.694X$ Chi-squared is 0.559 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.048ppm LC_{50} is 111.786ppm

95% confidence limits are 32.363 to 386.121ppm

Appendix Table CCCX: Cytotoxicity effect of *L. camara* (r/PetE) extracts against *A. salina* after 18h of exposure

────────────────────────────────────────────────────────────────────── Regression equation: $Y = 3.714 + 0.894X$ Chi-squared is 1.335 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.439ppm LC_{50} is 27.467ppm 95% confidence limits are 16.142 to 46.739ppm

Appendix Table CCCXI: Cytotoxicity effect of *L. camara* (r/PetE) extracts against *A. salina* after 24h of exposure

Chi-squared is 0.098 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.183ppm LC_{50} is 15.254ppm 95% confidence limits are 9.656 to 24.098ppm

Appendix TableCCCXII: Cytotoxicity effect of *L. camara* (r/PetE) extracts against *A. salina* after 30h of exposure

Regression equation: $Y = 3.713 + 1.174X$ Chi-squared is 0.038 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.097ppm LC_{50} is 12.497ppm 95% confidence limits are 7.642 to 20.437ppm

Regression equation: $Y = 2.705 + 1.087X$ Chi-squared is 1.014 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.111ppm LC_{50} is 129.074ppm 95% confidence limits are 70.805 to 235.294ppm

Appendix Table CCCXIV: Cytotoxicity effect of *L. camara* (r/CHCl₃) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 1.935 + 1.954X$ Chi-squared is 2.670 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.569ppm LC_{50} is 37.045ppm 95% confidence limits are 28.147 to 48.756ppm

Appendix Table CCCXV: Cytotoxicity effect of *L. camara* (r/CHCl₃) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 1.766 + 2.266X$ Chi-squared is 0.571 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.427ppm LC_{50} is 26.716ppm 95% confidence limits are 20.557 to 34.720ppm

Appendix Table CCCXVI: Cytotoxicity effect of *L. camara* (r/CHCl₃) extracts against *A. salina* after 24h of exposure

Regression equation: $Y = 2.265 + 2.048X$ Chi-squared is 0.313 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.335 ppm LC_{50} is 21.630ppm 95% confidence limits are 15.865 to 29.489ppm

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Regression equation: $Y = 2.504 + 2.027X$ Chi-squared is 0.321 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.231ppm LC_{50} is 17.035ppm 95% confidence limits are 11.899 to 24.388ppm

Appendix Table CCCXVIII: Cytotoxicity effect of *L. camara* (r/CH₃OH) extracts against *A. salina* after 6h of exposure

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Regression equation: $Y = 1.796 + 1.361X$ Chi-squared is 2.094 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 2.354ppm

 LC_{50} is 225.929ppm

95% confidence limits are 110.841 to 460.514ppm

Appendix TableCCCXIX: Cytotoxicity effect of *L. camara* (r/CH₃OH) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 1.519 + 2.028X$ Chi-squared is 4.083 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.716ppm LC_{50} is 52.056ppm 95% confidence limits are 40.162 to 67.472ppm

Appendix Table CCCXX: Cytotoxicity effect of *L. camara* (r/CH₃OH) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 1.197 + 2.381X$ Chi-squared is 1.080 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.597ppm LC_{50} is 39.515 ppm 95% confidence limits are 31.208 to 50.033ppm

Regression equation: $Y = 1.616 + 2.376X$ Chi-squared is 0.372 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.424ppm LC_{50} is 26.555ppm 95% confidence limits are 20.647 to 34.153ppm

Appendix Table CCCXXII: Cytotoxicity effect of *L. camara* (r/CH₃OH) extracts against *A. salina* after 30h of exposure

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Regression equation: $Y = 2.442 + 2.047X$ Chi-squared is 0.132 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.249ppm LC_{50} is 17.761ppm 95% confidence limits are 12.571 to 25.093ppm

Appendix Table CCCXXIII: Cytotoxicity effect of *M. piperita* (wp/PetE) extracts against *A. salina* after 6h of exposure

Regression equation: $Y = 3.137 + 0.740X$ Chi-squared is 1.413 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.518ppm LC_{50} is 329.968ppm 95% confidence limits are 46.135 to 2360.012ppm

Appendix Table CCCXXIV: Cytotoxicity effect of *M. piperita* (wp/PetE) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 2.516 + 1.588X$ Chi-squared is 8.472 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LC_{50} is 1.564ppm LC_{50} is 36.670ppm 95% confidence limits are 21.071 to 63.817ppm

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Regression equation: $Y = 2.657 + 1.697X$ Chi-squared is 9.267 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.380 ppm LC_{50} is 24.011ppm 95% confidence limits are 18.045 to 31.948ppm

Appendix Table CCCXXVI: Cytotoxicity effect of *M.piperita* (wp/PetE) extracts against *A. salina* after 24h of exposure

Regression equation: $Y = 3.394 + 1.326X$ Chi-squared is 1.573 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.211 ppm LC_{50} is 16.253ppm 95% confidence limits are 11.137 to 23.719ppm

Appendix Table CCCXXVII: Cytotoxicity effect of *M. piperita* (wp/PetE) extracts against *A. salina* after 30h of exposure

Regression equation: $Y = 3.362 + 1.670X$ Chi-squared is 3.583 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 0.981ppm LC_{50} is 9.573ppm 95% confidence limits are 7.003 to 13.084ppm

Appendix Table CCCXXVIII: Cytotoxicity effect of *M. piperita* (wp/CHCl₃) extracts against *A. salina* after 6h of exposure

Regression equation: $Y = 2.638 + 1.335X$ Chi-squared is 3.538 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.769ppm LC_{50} is 58.807 ppm 95% confidence limits are 36.827 to 93.905ppm

Regression equation: $Y = 1.364 + 2.487X$ Chi-squared is 1.385 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.462ppm LC_{50} is 28.980ppm 95% confidence limits are 23.061 to 36.418ppm

Appendix Table CCCXXXI: Cytotoxicity effect of *M. piperita* (wp/CHCl₃) extracts against *A. salina* after 24h of exposure

Regression equation: $Y = 2.731 + 1.749X$ Chi-squared is 2.382 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.297ppm LC_{50} is 19.830ppm 95% confidence limits are 14.422 to 27.265ppm

Appendix Table CCCXXX: Cytotoxicity effect of *M. piperita* (wp/CHCl₃) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 2.395 + 1.933X$ Chi-squared is 3.995 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.348ppm LC50 is 22.295ppm 95% confidence limits are 17.025 to 29.196ppm

Appendix Table CCCXXXII: Cytotoxicity effect of *M. piperita* (wp/CHCl₃) extracts against *A. salina* after 30h of exposure

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Regression equation: $Y = 2.925 + 1.832X$ Chi-squared is 1.839 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.133ppm LC_{50} is 13.573ppm 95% confidence limits are 9.891 to 18.626ppm

Regression equation: $Y = 2.317 + 0.986X$ Chi-squared is 0.479 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 2.720ppm LC_{50} is 525.030 ppm 95% confidence limits are 180.989 to 1523.052ppm

Appendix Table CCCXXXIV: Cytotoxicity effect of *M. piperita* (wp/CH₃OH) extracts against *A. salina* after 12h of exposure

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Regression equation: $Y = 1.438 + 1.499X$ Chi-squared is 1.670 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.377 ppm LC_{50} is 238.022ppm 95% confidence limits are 156.184 to 362.742ppm **Appendix Table CCCXXXV:** Cytotoxicity effect of *M. piperita* (wp/CH₃OH) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 2.341 + 1.204X$ Chi-squared is 0.069 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.210 ppm LC_{50} is 162.029ppm 95% confidence limits are 104.473 to 251.292ppm

Appendix Table CCCXXXVI: Cytotoxicity effect of *M. piperita* (wp/CH₃OH) extracts against *A. salina* after 24h of exposure

Regression equation: $Y = 1.997 + 1.422X$ Chi-squared is 0.990 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.112 ppm LC_{50} is 129.544 ppm 95% confidence limits are 90.751 to 184.919ppm

Regression equation: $Y = 2.280 + 1.435X$ Chi-squared is 0.638 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.895ppm LC_{50} is 78.572ppm 95% confidence limits are 55.163 to 111.916ppm

Appendix Table CCCXXXVIII: Cytotoxicity effect of *Mi. pudica* (wp/PetE) extracts against *A. salina* after 6h of exposure

Regression equation: $Y = 2.421 + 1.093X$ Chi-squared is 1.056 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.359ppm LC_{50} is 228.574ppm

95% confidence limits are 66.397 to 786.869ppm

Appendix Table CCCXXXIX: Cytotoxicity effect of *Mi. pudica* (wp/PetE) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 1.626 + 1.899X$ Chi-squared is 0.856 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.777ppm LC_{50} is 59.827ppm 95% confidence limits are 42.662 to 83.899ppm

Appendix Table CCCXL: Cytotoxicity effect of *Mi. pudica* (wp/PetE) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 2.087 + 1.744X$ Chi-squared is 1.558 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.670ppm LC50 is 46.757ppm 95% confidence limits are 33.732 to 64.812ppm

Regression equation: $Y = 2.519 + 1.562$ X Chi-squared is 1.033 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.588ppm LC_{50} is 38.768ppm 95% confidence limits are 27.593 to 54.470ppm

Appendix Table CCCXLII: Cytotoxicity effect of *Mi. pudica* (wp/PetE) extracts against *A. salina* after 30h of exposure

Regression equation: $Y = 2.138 + 2.079X$ Chi-squared is 0.385 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.377ppm LC_{50} is 23.815ppm

95% confidence limits are 18.432 to 30.769ppm

Appendix Table CCCXLIII: Cytotoxicity effect of *Mi. pudica* (wp/CHCl₃) extracts against *A. salina* after 6h of exposure

Regression equation: $Y = 2.424 + 1.145X$ Chi-squared is 2.247 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.251ppm LC_{50} is 178.222ppm 95% confidence limits are 90.457 to 351.137ppm

Appendix Table CCCXLIV: Cytotoxicity effect of *Mi. pudica* (wp/CHCl₃) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 1.650 + 1.778X$ Chi-squared is 4.554 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.884ppm LC_{50} is 76.568 ppm 95% confidence limits are 56.605 to 103.571ppm

Regression equation: $Y = 1.733 + 1.874X$ Chi-squared is 2.307 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.743ppm LC_{50} is 55.318ppm 95% confidence limits are 41.845 to 73.129ppm

Appendix Table CCCXLVI: Cytotoxicity effect of *Mi. pudica* (wp/CHCl3) extracts against *A. salina* after 24h of exposure

Regression equation: $Y = 2.208 + 1.655X$ Chi-squared is 4.163 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.687ppm LC_{50} is 48.650ppm 95% confidence limits are 35.803 to 66.108ppm

Appendix Table CCCXLVII: Cytotoxicity effect of *Mi. pudica* (wp/CHCl3) extracts against *A. salina* after 30h of exposure

Regression equation: $Y = 2.507 + 1.618X$ Chi-squared is 3.430 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.541ppm LC_{50} is 34.767ppm 95% confidence limits are 25.079 to 48.195ppm

Appendix TableCCCXLVIII: Cytotoxicity effect of *Mi. pudica* (wp/CH3OH) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 2.341 + 0.856X$ Chi-squared is 0.520 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 3.108 ppm LC_{50} is 1282.236 ppm 95% confidence limits are 170.560 to 9639.580ppm

Regression equation: $Y = 2.535 + 0.949X$ Chi-squared is 0.583 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.597ppm LC_{50} is 395.636ppm 95% confidence limits are 167.372 to 935.214ppm

Appendix Table CCCL: Cytotoxicity effect of *Mi. pudica* (wp/CH₃OH) extracts against *A. salina* after 24h of exposure

Regression equation: $Y = 2.463 + 1.178X$ Chi-squared is 0.542 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.153ppm LC_{50} is 142.187ppm 95% confidence limits are 92.397 to 218.807ppm

Appendix Table CCCLI: Cytotoxicity effect of *Mi. pudica* (wp/CH₃OH) extracts against *A. salina* after 30h of exposure

Regression equation: $Y = 2.141 + 1.476X$ Chi-squared is 0.584 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.938ppm LC_{50} is 86.651ppm 95% confidence limits are 61.605 to 121.880ppm

Appendix Table CCCLII: Cytotoxicity effect of *P. hysterophorus* (wp/PetE) extracts against *A. salina* after 6h of exposure

Regression equation: $Y = 3.065 + 0.738X$ Chi-squared is 0.147 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.621ppm LC_{50} is 418.086 ppm 95% confidence limits are 84.846 to 2060.159ppm

Regression equation: $Y = 2.324 + 1.422X$ Chi-squared is 1.012 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.881ppm LC_{50} is 76.099ppm 95% confidence limits are 52.576 to 110.146ppm

Appendix Table CCCLV: Cytotoxicity effect of *P. hysterophorus* (wp/PetE) extracts against *A. salina* after 24h of exposure

Regression equation: $Y = 2.459 + 1.581X$ Chi-squared is 0.366 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.607ppm LC_{50} is 40.454ppm 95% confidence limits are 29.252 to 55.946ppm

Appendix Table CCCLIV: Cytotoxicity effect of *P. hysterophorus* (wp/PetE) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 2.494 + 1.455$ X Chi-squared is 0.691 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.723ppm LC_{50} is 52.790ppm 95% confidence limits are 37.533 to 74.249ppm

Appendix Table CCCLVI: Cytotoxicity effect of *P. hysterophorus* (wp/PetE) extracts against *A. salina* after 30h of exposure

Regression equation: $Y = 2.531 + 1.710X$ Chi-squared is 0.202 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.444 ppm LC_{50} is 27.782ppm 95% confidence limits are 19.980 to 38.629ppm

Regression equation: $Y = 2.228 + 1.170X$ Chi-squared is 1.599 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.369ppm LC_{50} is 233.829ppm 95% confidence limits are 107.394 to 509.118ppm

Appendix Table CCCLIX: Cytotoxicity effect of *P. hysterophorus* (wp/CHCl3) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 1.393 + 1.889X$ Chi-squared is 4.705 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.910ppm LC_{50} is 81.277ppm 95% confidence limits are 60.728 to 108.778ppm

Regression equation: $Y = 1.373 + 1.875X$ Chi-squared is 3.480 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.935ppm LC_{50} is 86.089ppm 95% confidence limits are 63.948 to 115.898ppm

Appendix Table CCCLX: Cytotoxicity effect of *P. hysterophorus* (wp/CHCl3) extracts against *A. salina* after 24h of exposure

Regression equation: $Y = 1.789 + 1.814X$ Chi-squared is 3.747 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.770ppm LC_{50} is 58.931ppm 95% confidence limits are 44.239 to 78.501ppm

Regression equation: $Y = 1.963 + 1.816X$ Chi-squared is 0.508 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.673ppm LC_{50} is 47.061ppm 95% confidence limits are 35.472 to 62.436ppm

Appendix Table CCCLXIII: Cytotoxicity effect of *P. hysterophorus* (wp/CH₃OH) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 1.253 + 2.090X$ Chi-squared is 2.665 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.793ppm LC_{50} is 62.016ppm 95% confidence limits are 48.049 to 80.041ppm

Regression equation: $Y = 2.493 + 1.066X$ Chi-squared is 1.296 with 3 degrees of freedom No significant heterogeneity Log LC_{50} Is 2.352 ppm LC_{50} is 224.663ppm 95% confidence limits are 98.071 to 514.660ppm Appendix Table CCCLXIV: Cytotoxicity effect of *P. hysterophorus* (wp/CH₃OH) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 1.112 + 2.321X$ Chi-squared is 2.598 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.675ppm LC_{50} is 47.302ppm 95% confidence limits are 37.421 to 59.791ppm

Regression equation: $Y = 1.345 + 2.223X$ Chi-squared is 4.231 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.644ppm LC_{50} is 44.060 ppm 95% confidence limits are 33.933 to 57.207ppm

Appendix Table CCCLXVI: Cytotoxicity effect of *P.hysterophorus* (wp/CH₃OH) extracts against *A. salina* after 30h of exposure

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Regression equation: $Y = 1.375 + 2.397X$ Chi-squared is 1.864 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.512ppm LC_{50} is 32.513ppm

95% confidence limits are 25.566 to 41.349ppm

Appendix Table CCCLXVII: Cytotoxicity effect of *Ph. niruri* (wp/PetE) extracts against *A. salina* after 6h of exposure

Regression equation: $Y = 2.829 + 0.589X$ Chi-squared is 1.607 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 3.686 ppm LC_{50} is 4845.618 ppm 95% confidence limits are 181.877 to 129098.700ppm

Appendix Table CCCLXVIII: Cytotoxicity effect of *Ph. niruri* (wp/PetE) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 2.915 + 0.700X$ Chi-squared is 0.823 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.980ppm LC_{50} is 955.244ppm

95% confidence limits are 195.075 to 4677.654ppm

Regression equation: $Y = 2.893 + 0.837X$ Chi-squared is 0.975 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.517 ppm LC_{50} is 329.020ppm 95% confidence limits are 141.410 to 765.534ppm

Appendix Table CCCLXX: Cytotoxicity effect of *Ph. niruri* (wp/PetE) extracts against *A. salina* after 24h of exposure

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Regression equation: $Y = 2.628 + 1.163X$ Chi-squared is 0.559 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.040ppm LC_{50} is 109.640ppm 95% confidence limits are 73.122 to 164.395ppm

Appendix Table CCCLXXI: Cytotoxicity effect of *Ph. niruri* (wp/PetE) extracts against *A. salina* after 30h of exposure

Regression equation: $Y = 2.830 + 1.287X$ Chi-squared is 4.014 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.687 ppm LC_{50} is 48.604ppm 95% confidence limits are 33.656 to 70.189ppm

Appendix Table CCCLXXII: Cytotoxicity effect of *Ph. niruri* (wp/CHCl₃) extracts against *A. salina* after 6h of exposure

Regression equation: $Y = 2.897 + 0.655X$ Chi-squared is 1.381 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 3.209ppm LC_{50} is 1617.613ppm 95% confidence limits are 111.518 to 23464.230ppm

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Regression equation: $Y = 3.684 + 0.638X$ Chi-squared is 4.986 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.063 ppm LC_{50} is 115.700ppm 95% confidence limits are 45.056 to 297.106ppm

Appendix Table CCCLXXIV: Cytotoxicity effect of *Ph. niruri* (wp/CHCl₃) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 3.765 + 0.805X$ Chi-squared is 8.730 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.533ppm LC_{50} is 34.150 ppm 95% confidence limits are 19.858 to 58.728ppm

Appendix Table CCCLXXV: Cytotoxicity effect of *Ph. niruri* (wp/CHCl₃) extracts against *A. salina* after 24h of exposure

Regression equation: $Y = 3.724 + 0.907X$ Chi-squared is 9.694 with 4 degrees of freedom Variance has been adjusted for heterogeneity Log LC_{50} is 1.407 ppm LC_{50} is 25.543ppm 95% confidence limits are 11.738 to 55.582ppm

Appendix Table CCCLXXVI: Cytotoxicity effect of *Ph. niruri* (wp/CHCl₃) extracts against *A. salina* after 30h of exposure

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Regression equation: $Y = 4.336 + 0.479X$ Chi-squared is 1.338 with 3 degrees of freedom No significant heterogeneity LOG LC_{50} is 1.386 ppm LC⁵⁰ is 24.331 ppm 95% confidence limits are 9.212 to 64.267 ppm

Regression equation: $Y = 1.511 + 1.093X$ Chi-squared is 2.422 with 4 degrees of freedom No significant heterogeneity LOG LC_{50} is 3.192 ppm LC_{50} is 1557.524 ppm 95% confidence limits are 555.412 to 4367.718 ppm

Appendix Table CCCLXVIII: Cytotoxicity effect of *Ph. niruri* (wp/CH₃OH) extracts against *A. salina* after 12h of exposure

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Regression equation: $Y = 1.700 + 1.149X$ Chi-squared is 1.572 with 4 degrees of freedom No significant heterogeneity LOG LC_{50} is 2.871 ppm LC_{50} is 743.490 ppm 95% confidence limits are 381.772 to 1447.926 ppm

Appendix Table CCLXXIX: Cytotoxicity effect of *Ph. niruri* (wp/CH₃OH) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 1.876 + 1.208X$ Chi-squared is 6.892 with 4 degrees of freedom No significant heterogeneity LOG LC_{50} is 2.585 ppm LC_{50} is 384.977 ppm 95% confidence limits are 240.904 to 615.215 ppm

Appendix Table CCCLXXX: Cytotoxicity effect of *Ph. niruri* (wp/CH₃OH) extracts against *A. salina* after 24h of exposure

Regression equation: $Y = 2.257 + 1.106$ X Chi-squared is 3.927 with 4 degrees of freedom No significant heterogeneity LOG LC_{50} is 2.479 ppm LC_{50} is 301.357 ppm 95% confidence limits are 189.072 to 480.326 ppm

Regression equation: $Y = 1.993 + 1.377X$ Chi-squared is 2.027 with 4 degrees of freedom No significant heterogeneity LOG LC_{50} is 2.183 ppm LC_{50} is 152.499ppm 95% confidence limits are 108.849 to 213.652ppm

Appendix Table CCCLXXXIII: Cytotoxicity effect of *Po. hydropiper* (wp/Pet.E) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 3.825 + 0.547X$ Chi-squared is 0.104 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.150 ppm LC_{50} is 141.108ppm 95% confidence limits are 42.669 to 466.643ppm

Appendix Table CCCXXXII: Cytotoxicity effect of *Po. hydropiper* (wp/Pet.E) extracts against *A. salina* after 6h of exposure

Regression equation: $Y = 1.564 + 0.949X$ Chi-squared is 0.068 with 1 degrees of freedom No significant heterogeneity Log LC_{50} is 3.621ppm LC_{50} is 4175.449 ppm 95% confidence limits are 4.825 to 3613104ppm

Appendix Table CCCLXXXIV: Cytotoxicity effect of *Po. hydropiper* (wp/Pet.E) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 3.582 + 1.203X$ Chi-squared is 0.988 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.179ppm LC⁵⁰ is 15.102ppm 95% confidence limits are 9.676 to 23.573ppm

Regression equation: $Y = 3.387 + 1.559X$ Chi-squared is 1.216 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.035ppm LC_{50} is 10.831ppm 95% confidence limits are 7.263 to 16.151ppm

Appendix Table CCCLXXXVI: Cytotoxicity effect of *Po. hydropiper* (wp/Pet.E) extracts against *A. salina* after 30h of exposure

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Regression equation: $Y = 3.317 + 1.772X$

Chi-squared is 0.245 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 0.949ppm LC_{50} is 8.901ppm 95% confidence limits are 6.013 to 13.17ppm

Appendix Table CCCLXXXVII: Cytotoxicity effect of *Po. hydropiper* (wp/CHCl3) extracts against *A. salina* after 6h of exposure

Regression equation: $Y = 2.930 + 1.124X$ Chi-squared is 0.394 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.842 ppm LC_{50} is 69.464ppm 95% confidence limits are 38.534 to 125.218ppm

Appendix Table CCCLXXXVIII: Cytotoxicity effect of *Po. hydropiper* (wp/CHCl3) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 3.421 + 1.230X$ Chi-squared is 4.832 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.283ppm LC_{50} is 19.208ppm 95% confidence limits are 13.278 to 27.786ppm

Regression equation: $Y = 4.158 + 1.150X$ Chi-squared is 2.025 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 0.732ppm LC_{50} is 5.400ppm 95% confidence limits are 3.156 to 9.238ppm

Appendix Table CCCXC: Cytotoxicity effect of *Po. hydropiper* (wp/CHCl₃) extracts against *A. salina* after 24h of exposure

Regression equation: $Y = 4.399 + 1.302X$ Chi-squared is 1.103 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 0.461ppm LC_{50} is 2.892ppm

95% confidence limits are 1.479 to 5.653ppm

Appendix Table CCCXCI: Cytotoxicity effect of *Po. hydropiper* (wp/CHCl₃) extracts against *A. salina* after 30h of exposure

Regression equation: $Y = 4.789 + 1.045X$ Chi-squared is 0.813 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 0.202 ppm LC_{50} is 1.590ppm 95% confidence limits are 0.439 to 5.760ppm

Appendix Table CCCXCII: Cytotoxicity effect of *Po. hydropiper* (wp/CH3OH) extracts against *A. salina* after 6h of exposure

Regression equation: $Y = 3.467 + 0.416X$ Chi-squared is 1.435 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 3.686ppm LC_{50} is 4852.087ppm 95% confidence limits are 11.942 to 1971432ppm

Regression equation: $Y = 2.250 + 1.588X$ Chi-squared is 11.149 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LC_{50} is 1.732ppm LC_{50} is 53.895ppm 95% confidence limits are 29.307 to 99.111ppm

Appendix Table CCCXCIV: Cytotoxicity effect of *Po. hydropiper* (wp/CH₃OH) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 2.240 + 1.885X$ Chi-squared is 8.619 with 2 degrees of freedom Variance has been adjusted for heterogeneity Log LC_{50} is 1.464ppm LC_{50} is 29.110ppm 95% confidence limits are 15.499 to 54.674ppm

Appendix Table CCCXCV: Cytotoxicity effect of *Po. hydropiper* (wp/CH₃OH) extracts against *A. salina* after 24h of exposure

Regression equation: $Y = 2.539 + 1.752X$ Chi-squared is 8.104 with 2 degrees of freedom Variance has been adjusted for heterogeneity Log LC_{50} is 1.405ppm LC_{50} is 25.398ppm 95% confidence limits are 12.914 to 49.950ppm

Appendix Table CCCXCVI: Cytotoxicity effect of *Po. hydropiper* (wp/CH₃OH) extracts against *A. salina* after 30h of exposure

Regression equation: $Y = 3.001 + 1.512X$ Chi-squared is 10.610 with 2 degrees of freedom Variance has been adjusted for heterogeneity Log LC_{50} is 1.322 ppm LC_{50} is 20.995ppm 95% confidence limits are 7.963 to 55.354ppm

Regression equation: $Y = 2.655 + 0.968X$ Chi-squared is 1.448 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.424 ppm LC_{50} is 265.189ppm 95% confidence limits are 71.529 to 983.166ppm

Appendix Table CCCXCVIII: Cytotoxicity effect of *Pz. zeylanica* (wp/PetE) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 3.059 + 0.925X$ Chi-squared is 4.882 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.099 ppm LC_{50} is 125.528 ppm 95% confidence limits are 49.105 to 320.887ppm

Appendix Table CCCXCIX: Cytotoxicity effect of *Pz. zeylanica* (wp/PetE) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 3.237 + 0.975X$ Chi-squared is 4.183 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.807 ppm LC_{50} is 64.177ppm 95% confidence limits are 33.523 to 122.864ppm

Appendix Table CD: Cytotoxicity effect of *Pz. zeylanica* (wp/PetE) extracts against *A. salina* after 24h of exposure

Regression equation: $Y = 3.434 + 1.047X$ Chi-squared is 1.702 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.496ppm LC_{50} is 31.359ppm 95% confidence limits are 19.750 to 49.793ppm

A. salina after 30h of exposure ──────────────────────────────────────────────────────────────────────

Appendix Table CDI: Cytotoxicity effect of *Pz. zeylanica* (wp/PetE) extracts against

Regression equation: $Y = 3.884 + 1.046X$ Chi-squared is 5.126 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.066ppm LC_{50} is 11.648ppm 95% confidence limits are 7.431 to 18.259ppm

Appendix Table CDII: Cytotoxicity effect of *Pz. zeylanica* (wp/CHCl₃) extracts against *A. salina* after 6h of exposure

Regression equation: $Y = 2.472 + 0.505X$ Chi-squared is 1.143 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 5.006 ppm LC⁵⁰ is 101415.100ppm 95% confidence limits are 0.006 to 1.751E+12ppm

Appendix Table CDIII: Cytotoxicity effect of *Pz. zeylanica* (wp/CHCl₃) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 2.391 + 0.937X$ Chi-squared is 1.444 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.784ppm LC_{50} is 608.430ppm 95% confidence limits are 123.423 to 2999.348ppm

Appendix Table CDIV: Cytotoxicity effect of *Pz. zeylanica* (wp/CHCl₃) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 3.151 + 0.711X$ Chi-squared is 0.477 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.603 ppm LC_{50} is 400.674 ppm 95% confidence limits are 79.628 to 2016.126ppm

Regression equation: $Y = 3.290 + 0.762X$ Chi-squared is 0.4610 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.243ppm LC_{50} is 175.094ppm 95% confidence limits are 64.678 to 474.008ppm

Appendix Table CDVI: Cytotoxicity effect of *Pz. zeylanica* (wp/CHCl₃) extracts against *A. salina* after 30h of exposure

Regression equation: $Y = 2.725 + 1.292X$ Chi-squared is 1.266 with 3 degrees of freedom No significant heterogeneity

Log LC_{50} is 1.761ppm

- LC_{50} is 57.682ppm
- 95% confidence limits are 39.396 to 84.458ppm

Appendix Table CDVII: Cytotoxicity effect of *Pz. zeylanica* (wp/CH₃OH) extracts against *A. salina* after 6h of exposure

Regression equation: $Y = 2.847 + 0.760X$ Chi-squared is 0.274 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.834ppm LC_{50} is 682.599ppm 95% confidence limits are 96.215 to 4842.714ppm

Appendix Table CDVIII: Cytotoxicity effect of *Pz. zeylanica* (wp/CH₃OH) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 3.591 + 0.5840X$ Chi-squared is 0.110 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.412 ppm LC_{50} is 257.960ppm 95% confidence limits are 53.586 to 1241.796ppm

Regression equation: $Y = 3.398 + 0.825X$ Chi-squared is 0.330 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.941ppm LC_{50} is 87.372ppm 95% confidence limits are 45.519 to 167.707ppm

────────────────────────────────────────────────────────────────────── Regression equation: $Y = 3.059 + 1.191X$ Chi-squared is 1.338 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.629ppm LC_{50} is 42.562 ppm 95% confidence limits are 28.203 to 64.234ppm

Appendix Table CDXI: Cytotoxicity effect of *Pz. zeylanica* (wp/CH₃OH) extracts against *A. salina* after 30h of exposure

Chi-squared is 1.687 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.515ppm LC_{50} is 32.749ppm 95% confidence limits are 22.271 to 48.157ppm

Appendix Table CDXII: Cytotoxicity effect of *S. nodiflora* (wp/PetE) extracts against *A. salina* after 6h of exposure

Regression equation: $Y = 2.961 + 0.709X$ Chi-squared is 0.314 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.878ppm LC⁵⁰ is 754.710ppm 95% confidence limits are 113.488 to 5018.917ppm

Appendix Table CDXIII: Cytotoxicity effect of *S. nodiflora* (wp/PetE) extracts against *A. salina* after 12h of exposure

────────────────────────────────────────────────────────────────────── Regression equation: $Y = 3.508 + 0.637X$ Chi-squared is 1.628 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.342 ppm LC_{50} is 219.824ppm 95% confidence limits are 63.008 to 766.934ppm

Appendix Table CDXIV: Cytotoxicity effect of *S. nodiflora* (wp/PetE) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 3.958 + 0.704X$ Chi-squared is 0.416 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.480ppm LC_{50} is 30.223 ppm 95% confidence limits are 16.306 to 56.019ppm

Appendix Table CDXV: Cytotoxicity effect of *S. nodiflora* (wp/PetE) extracts against *A. salina* after 24h of exposure

Regression equation: $Y = 3.777 + 1.127X$ Chi-squared is 2.792 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.086ppm LC_{50} is 12.179ppm 95% confidence limits are 7.252 to 20.456ppm

Appendix Table CDXVI: Cytotoxicity effect of *S. nodiflora* (wp/PetE) extracts against *A. salina* after 30h of exposure

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Regression equation: $Y = 3.650 + 1.278X$ Chi-squared is 3.344 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.056ppm LC_{50} is 11.380ppm 95% confidence limits are 7.085 to 18.278ppm

Regression equation: $Y = 3.275 + 0.550 X$ Chi-squared is 0.525 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 3.137 ppm LC_{50} is 1372.014 ppm 95% confidence limits are 147.422 to 12768.92ppm

Appendix Table CDXIX: Cytotoxicity effect of *S. nodiflora* (wp/CHCl₃) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 2.692 + 1.066X$ Chi-squared is 10.902 with 4 degrees of freedom Variance has been adjusted for heterogeneity Log LC_{50} is 2.166 ppm LC_{50} is 146.676ppm 95% confidence limits are 66.514 to 323.450ppm

Appendix Table CDXVIII: Cytotoxicity effect of *S. nodiflora* (wp/CHCl₃) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 2.734 + 0.992X$ Chi-squared is 11.439 with 4 degrees of freedom Variance has been adjusted for heterogeneity Log LC_{50} is 2.285ppm LC_{50} is 192.840ppm 95% confidence limits are 74.018 to 502.408ppm

Appendix Table CDXX: Cytotoxicity effect of *S. nodiflora* (wp/CHCl₃) extracts against *A. salina* after 24h of exposure

Regression equation: $Y = 2.977 + 1.073X$ Chi-squared is 12.369 with 4 degrees of freedom Variance has been adjusted for heterogeneity Log LC_{50} is 1.8895 ppm LC_{50} is 76.688ppm 95% confidence limits are 36.959 to 159.121ppm

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Regression equation: $Y = 3.052 + 1.103X$

Chi-squared is 11.209 with 4 degrees of freedom Variance has been adjusted for heterogeneity Log LC_{50} is 1.765ppm LC_{50} is 58.268ppm 95% confidence limits are 29.406 to 115.461ppm

Appendix Table CDXXII: Cytotoxicity effect of *S. nodiflora* (wp/CH₃OH) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 1.899 + 0.873X$ Chi-squared is 1.515 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 3.550ppm LC_{50} is 3548.378ppm 95% confidence limits are 462.319 to 27234.43ppm

Appendix Table CDXXIII: Cytotoxicity effect of *S. nodiflora* (wp/CH₃OH) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 1.462 + 1.183X$ Chi-squared is 2.785 with 3 degrees of freedom No significant heterogeneity Log Lc_{50} is 2.990ppm Lc_{50} is 977.604ppm 95% confidence limits are 441.563 to 2164.381ppm

Appendix Table CDXXIV: Cytotoxicity effect of *S. nodiflora* (wp/CH₃OH) extracts against *A. salina* after 24h of exposure

Regression equation: $Y = 2.264 + 1.000X$ Chi-squared is 0.430 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.735ppm LC_{50} is 543.606ppm 95% confidence limits are 278.778 to 1060.010ppm

Regression equation: $Y = 2.008 + 1.304X$ Chi-squared is 1.320 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.295 ppm LC_{50} is 197.230 ppm 95% confidence limits are 135.584 to 286.905 ppm

Appendix Table CDXXVI: Cytotoxicity effect of *Z. zerumbet* (ap/PetE) extracts against *A. salina* after 6h of exposure

────────────────────────────────────────────────────────────────────── Regression equation: $Y = 3.156 + 0.793X$ Chi-squared is 0.421 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.325ppm LC₅₀ is 211.379ppm 95% confidence limits are 47.526 to 940.138ppm

Appendix Table CDXXVII: Cytotoxicity effect of *Z. zerumbet* (ap/PetE) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 3.030 + 1.385X$ Chi-squared is 0.464 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.422ppm LC_{50} is 26.447ppm 95% confidence limits are 18.517 to 37.772ppm

Appendix Table CDXXVIII: Cytotoxicity effect of *Z. zerumbet* (ap/PetE) extracts against *A. salina* after 18h of exposure

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Regression equation: $Y = 2.878 + 1.656X$ Chi-squared is 0.319 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.281ppm LC_{50} is 19.114ppm 95% confidence limits are 14.274 to 25.596ppm

Regression equation: $Y = 3.172 + 1.643X$ Chi-squared is 1.348 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.113ppm LC_{50} is 12.968ppm 95% confidence limits are 9.570 to 17.572ppm

Appendix Table CDXXXI: Cytotoxicity effect of *Z. zerumbet* (ap/CHCl₃) extracts against *A. salina* after 6h of exposure

Regression equation: $Y = 2.474 + 0.891X$ Chi-squared is 1.700 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.834 ppm LC_{50} is 682.473ppm 95% confidence limits are 121.602 to 3830.261ppm

Regression equation: $Y = 3.165 + 1.766X$ Chi-squared is 1.885 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.039ppm LC_{50} is 10.938ppm 95% confidence limits are 8.170 to 14.643ppm

Appendix Table CDLXXXII: Cytotoxicity effect of *Z. zerumbet* (ap/CHCl₃) extracts against *A. salina* after 12h of exposure

Dose				Ldos #U Kl %Kill Cr% E Pr Ex Pr Wk Pro Weght F Pro		
100.000		2.000 30 18 60.000 60 5.25 5.146			5.240 19.02 5.146	
50.000		1.699 30 13 43.333 43 4.82 4.804			4.838 18.81 4.826	
25.000		1.398 30 12 40.000 40 4.75 4.462			4.780 16.74 4.506	
12.500		1.097 30 4 13.333 13 3.87 4.120			3.904 14.13 4.187	

Regression equation: $Y = 3.022 + 1.062X$ Chi-squared is 2.930 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.863ppm LC_{50} is 72.906ppm 95% confidence limits are 45.200 to 117.596ppm

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Regression equation: $Y = 2.195 + 1.883X$ Chi-squared is 1.682 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.489ppm LC_{50} is 30.866ppm 95% confidence limits are 23.005 to 41.412ppm

Appendix Table CDXXXIV: Cytotoxicity effect of *Z. zerumbet* (ap/CHCl₃) extracts against *A. salina* after 24h of exposure

Regression equation: $Y = 2.721 + 1.636X$ Chi-squared is 1.376 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.393 ppm LC_{50} is 24.701ppm 95% confidence limits are 17.227 to 35.417ppm

Appendix Table CDXXXV: Cytotoxicity effect of *Z. zerumbet* (ap/CHCl₃) extracts against *A. salina* after 30h of exposure

Regression equation: $Y = 2.934 + 1.560X$ Chi-squared is 1.336 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.325ppm LC_{50} is 21.121 ppm 95% confidence limits are 14.056 to 31.738ppm

Appendix Table CDXXXVI: Cytotoxicity effect of *Z. zerumbet* (ap/CH₃OH) extracts against *A. salina* after 6h of exposure

Regression equation: $Y = 1.692 + 1.266X$ Chi-squared is 1.794 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.613ppm LC_{50} is 410.045ppm 95% confidence limits are 209.547 to 802.382ppm

Regression equation: $Y = 2.317 + 1.093X$ Chi-squared is 0.518 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.454ppm LC_{50} is 284.761ppm 95% confidence limits are 152.007 to 533.455ppm

Appendix Table CDXXXIX: Cytotoxicity effect of *Z. zerumbet* (ap/CH₃OH) extracts against *A. salina* after 24h of exposure

Regression equation: $Y = 2.111 + 1.403X$ Chi-squared is 0.368 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.059ppm LC_{50} is 114.558 ppm 95% confidence limits are 80.456 to 163.113ppm

Appendix Table CDXXXVIII: Cytotoxicity effect of *Z. zerumbet* (ap/CH3OH) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 2.072 + 1.353X$ Chi-squared is 0.761 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.164 ppm LC_{50} is 145.906ppm 95% confidence limits are 99.457 to 214.047ppm

Appendix Table CDXL: Cytotoxicity effect of *Z. zerumbet* (ap/CH₃OH) extracts against *A. salina* after 30h of exposure

Regression equation: $Y = 2.189 + 1.443X$ Chi-squared is 1.840 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.947996ppm LC_{50} is 88.715ppm 95% confidence limits are 62.754 to 125.415ppm

Regression equation: $Y = 3.279 + 0.836X$ Chi-squared is 0.158 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 2.060ppm LC_{50} is 114.818 ppm 95% confidence limits are 36.676 to 359.449ppm

Appendix Table CDXLII: Cytotoxicity effect of *Z. zerumbet* (rh/PetE) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 2.530 + 1.549X$ Chi-squared is 4.250 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.594ppm LC_{50} is 39.290ppm 95% confidence limits are 27.847 to 55.437ppm

Appendix Table CDXLIII: Cytotoxicity effect of *Z. zerumbet* (rh/PetE) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 2.735 + 1.618$ X Chi-squared is 1.317 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.400ppm LC_{50} is 25.115ppm 95% confidence limits are 18.424 to 34.236ppm

Appendix Table CDXLIV: Cytotoxicity effect of *Z. zerumbet* (rh/PetE) extracts against *A. salina* after 24h of exposure

Regression equation: $Y = 2.837 + 1.743X$ Chi-squared is 0.242 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.241ppm LC_{50} is 17.403ppm 95% confidence limits are 12.847 to 23.57ppm

Regression equation: $Y = 2.780 + 1.958X$ Chi-squared is 0.628 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.134 ppm LC_{50} is 13.608ppm 95% confidence limits are 10.116 to 18.305ppm

Appendix Table CDXLVI: Cytotoxicity effect of *Z. zerumbet* (rh/CHCl3) extracts against *A. salina* after 6h of exposure

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Regression equation: $Y = 2.509 + 1.236X$ Chi-squared is 1.835 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.015 ppm LC_{50} is 103.476 ppm 95% confidence limits are 52.005 to 205.892ppm

Appendix Table CDXLVII: Cytotoxicity effect of *Z. zerumbet* (rh/CHCl₃) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 1.783 + 2.195X$ Chi-squared is 2.117 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.465ppm LC_{50} is 29.198ppm 95% confidence limits are 22.887 to 37.249ppm

Appendix Table CDXLVIII: Cytotoxicity effect of *Z. zerumbet* (rh/CHCl₃) extracts against *A. salina* after 18h of exposure

Regression equation: $Y = 2.163 + 2.345X$ Chi-squared is 0.785 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.210ppm LC_{50} is 16.212 ppm 95% confidence limits are 12.730 to 20.647ppm

Regression equation: $Y = 2.637 + 2.124X$ Chi-squared is 0.464 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.113ppm LC_{50} is 12.963ppm 95% confidence limits are 9.802 to 17.144ppm

Appendix Table CDL: Cytotoxicity effect of *Z. zerumbet* (rh/CHCl₃) extracts against *A. salina* after 30h of exposure

────────────────────────────────────────────────────────────────────── Regression equation: $Y = 2.533 + 2.353X$

Chi-squared is 1.750 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.048 ppm LC_{50} is 11.172 ppm 95% confidence limits are 8.534 to 14.624ppm

Appendix Table CDLI: Cytotoxicity effect of *Z. zerumbet* (rh/CH₃OH) extracts against *A. salina* after 6h of exposure

Regression equation: $Y = 1.487 + 1.309X$ Chi-squared is 0.304 with 1 degrees of freedom No significant heterogeneity Log LC_{50} is 2.684 ppm LC_{50} is 482.507 ppm 95% confidence limits are 203.485 to 1144.128ppm

Appendix Table CDLII: Cytotoxicity effect of *Z. zerumbet* (rh/CH₃OH) extracts against *A. salina* after 12h of exposure

Regression equation: $Y = 1.295 + 1.465X$ Chi-squared is 1.546 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.530 ppm LC_{50} is 338.584ppm 95% confidence limits are 200.216 to 572.579ppm

Regression equation: $Y = 1.564 + 1.478X$ Chi-squared is 1.653 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.324ppm LC_{50} is 210.761ppm 95% confidence limits are 140.841 to 315.391ppm

Appendix Table CDLV: Cytotoxicity effect of *Z. zerumbet* (rh/CH₃OH) extracts against *A. salina* after 30h of exposure

Regression equation: $Y = 1.742 + 1.635X$ Chi-squared is 0.818 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.993ppm LC_{50} is 98.301ppm 95% confidence limits are 72.434 to 133.405ppm

Regression equation: $Y = 2.093 + 1.395X$ Chi-squared is 1.245 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.084 ppm LC_{50} is 121.463ppm 95% confidence limits are 84.756 to 174.067ppm

Appendix Table CDLVI: Larvicidal effect of *E. mummularius* (wp/PetE) extracts against *C. quinquefasciatus* after 6h of exposure

Dose				Ldos #U Kl %Kill Cr% E Pr Ex Pr Wk Pro Weght F Pro		
200.000				2.301 30 15 50.000 50 5.00 4.855 5.020 18.81 4.872		
100.000				2.000 30 4 13.333 13 3.87 4.222 3.912 15.09 4.231		
50.000		1.699 30 3 10.000 10 3.72 3.589			3.750 8.07 3.589	
25.000				1.398 30 1 3.333 3 3.12 2.956 3.172 3.30 2.948		

Regression equation: $Y = -0.031 + 2.131X$ Chi-squared is 2.318 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 2.361 ppm LC_{50} is 229.552ppm 95% confidence limits are 140.986 to 373.752ppm

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Regression equation: $Y = 1.133 + 1.836X$ Chi-squared is 4.537 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.106 ppm LC_{50} is 127.677ppm 95% confidence limits are 89.302 to 182.542ppm

Appendix Table CDLVIII: Larvicidal effect of *E. mummularius* (wp/PetE) extracts against *C. quinquefasciatus larvae* after 18h of exposure

Regression equation: $Y = 0.655 + 2.324X$ Chi-squared is 6.215 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.869 ppm LC_{50} is 74.044ppm 95% confidence limits are 58.328 to 93.996ppm

Appendix Table CDLIX: Larvicidal effect of *E. mummularius* (wp/PetE) extracts against *C. quinquefasciatus* after 24h of exposure

Regression equation: $Y = 1.546 + 1.910X$ Chi-squared is 6.371 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.809 ppm LC_{50} is 64.412 ppm 95% confidence limits are 48.891 to 84.861ppm

Appendix Table CDLX: Larvicidal effect of *E. mummularius* (wp/PetE) extracts against *C. quinquefasciatus* after 30h of exposure

Regression equation: $Y = 2.155 + 1.717X$ Chi-squared is 5.675 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.657ppm LC_{50} is 45.345ppm 95% confidence limits are 34.157 to 60.198ppm

Regression equation: $Y = 1.082 + 1.415X$ Chi-squared is 0.182 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.769ppm LC_{50} is 587.413ppm 95% confidence limits are 272.445 to 1266.508ppm

Appendix Table CDLXII: Larvicidal effect of *E. mummularius* (wp/CHCl₃) extracts against *C. quinquefasciatus* after 12h of exposure

Regression equation: $Y = 1.646 + 1.458X$ Chi-squared is 1.706 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.300ppm LC_{50} is 199.652ppm 95% confidence limits are 134.205 to 297.015ppm **Appendix Table CDLXIII:** Larvicidal effect of *E. mummularius* (wp/CHCl3) extracts against *C. quinquefasciatus* after 18h of exposure

Regression equation: $Y = 1.624 + 1.737X$ Chi-squared is 2.982 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.944 ppm LC_{50} is 87.868ppm 95% confidence limits are 66.398 to 116.280ppm

Appendix Table CDLXIV: Larvicidal effect of *E. mummularius* (wp/CHCl3) extracts against *C. quinquefasciatus* after 24h of exposure

Regression equation: $Y = 1.352 + 1.955X$ Chi-squared is 2.805 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.866ppm LC_{50} is 73.468ppm 95% confidence limits are 57.038 to 94.632ppm

Regression equation: $Y = 0.966 + 2.273X$ Chi-squared is 3.386 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.774ppm LC_{50} is 59.492ppm 95% confidence limits are 46.785 to 75.650ppm

Appendix Table CDLXVI: Larvicidal effect of *E. mummularius* (wp/CH₃OH) extracts against *C. quinquefasciatus* after 6h of exposure

Regression equation: $Y = 2.397 + 0.664X$ Chi-squared is 0.722 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 3.920ppm LC⁵⁰ is 8322.446ppm 95% confidence limits are 443.818 to 156062.1ppm **Appendix Table CDLXVII:** Larvicidal effect of *E. mummularius* (wp/CH₃OH) extracts against *C. quinquefasciatus* after 12h of exposure

Regression equation: $Y = 2.590 + 0.807X$ Chi-squared is 2.543 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.988ppm LC_{50} is 973.453ppm 95% confidence limits are 340.280 to 2784.796ppm

Appendix Table CDLXVIII: Larvicidal effect of *E. mummularius* (wp/CH₃OH) extracts against *C. quinquefasciatus* after 18h of exposure

Regression equation: $Y = 3.003 + 0.733X$ Chi-squared is 0.543 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.724ppm LC_{50} is 529.690ppm 95% confidence limits are 221.355 to 1267.520ppm

Regression equation: $Y = 2.503 + 1.060X$ Chi-squared is 0.937 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.356 ppm LC_{50} is 226.776ppm 95% confidence limits are 145.669 to 353.042ppm

Appendix Table CDLXXI: Larvicidal effect of *L. camara* (ap/PetE) extracts against *C. quinquefasciatus* after 6h of exposure

Regression equation: $Y = 2.283 + 1.281X$ Chi-squared is 0.888 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 2.121 ppm LC_{50} is 132.032 ppm 95% confidence limits are 56.850 to 306.639ppm

Appendix Table CDLXX: Larvicidal effect of *E. mummularius* (wp/CH₃OH) extracts against *C. quinquefasciatus* after 30h of exposure

Regression equation: $Y = 2.263 + 1.269X$ Chi-squared is 1.880 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.157ppm LC_{50} is 143.461 ppm 95% confidence limits are 100.061 to 205.686ppm

Appendix Table CDLXXII: Larvicidal effect of *L. camara* (ap/PetE) extracts against *C. quinquefasciatus* after 12h of exposure

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Regression equation: $Y = 2.563 + 1.359X$ Chi-squared is 1.252 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.793ppm LC_{50} is 62.076ppm 95% confidence limits are 38.717 to 99.527ppm

Regression equation: $Y = 2.579 + 1.480X$ Chi-squared is 1.739 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.636ppm LC_{50} is 43.208ppm 95% confidence limits are 29.833 to 62.578ppm

Appendix Table CDLXXIV: Larvicidal effect of *L. camara* (ap/PetE) extracts against *C. quinquefasciatus* after 24h of exposure

────────────────────────────────────────────────────────────────────── Regression equation: $Y = 2.857 + 1.456X$ Chi-squared is 2.831 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.471ppm LC_{50} is 29.608ppm 95% confidence limits are 20.958 to 41.828ppm

Appendix Table CDLXXV: Larvicidal effect of *L. camara* (ap/PetE) extracts against *C. quinquefasciatus* after 30h of exposure

Regression equation: $Y = 3.000 + 1.469X$ Chi-squared is 2.653 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.361ppm LC_{50} is 22.950ppm 95% confidence limits are 16.330 to 32.252ppm

Appendix Table CDLXXVI: Larvicidal effect of *L. camara* (ap/CHCl3) extracts against *C. quinquefasciatus* after 6h of exposure

────────────────────────────────────────────────────────────────────── Regression equation: $Y = 2.563 + 0.840X$ Chi-squared is 0.238 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.902ppm LC_{50} is 797.276ppm 95% confidence limits are 199.363 to 3188.403ppm

Regression equation: $Y = 2.915 + 0.767X$ Chi-squared is 0.632 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.720ppm LC_{50} is 525.043ppm 95% confidence limits are 154.293 to 1786.669ppm

Appendix Table CDLXXVIII: Larvicidal effect of *L. camara* (ap/CHCl₃) extracts against *C. quinquefasciatus* after 18h of exposure

Regression equation: $Y = 2.679 + 1.014X$ Chi-squared is 2.212 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.289ppm LC50 is 194.735ppm 95% confidence limits are 111.424 to 340.337ppm

Appendix Table CDLXXIX: Larvicidal effect of *L. camara* (ap/CHCl₃) extracts against *C. quinquefasciatus* after 24h of exposure

Regression equation: $Y = 2.361 + 1.418X$ Chi-squared is 4.434 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.861ppm LC_{50} is 72.554ppm 95% confidence limits are 50.510 to 104.218ppm

Appendix Table CDLXXX: Larvicidal effect of *L. camara* (ap/CHCl₃) extracts against *C. quinquefasciatus* after 30h of exposure

Regression equation: $Y = 2.129 + 1.635X$ Chi-squared is 2.450 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.756ppm LC_{50} is 57.024ppm 95% confidence limits are 40.443 to 80.403ppm

Regression equation: $Y = -1.547 + 2.728X$ Chi-squared is 0.022 with 1 degrees of freedom No significant heterogeneity

Log LC_{50} is 2.400 ppm LC_{50} is 251.201ppm

95% confidence limits are 157.531 to 400.568ppm

Appendix Table CDLXXXII: Larvicidal effect of *L. camara* (ap/CH₃OH) extracts against *C. quinquefasciatus* after 12h of exposure

────────────────────────────────────────────────────────────────────── Regression equation: $Y = 0.882 + 1.843X$ Chi-squared is 1.114 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.234ppm

 LC_{50} is 171.323ppm

95% confidence limits are 111.619 to 262.962ppm

Appendix Table CDLXXXIII: Larvicidal effect of *L. camara* (ap/CH3OH) extracts against *C. quinquefasciatus* after 18h of exposure

Regression equation: $Y = 1.315 + 1.939X$ Chi-squared is 1.659 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.900ppm LC_{50} is 79.484ppm 95% confidence limits are 59.951 to 105.382ppm

Appendix Table CDLXXXIV: Larvicidal effect of *L. camara* (ap/CH₃OH) extracts against *C. quinquefasciatus* after 24h of exposure

Regression equation: $Y = 1.471 + 2.185X$ Chi-squared is 1.738 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.615ppm LC_{50} is 41.211 ppm 95% confidence limits are 32.159 to 52.811ppm

Regression equation: $Y = 1.422 + 2.288X$ Chi-squared is 1.919 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.564ppm LC_{50} is 36.658ppm 95% confidence limits are 28.523 to 47.115ppm

Appendix Table CDLXXXVI: Larvicidal effect of *L. camara* (r/PetE) extracts against *C. quinquefasciatus* after 6h of exposure

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Regression equation: $Y = -0.445 + 2.768X$ Chi-squared is 1.399 with 1 degrees of freedom

No significant heterogeneity

Log LC_{50} is 1.967ppm

 LC_{50} is 92.666ppm

95% confidence limits are 66.876 to 128.403ppm

Appendix Table CDLXXXVII: Larvicidal effect of *L. camara* (r/PetE) extracts against *C. quinquefasciatus* after 12h of exposure

Regression equation: $Y = 2.042 + 1.584X$ Chi-squared is 1.297 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.867ppm LC_{50} is 73.666ppm 95% confidence limits are 47.125 to 115.153ppm

Appendix Table CDLXXXVIII: Larvicidal effect of *L. camara* (r/PetE) extracts against *C. quinquefasciatus* after 18h of exposure

Regression equation: $Y = 1.762 + 1.984X$ Chi-squared is 1.040 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.632ppm LC_{50} is 42.835 ppm 95% confidence limits are 32.238 to 56.917ppm

Regression equation: $Y = 1.642 + 2.221X$ Chi-squared is 3.341 with 3 egrees of freedom No significant heterogeneity Log LC_{50} is 1.512ppm LC_{50} is 32.517ppm 95% confidence limits are 25.437 to 41.567ppm

Appendix Table CDXC: Larvicidal effect of *L. camara* (r/PetE) extracts against *C. quinquefasciatus* after 30h of exposure

Regression equation: $Y = 2.314 + 1.846X$

Chi-squared is 0.333 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.455ppm LC_{50} is 28.532ppm 95% confidence limits are 20.330 to 40.043ppm

Appendix Table CDXCI: Larvicidal effect of *L. camara* (r/CHCl₃) extracts against *C. quinquefasciatus* after 6h of exposure

Regression equation: $Y = 1.249 + 1.120X$ Chi-squared is 0.245 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 3.350 ppm LC_{50} is 2238.424ppm 95% confidence limits are 584.642 to 8570.274ppm

Appendix Table CDXCII: Larvicidal effect *L. camara* (r/CHCl₃) extracts against *C. quinquefasciatus* after 12h of exposure

Regression equation: $Y = 1.390 + 1.312X$ Chi-squared is 0.289 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.752 ppm LC_{50} is 565.086ppm 95% confidence limits are 333.691 to 956.937ppm

Regression equation: $Y = 1.933 + 1.161X$ Chi-squared is 1.351 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.641ppm LC_{50} is 438.020ppm 95% confidence limits are 261.029 to 735.022ppm

Appendix Table CDXCIV: Larvicidal effect of *L. camara* (r/CHCl3) extracts against *C. quinquefasciatus* after 24h of exposure

Regression equation: $Y = 1.688 + 1.323X$ Chi-squared is 1.960 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.503 ppm LC_{50} is 318.277ppm 95% confidence limits are 213.313 to 474.891ppm

Appendix Table CDXCV: Larvicidal effect of *L. camara* (r/CHCl3) extracts against *C. quinquefasciatus* after 30h of exposure

Regression equation: $Y = 1.316 + 1.604X$ Chi-squared is 1.911 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.297ppm LC_{50} is 198.309ppm 95% confidence limits are 144.549 to 272.064ppm

Appendix Table CDXCVI: Larvicidal effect of *L. camara* (r/CH3OH) extracts against *C. quinquefasciatus* after 6h of exposure

Regression equation: $Y = 1.927 + 1.004X$ Chi-squared is 0.659 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 3.059 ppm LC_{50} is 1146.454 ppm 95% confidence limits are 90.833 to 14469.99ppm

Regression equation: $Y = 1.400 + 1.629X$ Chi-squared is 0.836 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.211ppm LC_{50} is 162.446ppm 95% confidence limits are 102.209 to 258.182ppm

Appendix Table CDXCVIII: Larvicidal effect of *L. camara* (r/CH₃OH) extracts against *C. quinquefasciatus* after 18h of exposure

Regression equation: $Y = 1.818 + 1.630X$ Chi-squared is 0.409 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.952ppm LC_{50} is 89.496ppm 95% confidence limits are 63.497 to 126.141ppm

Appendix Table CDXCIX: Larvicidal effect of *L. camara* (r/CH₃OH) extracts against *C. quinquefasciatus* after 24h of exposure

Regression equation: $Y = 1.678 + 1.915X$ Chi-squared is 1.474 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.735ppm LC_{50} is 54.335ppm 95% confidence limits are 41.354 to 71.392ppm

Appendix Table D: Larvicidal effect of *L. camara* (r/CH₃OH) extracts against *C. quinquefasciatus* after 30h of exposure

Regression equation: $Y = 1.802 + 1.960X$ Chi-squared is 0.381 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.632ppm LC_{50} is 42.811 ppm 95% confidence limits are 32.019 to 57.242ppm

Appendix Table DI: Larvicidal effect of *M. piperita* (wp/PetE) extracts against *C. quinquefasciatus* after 6h of exposure

Appendix Table DII: Larvicidal effect of *M. piperita* (wp/PetE) extracts against *C. quinquefasciatus* after 12h of exposure

────────────────────────────────────────────────────────────────────── Regression equation: $Y = 1.305 + 1.795X$ Chi-squared is 0.206 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 2.058ppm LC_{50} is 114.399 ppm 95% confidence limits are 80.711 to 162.149ppm

Appendix Table DIII: Larvicidal effect of *M. piperita* (wp/PetE) extracts against *C. quinquefasciatus* after 18h of exposure

Regression equation: $Y = 1.782 + 1.692X$ Chi-squared is 0.620 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.902ppm LC_{50} is 79.776ppm 95% confidence limits are 57.825 to 110.059ppm

Appendix Table DIV: Larvicidal effect of *M. piperita* (wp/PetE) extracts against *C. quinquefasciatus* after 24h of exposure

Regression equation: $Y = 1.917 + 1.730X$ Chi-squared is 2.819 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.782ppm LC_{50} is 60.519 ppm 95% confidence limits are 44.858 to 81.648ppm

Regression equation: $Y = 1.808 + 1.919X$ Chi-squared is 4.127 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.663ppm LC_{50} is 46.065ppm 95% confidence limits are 35.055 to 60.533ppm

Appendix Table DVI: Larvicidal effect of *M. piperita* (wp/CHCl₃) extracts against *C. quinquefasciatus* after 6h of exposure

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Regression equation: $Y = -1.171 + 2.353X$

Chi-squared is 3.037 with 1 degrees of freedom No significant heterogeneity

Log LC_{50} is 2.622 ppm

 LC_{50} is 418.844 ppm

95% confidence limits are 272.140 to 644.633ppm

Appendix Table DVII: Larvicidal effect of *M. piperita* (wp/CHCl3) extracts against *C. quinquefasciatus* after 12h of exposure

Appendix Table DVIII: Larvicidal effect of *M. piperita* (wp/CHCl₃) extracts against *C. quinquefasciatus* after 18h of exposure

Regression equation: $Y = 0.111 + 2.160X$ Chi-squared is 2.122 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 2.263 ppm LC50 is 183.390ppm 95% confidence limits are 139.801 to 240.569ppm

Regression equation: $Y = 0.175 + 2.279X$ Chi-squared is 2.705 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.117ppm LC_{50} is 130.911ppm 95% confidence limits are 102.806 to 166.700ppm

Appendix Table DX: Larvicidal effect of *M. piperita* (wp/CHCl₃) extracts against *C. quinquefasciatus* after 30h of exposure

Regression equation: $Y = 1.680 + 1.670X$ Chi-squared is 0.130 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.988 ppm

- LC_{50} is 97.375ppm
- 95% confidence limits are 68.799 to 137.821ppm

Appendix Table DXI: Larvicidal effect of *M. piperita* (wp/CH₃OH) extracts against *C. quinquefasciatus* after 6h of exposure

Regression equation: $Y = -1.285 + 2.042X$ Chi-squared is 1.396 with 1 degrees of freedom No significant heterogeneity Log LC_{50} is 3.077 ppm LC_{50} is 1195.342ppm 95% confidence limits are 589.974 to 2421.874ppm

Appendix Table DXII: Larvicidal effect of *M. piperita* (wp/CH₃OH) extracts against *C. quinquefasciatus* after 12h of exposure

Regression equation: $Y = -0.389 + 1.778X$ Chi-squared is 1.473 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 3.0315 ppm LC_{50} is 1075.135ppm 95% confidence limits are 562.391 to 2055.359ppm

Regression equation: $Y = 0.087 + 1.728X$ Chi-squared is 0.134 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 2.843 ppm LC_{50} is 696.966ppm 95% confidence limits are 434.749 to 1117.337ppm

Appendix Table DXIV: Larvicidal effect of *M. piperita* (wp/CH₃OH) extracts against *C. quinquefasciatus* after 24h of exposure

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Regression equation: $Y = 0.590 + 1.625X$ Chi-squared is 0.760 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.713 ppm LC_{50} is 516.804ppm 95% confidence limits are 344.759 to 774.703ppm

Appendix Table DXV: Larvicidal effect of *M. piperita* (wp/CH₃OH) extracts against *C. quinquefasciatus* after 30h of exposure

Chi-squared is 1.209 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.491ppm LC_{50} is 309.859ppm 95% confidence limits are 235.467 to 407.753ppm

Appendix Table DXVI: Larvicidal effect of *Mi. pudica* (wp/PetE) extracts against *C. quinquefasciatus* after 6h of exposure

Regression equation: $Y = 0.556 + 1.745X$ Chi-squared is 0.411 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.547 ppm LC_{50} is 352.364ppm 95% confidence limits are 223.195 to 556.286ppm

Regression equation: $Y = 1.225 + 1.581X$ Chi-squared is 0.420 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.387ppm LC_{50} is 243.844ppm 95% confidence limits are 162.579 to 365.729ppm

Appendix Table DXVIII: Larvicidal effect of *Mi. pudica* (wp/PetE) extracts against *C. quinquefasciatus* after 18h of exposure

────────────────────────────────────────────────────────────────────── Regression equation: $Y = 1.705 + 1.452X$ Chi-squared is 2.040 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.269ppm LC_{50} is 185.822ppm 95% confidence limits are 125.898 to 274.267ppm

Appendix Table DXIX: Larvicidal effect of *Mi. pudica* (wp/PetE) extracts against *C. quinquefasciatus* after 24h of exposure

Chi-squared is 0.565 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.075ppm LC_{50} is 118.721 ppm 95% confidence limits are 84.386 to 167.026ppm

Appendix Table DXX: Larvicidal effect of *Mi. pudica* (wp/PetE) extracts against *C. quinquefasciatus* after 30h of exposure

────────────────────────────────────────────────────────────────────── Regression equation: $Y = 2.145 + 1.467X$ Chi-squared is 1.444 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.945ppm LC_{50} is 88.187ppm 95% confidence limits are 62.719 to 123.996ppm

95% confidence limits are 600.263 to 2237.904ppm

Appendix Table DXXI: Larvicidal effect of *Mi. pudica* (wp/CHCl3) extracts against *C. quinquefasciatus* after 6h of exposure

Appendix TableDXXIII: Larvicidal effect of *Mi. pudica* (wp/CHCl₃) extracts against *C. quinquefasciatus* after 18h of exposure

Regression equation: $Y = 1.309 + 1.287X$ Chi-squared is 2.218 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.869 ppm LC_{50} is 739.454ppm 95% confidence limits are 397.209 to 1376.586ppm

Appendix Table DXXII: Larvicidal effect of *Mi. pudica* (wp/CHCl₃) extracts against *C. quinquefasciatus* after 12h of exposure

Regression equation: $Y = 1.252 + 1.197X$ Chi-squared is 0.744 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 3.132 ppm LC_{50} is 1356.547ppm 95% confidence limits are 471.550 to 3902.496ppm **Appendix Table DXXIV:** Larvicidal effect of *Mi. pudica* (wp/CHCl₃) extracts against *C. quinquefasciatus* after 24h of exposure

Regression equation: $Y = 1.525 + 1.309X$ Chi-squared is 0.823 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.655ppm LC_{50} is 452.226ppm 95% confidence limits are 283.249 to 722.009ppm

Regression equation: $Y = 0.937 + 1.663X$ Chi-squared is 1.068 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.443ppm LC_{50} is 277.609ppm 95% confidence limits are 202.384 to 380.795ppm

Appendix Table DXXVII: Larvicidal effect of *P. hysterophorus* (wp/PetE) extracts against *C. quinquefasciatus* after 12h of exposure

Regression equation: $Y = 0.269 + 2.556X$ Chi-squared is 6.875 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.851ppm LC_{50} is 71.013ppm 95% confidence limits are 56.834 to 88.728ppm

Appendix Table DXXVI: Larvicidal effect of *P. hysterophorus* (wp/PetE) extracts against *C. quinquefasciatus* after 6h of exposure

Regression equation: $Y = 0.190 + 2.373X$ Chi-squared is 1.765 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 2.027ppm LC_{50} is 106.419ppm 95% confidence limits are 81.974 to 138.155ppm

Appendix Table DXXVIII: Larvicidal effect of *P. hysterophorus* (wp/PetE) extracts against *C. quinquefasciatus* after 18h of exposure

Regression equation: $Y = 2.133 + 1.554X$ Chi-squared is 1.253 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.845ppm LC_{50} is 69.907ppm 95% confidence limits are 44.616 to 109.534ppm

Regression equation: $Y = 2.494 + 1.507X$ Chi-squared is 0.513 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.663ppm LC_{50} is 46.054ppm 95% confidence limits are 31.609 to 67.101ppm

Appendix Table DXXX: Larvicidal effect of *P. hysterophorus* (wp/PetE) extracts against *C. quinquefasciatus* after 30h of exposure

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Regression equation: $Y = 2.386 + 1.657X$ Chi-squared is 0.509 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.578ppm LC_{50} is 37.858ppm

95% confidence limits are 27.206 to 52.681ppm

Appendix Table DXXXI: Larvicidal effect of *P. hysterophorus* (wp/CHCl₃) extracts against *C. quinquefasciatus* after 6h of exposure

Regression equation: $Y = 0.284 + 1.701X$ Chi-squared is 0.068 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.773 ppm LC_{50} is 593.445ppm 95% confidence limits are 389.409 to 904.389ppm

Appendix Table DXXXII: Larvicidal effect of *P. hysterophorus* (wp/CHCl3) extracts against *C. quinquefasciatus* after 12h of exposure

Regression equation: $Y = -0.329 + 2.102X$ Chi-squared is 1.247 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.536ppm LC_{50} is 343.456 ppm 95% confidence limits are 261.438 to 451.205ppm **Appendix Table DXXXIII:** Larvicidal effect of *P. hysterophorus* (wp/CHCl3) extracts against *C. quinquefasciatus* after 18h of exposure

Regression equation: $Y = 0.591 + 1.776X$ Chi-squared is 2.622 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 2.483ppm LC_{50} is 303.972ppm 95% confidence limits are 200.183 to 461.573ppm

Appendix Table DXXXV: Larvicidal effect of *P. hysterophorus* (wp/CHCl₃) extracts against *C. quinquefasciatus* after 30h of exposure

 LC_{50} is 131.807 ppm

95% confidence limits are 101.520 to 171.129ppm

Appendix Table DXXXIV: Larvicidal effect of *P. hysterophorus* (wp/CHCl3) extracts against *C. quinquefasciatus* after 24h of exposure

Regression equation: $Y = 1.049 + 1.700X$ Chi-squared is 1.570 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 2.324 ppm LC_{50} is 211.057 ppm 95% confidence limits are 148.368 to 300.235ppm

Appendix Table DXXXVI: Larvicidal effect of *P. hysterophorus* (wp/CH₃OH) extracts against *C. quinquefasciatus* after 6h of exposure

Regression equation: $Y = 1.439 + 1.498X$ Chi-squared is 2.437 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 2.377 ppm LC_{50} is 237.973ppm 95% confidence limits are 120.781 to 468.876ppm

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Regression equation: $Y = 2.005 + 1.407X$ Chi-squared is 0.287 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 2.129 ppm LC_{50} is 134.598 ppm 95% confidence limits are 83.031 to 218.190ppm

Appendix Table DXXXVIII: Larvicidal effect of *P. hysterophorus* (wp/CH₃OH) extracts against *C. quinquefasciatus* after 18h of exposure

Regression equation: $Y = 2.150 + 1.469X$ Chi-squared is 0.212 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.940ppm LC_{50} is 87.056ppm 95% confidence limits are 59.845 to 126.639ppm

Appendix Table DXXXIX: Larvicidal effect of *P. hysterophorus* (wp/CH3OH) extracts against *C. quinquefasciatus* after 24h of exposure

Regression equation: $Y = 1.892 + 1.827X$ Chi-squared is 0.078 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.701ppm LC_{50} is 50.229ppm 95% confidence limits are 37.872 to 66.618ppm

Appendix Table DXL: Larvicidal effect of *P. hysterophorus* (wp/CH₃OH) extracts against *C. quinquefasciatus* after 30h of exposure

Regression equation: $Y = 1.421 + 2.261X$ Chi-squared is 0.999 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.583ppm LC_{50} is 38.287 ppm 95% confidence limits are 29.676 to 49.396ppm

Regression equation: $Y = 2.294 + 1.224X$ Chi-squared is 1.390 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.210ppm LC_{50} is 162.346ppm 95% confidence limits are 64.957 to 405.746ppm

Appendix Table DXLII: Larvicidal effect of *Ph. niruri* (wp/PetE) extracts against *C. quinquefasciatus* after 12h of exposure

Regression equation: $Y = 2.585 + 1.356X$ Chi-squared is 2.411 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.781ppm LC_{50} is 60.402ppm 95% confidence limits are 38.045 to 95.898ppm

Appendix Table DXLIII: Larvicidal effect of *Ph. niruri* (wp/PetE) extracts against *C. quinquefasciatus* after 18h of exposure

Regression equation: $Y = 3.345 + 1.189X$ Chi-squared is 2.215 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.392 ppm LC_{50} is 24.680ppm 95% confidence limits are 16.716 to 36.438ppm

Appendix Table DXLIV: Larvicidal effect of *Ph. niruri* (wp/PetE) extracts against *C. quinquefasciatus* after 24h of exposure

────────────────────────────────────────────────────────────────────── Regression equation: $Y = 3.945 + 1.122X$ Chi-squared is 0.486 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 0.940ppm LC_{50} is 8.707 ppm 95% confidence limits are 5.523 to 13.729ppm

Regression equation: $Y = 4.252 + 1.566X$ Chi-squared is 0.305 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 0.477ppm LC_{50} is 3.390ppm bh 95% confidence limits are 1.672 to 5.391ppm

Appendix Table DXLVI: Larvicidal effect of *Ph. niruri* (wp/CHCl₃) extracts against *C. quinquefasciatus* after 6h of exposure

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Regression equation: $Y = 2.158 + 1.120X$ Chi-squared is 0.889 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.537 ppm LC_{50} is 344.236ppm 95% confidence limits are 124.367 to 952.811ppm

Appendix Table DXLVII: Larvicidal effect of *Ph. niruri* (wp/CHCl₃) extracts against *C. quinquefasciatus* after 12h of exposure

Regression equation: $Y = 2.453 + 1.234X$ Chi-squared is 1.086 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.064ppm LC_{50} is 115.823ppm 95% confidence limits are 70.603 to 190.005ppm

Appendix Table DXLVIII: Larvicidal effect of *Ph. niruri* (wp/CHCl₃) extracts against *C. quinquefasciatus* after 18h of exposure

Regression equation: $Y = 3.201 + 1.125X$ Chi-squared is 0.914 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.599ppm LC_{50} is 39.697ppm 95% confidence limits are 26.645 to 59.142ppm

Regression equation: $Y = 3.977 + 1.011X$ Chi-squared is 0.305 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.012ppm LC_{50} is 10.284ppm 95% confidence limits are 5.507 to 19.206ppm

Appendix Table DL: Larvicidal effect of *Ph. niruri* (wp/CHCl₃) extracts against *C. quinquefasciatus* after 30h of exposure

Regression equation: $Y = 4.456 + 1.072X$

Chi-squared is 0.585 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 0.508 ppm LC_{50} is 3.220ppm 95% confidence limits are 0.925 to 11.196ppm

Appendix Table DLI: Larvicidal effect of *Ph. niruri* (wp/CH₃OH) extracts against *C. quinquefasciatus* after 6h of exposure

Regression equation: $Y = -5.367 + 3.234X$ Chi-squared is 0.121 with 1 degrees of freedom No significant heterogeneity Log LC_{50} is 3.205 ppm LC_{50} is 1603.863ppm 95% confidence limits are 551.099 to 4667.72ppm

Appendix Table DLII: Larvicidal effect of *Ph. niruri* (wp/CH₃OH) extracts against *C. quinquefasciatus* after 12h of exposure

Regression equation: $Y = -1.687 + 2.156X$ Chi-squared is 0.408 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 3.101 ppm LC_{50} is 1262.913 ppm 95% confidence limits are 648.058 to 2461.124ppm

No significant heterogeneity

- Log LC_{50} is 2.968ppm
- LC_{50} is 928.145ppm
- 95% confidence limits are 614.832 to 1401.117ppm

Appendix Table DLIV: Larvicidal effect of *Ph. niruri* (wp/CH₃OH) extracts against *C. quinquefasciatus* after 24h of exposure

Regression equation: $Y = -0.800 + 2.100X$ Chi-squared is 1.768 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.762 ppm LC_{50} is 577.739ppm 95% confidence limits are 437.340 to 763.209ppm

Appendix Table DLV: Larvicidal effect of *Ph. niruri* (wp/CH₃OH) extracts against *C. quinquefasciatus* after 30h of exposure

Chi-squared is 4.497 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.415 ppm LC_{50} is 259.864ppm 95% confidence limits are 206.685 to 326.728ppm

Appendix Table DLVI: Larvicidal effect of *Po. hydropiper* (wp/PetE) extracts against *C. quinquefasciatus* after 6h of exposure

Regression equation: $Y = 2.532 + 1.103X$ Chi-squared is 1.193 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.237 ppm LC_{50} is 172.630ppm 95% confidence limits are 61.302 to 486.138ppm

Regression equation: $Y = 2.224 + 1.482X$ Chi-squared is 0.451 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.873ppm LC_{50} is 74.608ppm 95% confidence limits are 46.201 to 120.481ppm

Regression equation: $Y = 2.239 + 1.549X$ Chi-squared is 1.829 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.782ppm LC_{50} is 60.534 ppm 95% confidence limits are 40.12327 to 91.328ppm **Appendix Table DLIX:** Larvicidal effect of *Po. hydropiper* (wp/PetE) extracts against *C. quinquefasciatus* after 24h of exposure

Regression equation: $Y = 2.265 + 1.684X$ Chi-squared is 2.617 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.624ppm LC_{50} is 42.100ppm 95% confidence limits are 30.382 to 58.337ppm

Appendix Table DLX: Larvicidal effect of *Po. hydropiper* (wp/PetE) extracts against *C. quinquefasciatus* after 30h of exposure

Regression equation: $Y = 2.366 + 1.928X$ Chi-squared is 2.438 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.366ppm LC⁵⁰ is 23.246ppm 95% confidence limits are 17.728 to 30.483ppm

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Regression equation: $Y = 2.697 + 0.705X$ Chi-squared is 0.263 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 3.267 ppm LC_{50} is 1851.189 ppm 95% confidence limits are 126.670 to 27053.9ppm

Appendix Table DLXII: Larvicidal effect of *Po. hydropiper* (wp/CHCl₃) extracts against *C. quinquefasciatus* after 12h of exposure

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Regression equation: $Y = 2.349 + 1.114X$ Chi-squared is 1.355 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.380ppm LC_{50} is 240.028ppm 95% confidence limits are 109.256 to 527.321ppm

Appendix Table DLXIII: Larvicidal effect of *Po. hydropiper* (wp/CHCl₃) extracts against *C. quinquefasciatus* after 18h of exposure

Regression equation: $Y = 2.743 + 1.032X$ Chi-squared is 1.769 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.186 ppm LC_{50} is 153.594ppm 95% confidence limits are 78.414 to 300.853ppm

Appendix Table DLXIV: Larvicidal effect of *Po. hydropiper* (wp/CHCl₃) extracts against *C. quinquefasciatus* after 24h of exposure

Regression equation: $Y = 3.014 + 1.031X$ Chi-squared is 2.181 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.926ppm LC_{50} is 84.288 ppm 95% confidence limits are 50.332 to 141.150ppm

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Regression equation: $Y = 2.695 + 1.401X$ Chi-squared is 2.560 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.645 ppm LC_{50} is 44.154 ppm 95% confidence limits are 31.636 to 61.625ppm

Appendix Table DLXVI: Larvicidal effect of *Po. hydropiper* (wp/CH₃OH) extracts against *C. quinquefasciatus* after 6h of exposure

Regression equation: $Y = 2.335 + 1.029X$ Chi-squared is 0.286 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.589ppm LC_{50} is 388.521 ppm 95% confidence limits are 133.247 to 1132.844ppm **Appendix Table DLXVII:** Larvicidal effect of *Po. hydropiper* (wp/CH₃OH) extracts against *C. quinquefasciatus* after 12h of exposure

Regression equation: $Y = 2.423 + 1.028X$ Chi-squared is 0.799 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.506 ppm LC_{50} is 320.718ppm 95% confidence limits are 121.140 to 849.101ppm

Appendix Table DLXVIII: Larvicidal effect of *Po. hydropiper* (wp/CH₃OH) extracts against *C. quinquefasciatus* after 18h of exposure

Regression equation: $Y = 3.002 + 0.765X$ Chi-squared is 0.956 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.612ppm LC_{50} is 408.946 ppm 95% confidence limits are 103.401 to 1617.363ppm

Regression equation: $Y = 4.006 + 0.461X$ Chi-squared is 0.596 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.157 ppm LC_{50} is 143.576ppm 95% confidence limits are 34.461 to 598.190ppm

Appendix Table DLXXI: Larvicidal effect of *Pz. zeylanica* (wp/PetE) extracts against *C. quinquefasciatus* after 6h of exposure

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Regression equation: $Y = 1.564 + 0.949X$

Chi-squared is 0.068 with 1 degrees of freedom

No significant heterogeneity

Log LC_{50} is 3.621 ppm

 LC_{50} is 4175.449 ppm

95% confidence limits are 4.825 to 3613104 ppm

Appendix Table DLXX: Larvicidal effect of *Po. hydropiper* (wp/CH₃OH) extracts against *C. quinquefasciatus* after 30h of exposure

Regression equation: $Y = 4.165 + 0.628X$ Chi-squared is 1.939 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.331ppm LC_{50} is 21.432ppm 95% confidence limits are 10.213 to 44.978ppm

Appendix Table DLXXII: Larvicidal effect of *Pz. zeylanica* (wp/PetE) extracts against *C. quinquefasciatus* after 12h of exposure

────────────────────────────────────────────────────────────────────── Regression equation: $Y = 1.410 + 1.285X$ Chi-squared is 0.3743358 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 2.793ppm LC_{50} is 620.466ppm 95% confidence limits are 133.242 to 2889.315ppm

α **Appendices** \Box **IES, RU**

Regression equation: $Y = 0.958 + 1.678X$ Chi-squared is 0.4213114 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 2.408 ppm LC_{50} is 256.030ppm

95% confidence limits are 133.495 to 491.042ppm

Appendix Table DLXXIV: Larvicidal effect of *Pz. zeylanica* (wp/PetE) extracts against *C. quinquefasciatus* after 24h of exposure

Regression equation: $Y = 2.394 + 1.377X$ Chi-squared is 0.301 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.893ppm LC_{50} is 78.121ppm 95% confidence limits are 53.341 to 114.414ppm **Appendix Table DLXXV:** Larvicidal effect of *Pz. zeylanica* (wp/PetE) extracts against *C. quinquefasciatus* after 30h of exposure

Regression equation: $Y = 2.927 + 1.497X$ Chi-squared is 0.143 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.384ppm LC_{50} is 24.226ppm 95% confidence limits are 17.392 to 33.746ppm

Appendix Table DLXXVI: Larvicidal effect of *Pz. zeylanica* (wp/CHCl₃) extracts against *C. quinquefasciatus* after 6h of exposure

Regression equation: $Y = 2.758 + 0.648X$ Chi-squared is 1.313 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 3.459ppm LC_{50} is 2878.285ppm 95% confidence limits are 219.310 to 37775.37ppm

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Regression equation: $Y = 2.481 + 1.004X$ Chi-squared is 1.143 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.510 ppm LC_{50} is 323.224ppm 95% confidence limits are 159.599 to 654.602ppm

Appendix Table DLXXIX: Larvicidal effect of *Pz. zeylanica* (wp/CHCl₃) extracts against *C. quinquefasciatus* after 24h of exposure

Regression equation: $Y = 2.325 + 1.342X$ Chi-squared is 3.139 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.993 ppm LC_{50} is 98.301ppm 95% confidence limits are 69.327 to 139.382ppm

Appendix Table DLXXVIII: Larvicidal effect of *Pz. zeylanica* (wp/CHCl₃) extracts against *C. quinquefasciatus* after 18h of exposure

────────────────────────────────────────────────────────────────────── Regression equation: $Y = 2.406 + 1.234X$ Chi-squared is 0.756 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.101 ppm LC_{50} is 126.241ppm 95% confidence limits are 84.859 to 187.804ppm

Appendices CXLV IES, RU

Appendix Table DLXXX: Larvicidal effect of *Pz. zeylanica* (wp/CHCl₃) extracts against *C. quinquefasciatus* after 30h of exposure

Dose	Ldos						#U Kl %Kill Cr% E Pr Ex Pr Wk Pro Weght F Pro		
400.000				2.602 30 27 90.000 90 6.28 6.058				6.210 13.17 5.997	
200.000	2.301			30 22 73.333 73 5.61 5.636				5.610 16.74 5.589	
100.000	2.000			30 14 46.667 47 4.92 5.213				4.942 18.81	5.180
50.000	1.699			30 11 36.667 37 4.67 4.790				4.662 18.48 4.771	
25.000	1.398	30	9	30.000		30 4.48 4.368		4.490 15.96 4.363	
12.500	1.097	30		5 16.667		17 4.05 3.945		4.062 12.15 3.954	

Regression equation: $Y = 2.465 + 1.357X$ Chi-squared is 2.291 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 1.867 ppm LC_{50} is 73.680ppm 95% confidence limits are 52.412 to 103.580ppm

Regression equation: $Y = -1.608 + 2.662X$ Chi-squared is 0.529 with 1 degrees of freedom

No significant heterogeneity Log LC_{50} is 2.482 ppm LC_{50} is 303.296ppm

95% confidence limits are 166.973 to 550.917ppm

Appendix Table DXXXII: Larvicidal effect of *S. nodiflora* (wp/PetE) extracts against *C. quinquefasciatus* after 12h of exposure

Regression equation: $Y = -1.908 + 3.139X$ Chi-squared is 0.302 with 1 degrees of freedom No significant heterogeneity

Log LC_{50} is 2.201ppm

LC⁵⁰ is 158.823ppm

95% confidence limits are 123.539 to 204.183ppm

Appendix Table DXXXIII: Larvicidal effect of *S. nodiflora* (wp/PetE) extracts against *C. quinquefasciatus* after 18h of exposure

Regression equation: $Y = 1.033 + 1.901X$ Chi-squared is 8.627 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.087 ppm LC_{50} is 122.058ppm 95% confidence limits are 86.875 to 171.491ppm

Appendix Table DXXXIV: Larvicidal effect of *S. nodiflora* (wp/PetE) extracts against *C. quinquefasciatus* after 24h of exposure

Regression equation: $Y = 2.639 + 1.141X$ Chi-squared is 1.738 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.069ppm LC₅₀ is 117.128ppm 95% confidence limits are 52.580 to 260.918ppm

Regression equation: $Y = 3.511 + 0.875X$ Chi-squared is 3.793 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.702ppm LC_{50} is 50.361ppm 95% confidence limits are 26.170 to 96.914ppm

Appendix Table DCXXXVII: Larvicidal effect of *S. nodiflora*(wp/CHCl₃) extracts against *C. quinquefasciatus* after 12h of exposure

Regression equation: $Y = 0.439 + 1.721X$ Chi-squared is 0.655 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.650ppm LC_{50} is 446.388ppm 95% confidence limits are 258.934 to 769.548ppm

Appendix Table DLXXXVI: Larvicidal effect of *S. nodiflora* (wp/CHCl3) extracts against *C. quinquefasciatus* after 6h of exposure

Regression equation: $Y = 1.236 + 1.092X$ Chi-squared is 0.610 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 3.446ppm LC_{50} is 2793.540ppm 95% confidence limits are 190.263 to 41016.220ppm **Appendix Table DLXXXVIII:** Larvicidal effect of *S. nodiflora* (wp/CHCl3) extracts against *C. quinquefasciatus* after 18h of exposure

Regression equation: $Y = 0.938 + 1.776X$ Chi-squared is 0.835 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.286ppm LC_{50} is 193.416ppm 95% confidence limits are 139.520 to 268.130ppm

Regression equation: $Y = 1.362 + 1.753X$ Chi-squared is 2.254 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.076ppm LC_{50} is 119.114ppm 95% confidence limits are 88.830 to 159.723ppm

Appendix Table DXCI: Larvicidal effect of *S. nodiflora* (wp/CH₃OH) extracts against *C. quinquefasciatus* after 6h of exposure

Regression equation: $Y = 2.028 + 0.921X$ Chi-squared is 0.431 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 3.227ppm LC_{50} is 1686.443ppm 95% confidence limits are 252.683 to 11255.57ppm

Appendix Table DXC: Larvicidal effect of *S. nodiflora* (wp/CHCl₃) extracts against *C. quinquefasciatus* after 30h of exposure

Regression equation: $Y = 1.506 + 1.750X$ Chi-squared is 2.061 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.996ppm LC_{50} is 99.141ppm 95% confidence limits are 73.979 to 132.860ppm

Appendix Table DXCII: Larvicidal effect of *S. nodiflora* (wp/CH₃OH) extracts against *C. quinquefasciatus* after 12h of exposure

Regression equation: $Y = 1.905 + 1.170X$ Chi-squared is 0.611 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.645ppm LC_{50} is 441.278 ppm 95% confidence limits are 215.263 to 904.595ppm

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Regression equation: $Y = 1.721 + 1.430X$ Chi-squared is 0.436 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.293 ppm LC_{50} is 196.477ppm 95% confidence limits are 131.411 to 293.762ppm

Appendix Table DXCV: Larvicidal effect of *S. nodiflora* (wp/CH₃OH) extracts against *C. quinquefasciatus* after 30h of exposure

Regression equation: $Y = 1.541 + 1.679X$ Chi-squared is 3.335 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.060 ppm LC_{50} is 114.765ppm 95% confidence limits are 85.286 to 154.434ppm

Appendix Table DXCIV: Larvicidal effect of *S. nodiflora* (wp/CH3OH) extracts against *C. quinquefasciatus* after 24h of exposure

Regression equation: $Y = 1.734 + 1.492X$ Chi-squared is 0.479 with 4 degrees of freedom No significant heterogeneity Log LC_{50} is 2.188 ppm LC_{50} is 154.247 ppm 95% confidence limits are 108.216 to 219.856ppm

Appendices CXLIX IES, RU

Appendix Table DXCVI: Larvicidal effect of *Z. zerumbet* (ap/PetE) extracts against *C. quinquefasciatus* after 6h of exposure

Dose					Ldos #U Kl %Kill Cr% E Pr Ex Pr Wk Pro Weght F Pro		
400.000					2.602 30 16 53.333 53 5.08 4.990 5.065 19.02 4.988		
200.000			2.301 30 8 26.667 27 4.39 4.511			4.376 17.43 4.500	
100.000	2.000		30 5 16.667 17 4.05 4.032			4.037 13.17 4.011	
50.000	1.699	$\overline{30}$	2 6.667 7 3.52 3.553			3.519 8.07 3.523	
25.000	1.398	30	1 3.333 3 3.12 3.074			3.135 3.93 3.034	

Regression equation: $Y = 0.766 + 1.623X$ Chi-squared is 0.428 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.609ppm LC_{50} is 406.664ppm 95% confidence limits are 238.150 to 694.419ppm

Regression equation: $Y = 1.610 + 1.516X$ Chi-squared is 1.863 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.237ppm LC_{50} is 172.488ppm 95% confidence limits are 120.160 to 247.606ppm

Appendix Table DXCIX: Larvicidal effect of *Z. zerumbet* (ap/PetE) extracts against *C. quinquefasciatus* after 24h of exposure

Regression equation: $Y = 2.073 + 1.583X$ Chi-squared is 1.701 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.849ppm LC_{50} is 70.661 ppm 95% confidence limits are 50.046 to 99.768ppm

Appendix Table DXCVIII: Larvicidal effect of *Z. zerumbet* (ap/PetE) extracts against *C. quinquefasciatus* after 18h of exposure

Regression equation: $Y = 1.435 + 1.837X$ Chi-squared is 4.016 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.941ppm LC_{50} is 87.267ppm 95% confidence limits are 65.915 to 115.536ppm

Appendix Table DC: Larvicidal effect of *Z. zerumbet* (ap/PetE) extracts against *C. quinquefasciatus* after 30h of exposure

Regression equation: $Y = 1.125 + 2.220X$ Chi-squared is 0.453 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 1.746ppm LC_{50} is 55.681ppm 95% confidence limits are 42.761 to 72.504ppm

Appendix Table DCI: Larvicidal effect of *Z. zerumbet* (ap/CHCl3) extracts against

Regression equation: $Y = -0.168 + 1.619X$ Chi-squared is 0.032 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 3.191 ppm LC_{50} is 1553.064ppm 95% confidence limits are 608.079 to 3966.607ppm

Appendix Table DCII: Larvicidal effect of *Z. zerumbet* (ap/CHCl₃) extracts against *C. quinquefasciatus* after 12h of exposure

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Regression equation: $Y = 1.499 + 1.284X$ Chi-squared is 0.220 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.727 ppm LC_{50} is 532.872ppm 95% confidence limits are 317.245 to 895.057ppm **Appendix Table DCIII:** Larvicidal effect of *Z. zerumbet* (ap/CHCl₃) extracts against *C. quinquefasciatus* after 18h of exposure

Regression equation: $Y = 1.976 + 1.151X$ Chi-squared is 0.321 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.627ppm LC_{50} is 423.255ppm 95% confidence limits are 253.320 to 707.189ppm

Appendix Table DCIV: Larvicidal effect of *Z. zerumbet* (ap/CHCl3) extracts against *C. quinquefasciatus* after 24h of exposure

Regression equation: $Y = 1.712 + 1.348X$ Chi-squared is 0.149 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.439 ppm LC_{50} is 274.740ppm 95% confidence limits are 188.522 to 400.389ppm

Regression equation: $Y = 0.915 + 1.848X$ Chi-squared is 2.616 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.210ppm LC_{50} is 162.143ppm 95% confidence limits are 122.329 to 214.917ppm

Appendix Table DCVI: Larvicidal effect of *Z. zerumbet* (rh/PetE) extracts against *C. quinquefasciatus* after 6h of exposure

Regression equation: $Y = 2.712 + 1.371X$ Chi-squared is 1.802 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.668ppm LC_{50} is 46.592ppm 95% confidence limits are 25.893 to 83.837ppm

Appendix Table DCVII: Larvicidal effect of *Z. zerumbet* (rh/PetE) extracts against *C. quinquefasciatus* after 12h of exposure

Regression equation: $Y = 3.173 + 1.510X$ Chi-squared is 1.495 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.210ppm LC_{50} is 16.216ppm 95% confidence limits are 11.574 to 22.720ppm

Appendix Table DCVIII: Larvicidal effect of *Z. zerumbet* (rh/PetE) extracts against *C. quinquefasciatus* after 18h of exposure

Regression equation: $Y = 3.262 + 1.704X$ Chi-squared is 2.238 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 1.020ppm LC_{50} is 10.475ppm 95% confidence limits are 7.767 to 14.126ppm

Regression equation: $Y = 3.623 + 1.552X$ Chi-squared is 0.798 with 2 degrees of freedom No significant heterogeneity Log LC_{50} IS 0.887ppm LC_{50} is 7.717ppm 95% confidence limits are 5.411 to 11.007ppm

Appendix Table DCX: Larvicidal effect of *Z. zerumbet* (rh/PetE) extracts against *C. quinquefasciatus* after 30h of exposure

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Regression equation: $Y = 3.892 + 1.514X$ Chi-squared is 0.651 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 0.732ppm LC_{50} is 5.389ppm 95% confidence limits are 3.564 to 8.150ppm

Appendix Table DCXI: Larvicidal effect of *Z. zerumbet* (rh/CHCl₃) extracts against *C. quinquefasciatus* after 6h of exposure

Regression equation: $Y = -0.454 + 1.868X$ Chi-squared is 0.318 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 2.919ppm LC_{50} is 830.202 ppm 95% confidence limits are 500.995 to 1375.732ppm

Appendix Table DCXII: Larvicidal effect of *Z. zerumbet* (rh/CHCl₃) extracts against *C. quinquefasciatus* after 12h of exposure

Regression equation: $Y = 1.195 + 1.381X$ Chi-squared is 0.339 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 2.755ppm LC_{50} is 569.500ppm 95% confidence limits are 341.515 to 949.682ppm

Regression equation: $Y = 1.829 + 1.204X$ Chi-squared is 0.997 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.634ppm LC_{50} is 430.237ppm 95% confidence limits are 262.051 to 706.368ppm

Appendix Table DCXIV: Larvicidal effect of *Z. zerumbet* (rh/CHCl3) extracts against *C. quinquefasciatus* after 24h of exposure

Regression equation: $Y = 1.497 + 1.497X$ Chi-squared is 0.632 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.340 ppm LC_{50} is 218.993ppm 95% confidence limits are 157.124 to 305.225ppm

Appendix Table DCXV: Larvicidal effect of *Z. zerumbet* (rh/CHCl₃) extracts against *C. quinquefasciatus* after 30h of exposure

No significant heterogeneity

Log LC_{50} is 2.238 ppm

 LC_{50} is 172.985ppm

95% confidence limits are 124.531 to 240.292ppm

Appendix Table DCXVI: Larvicidal effect of *Z. zerumbet* (rh/CH₃OH) extracts against *C. quinquefasciatus* after 6h of exposure

Regression equation: $Y = 1.202 + 1.275X$ Chi-squared is 1.334 with 3 degrees of freedom No significant heterogeneity Log LC_{50} IS 2.978ppm LC_{50} is 949.989ppm 95% confidence limits are 457.238 to 1973.761ppm

Regression equation: $Y = 0.895 + 1.549X$ Chi-squared is 1.480 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.651ppm LC_{50} is 447.359ppm 95% confidence limits are 301.568 to 663.632ppm

Appendix Table DCXVIII: Larvicidal effect of *Z. zerumbet* (rh/CH₃OH) extracts against *C. quinquefasciatus* after 18h of exposure

────────────────────────────────────────────────────────────────────── Regression equation: $Y = 0.859 + 1.695X$ Chi-squared is 0.909 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.443 ppm LC_{50} is 277.516ppm 95% confidence limits are 203.366 to 378.701ppm

Appendix Table DCXIX: Larvicidal effect of *Z. zerumbet* (rh/CH₃OH) extracts against *C. quinquefasciatus* after 24h of exposure

Regression equation: $Y = -0.042 + 2.206X$ Chi-squared is 2.303 with 3 degrees of freedom No significant heterogeneity Log LC_{50} is 2.286ppm LC_{50} is 193.248ppm 95% confidence limits are 151.477 to 246.538ppm

Appendix Table DCXX: Larvicidal effect of *Z. zerumbet* (rh/CH₃OH) extracts against *C. quinquefasciatus* after 30h of exposure

Regression equation: $Y = 0.244 + 2.118X$ Chi-squared is 0.093 with 2 degrees of freedom No significant heterogeneity Log LC_{50} is 2.245 ppm LC_{50} is 175.940 ppm 95% confidence limits are 133.551 to 231.784ppm

Regression equation: $Y = 1.258 + 2.332X$ Chi-squared is 0.505 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.604mg/cm² LD_{50} is 4.022 mg/cm² 95% confidence limits are 1.174 to 13.784 mg/cm²

Appendix Table DCXXII: Dose mortality effect of *E. nummularius* (wp/PetE) extracts against *T. castaneum* after12h of exposure

Regression equation: $Y = -0.216 + 4.711X$ Chi-squared is 58.525 with 4 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.107 mg/cm² LD_{50} is 1.280mg/cm² 95% confidence limits are 0.789 to 2.078 mq/cm²

α **Appendices** \Box **IES, RU**

Regression equation: $Y = -0.193 + 4.781X$ Chi-squared is 92.388 with 4 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.086 mg/cm² LD_{50} is 1.220 mg/cm² 95% confidence limits are 0.691 to $2.153mg/cm²$

Regression equation: $Y = -0.204 + 5.005X$ Chi-squared is 35.827 with 4 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.040 mg/cm² LD_{50} is 1.096 mg/cm² 95% confidence limits are 0.799 to $1.502mg/cm²$

Regression equation: $Y = -0.007 + 4.942X$ Chi-squared is 28.798 with 4 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.013mg/cm² LD_{50} is 1.031mg/cm² 95% confidence limits are 0.779 to $1.364mg/cm²$

Appendix Table DCXXVII: Dose mortality effect of *E. nummularius* (wp/CHCl3) extracts against *T. castaneum* after 12h of exposure

Regression equation: $Y = 3.105 + 3.704X$ Chi-squared is 1.408 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is $0.512mg/cm^{2}$ LD_{50} is 3.248mg/cm² 95% confidence limits are 20.839 to 3.717mg/cm²

Appendix Table DCXXVI: Dose mortality effect of *E. nummularius* (wp/CHCl3) extracts against *T. castaneum* after 30min of exposure

Regression equation: $Y = 7.361 + -6.957X$ Chi-squared is -000008 with 0 degrees of freedom No significant heterogeneity

- Log LD_{50} is 0.339 mg/cm²
- LD_{50} is 2.185mg/cm²
- 95% confidence limits are 0.469 to 10.186 mg/cm²

Appendix Table DCXXVIII: Dose mortality effect of *E. nummularius* (wp/CHCl3) extracts against *T. castaneum* after 24h of exposur**e**

Regression equation: $Y = 3.042 + 4.116X$ Chi-squared is 0.749 with 3 degrees of freedom No significant heterogeneity Log LD_{50} izs 0.476 mg/cm² LD_{50} is 2.990mg/cm² 95% confidence limits are 2.663 to 3.358mg/cm²

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Regression equation: $Y = 2.793 + 5.483X$ Chi-squared is 3.278 with 4 degrees of freedom No significant heterogeneity Log LD_{50} is 0.402 mg/cm² LD_{50} is 2.526mq/cm² 95% confidence limits are 2.313 to 2.759mg/cm²

Appendix Table DCXXXI: Dose mortality effect of *E. nummularius* (wp/CH₃OH) extracts against *T. castaneum* after 30min of exposure

Regression equation: $Y = 2.717 + 2.203X$ Chi-squared is 1.345 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.036 mg/cm² LD_{50} is 10.876mg/cm² 95% confidence limits are 1.195 to 98.945mg/cm²

Regression equation: $Y = 3.131 + 5.446X$ Chi-squared is 5.283 with 4 degrees of freedom No significant heterogeneity Log LD_{50} is 0.343 mg/cm² LD_{50} is 2.204mg/cm² 95% confidence limits are 2.000 to 2.429 mg/cm²

α **Appendices** \Box **IES, RU**

Appendix Table DCXXXII: Dose mortality effect of *E. nummularius* (wp/CH₃OH) extracts against *T. castaneum* after 12h of exposure

Dose				Ldos #U Kl %Kill Cr% EPr ExPr WkPro Weght FPro	
				3.567 0.552 30 19 63.333 63 5.33 5.399 5.318 18.48 5.389 3.057 0.485 30 17 56.667 57 5.18 5.094 5.175 19.11 5.086 2.548 0.406 30 12 40.000 40 4.75 4.734 4.740 18.48 4.729	
				2.038 0.309 30 7 23.333 23 4.26 4.293 4.252 15.09 4.291	

Regression equation: $Y = 2.895 + 4.516X$ Chi-squared is 0.268 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.466mg/cm² LD_{50} is 2.925mg/cm² 95% confidence limits are 2.590 to 3.305mg/cm²

Regression equation: $Y = 2.274 + 9.789X$ Chi-squared is 12.185 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 0.278 mg/cm² LD_{50} is 1.899mg/cm² 95% confidence limits are 1.645 to 2.191mg/cm²

Appendix TableDCXXXV: Dose mortality effect of *E. nummularius* (wp/CH₃OH) extracts against *T. castaneum* after 48h of exposure

Dose				Ldos #U Kl %Kill Cr% E Pr Ex Pr Wk Pro Weght F Pro	
				2.038 0.309 30 27 90.000 90 6.28 6.280 6.230 11.10 6.230 1,529 0,184 30 2 6,667 7 3,52 3,520 3,519 8,07 3,519	

Regression equation: $Y = -0.487 + 21.724X$ Chi-squared is -0.00003 with 0 degrees of freedom No significant heterogeneity Log LD_{50} is 0.253 mg/cm² LD_{50} is 1.789mg/cm² 95% confidence limits are 1.706 to 1.876 mg/cm²

Regression equation: $Y = 1.139 + 14.147X$ Chi-squared is 2.918 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.273 mg/cm² LD_{50} is 1.875mg/cm² 95% confidence limits are 1.772 to 1.984mq/cm²

Appendix Table DCXXXVI: Dose mortality effect of *L. camara* (ap/PetE) extracts against *T. castaneum* after 12h of exposure

Dose				Ldos #U Kl %Kill Cr% E Pr Ex Pr Wk Pro Weght F Pro	
				4.076 0.610 30 9 30.000 30 4.48 4.576 4.460 17.43 4.565 3.567 0.552 30 7 23.333 23 4.26 4.044 4.283 13.17 4.025	
				3.057 0.485 30 1 3.333 3 3.12 3.428 3.180 7.14 3.401	

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Regression equation: $Y = -1.120 + 9.316X$ Chi-squared is 1.416 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.657 mg/cm² LD_{50} is 4.539 mg/cm² 95% confidence limits are 3.824 to 5.388 mg/cm²

Regression equation: $Y = 1.431 + 5.799X$ Chi-squared is 0.567 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.615 mg/cm² LD_{50} is 4.125mg/cm² 95% confidence limits are 3.572 to 4.763mq/cm²

Appendix Table DCXXXIX: Dose mortality effect of *L. camara* (ap/PetE) extracts against *T. castaneum* after 48h of exposure

Regression equation: $Y = 1.161 + 7.922X$ Chi-squared is 0.628 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.485mg/cm² LD_{50} is 3.052mg/cm² 95% confidence limits are 2.856 to 3.261mg/cm²

Regression equation: $Y = 1.247 + 6.772X$ Chi-squared is 0.858 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.554 mg/cm² LD_{50} is 3.583mg/cm² 95% confidence limits are 3.281 to 3.912mg/ $cm²$

Appendix Table DCXL: Dose mortality effect of *L. camara* (ap/CHCl₃) extracts against *T. castaneum* after 12h of exposure

Dose				Ldos #U Kl %Kill Cr% E Pr Ex Pr Wk Pro Weght F Pro	
				2.548 0.406 30 15 50.000 50 5.00 4.817 5.020 18.81 4.838	
				2.038 0.309 30 5 16.667 17 4.05 4.305 4.074 15.96 4.318	
				1.529 0.184 30 2 6.667 7 3.52 3.646 3.529 9.06 3.647	
1.019				0.008 30 1 3.333 3 3.12 2.716 3.379 2.28 2.701	

Regression equation: $Y = 2.657 + 5.370X$ Chi-squared is 2.742 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.436 mg/cm² LD_{50} is 2.731 mg/cm² 95% confidence limits are 2.253 to 3.310 mg/cm²

Appendices CLX IES, RU

Regression equation: $Y = 3.249 + 4.674X$ Chi-squared is 1.296 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.375 mg/cm² LD_{50} is 2.370 mg/cm² 95% confidence limits are 2.018 to 2.783 mq/cm²

Appendix Table DCXLII: Dose mortality effect of *L. camara* (ap/CHCl₃) extracts against *T. castaneum* after 36h of exposure

Regression equation: $Y = -0.520 + 4.261X$ Chi-squared is 7.061 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.296 mg/cm² LD_{50} is 1.975mg/cm² 95% confidence limits are 1.721 to 2.266mg/cm²

Appendix Table DCXLIII: Dose mortality effect of *L. camara* (ap/CHCl₃) extracts against *T. castaneum* after 48h of exposure

Regression equation: $Y = -1.079 + 5.128X$ Chi-squared is 6.500 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.185mg/cm² LD_{50} is 1.533mg/cm² 95% confidence limits are 1.369 to 1.716mg/cm²

Appendix Table DCXLIV: Dose mortality effect of *L. camara* (ap/CH₃OH) extracts against *T. castaneum* after 12h of exposure

Regression equation: $Y = 1.709 + 2.307X$ Chi-squared is 1.679 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.426mg/cm² LD_{50} is 2.668mg/cm²

95% confidence limits are 1.625 to 4.381 mq/cm²

Regression equation: $Y = 3.340 + 1.151X$ Chi-squared is 0.059 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.442 mg/cm² LD_{50} is 2.765mg/cm² 95% confidence limits are 1.095 to 6.982 mq/cm²

Appendix Table DCXLVII: Dose mortality effect of *L. camara* (ap/CH₃OH) extracts against *T. castaneum* after 48h of exposure

Regression equation: $Y = 2.205 + 3.101X$ Chi-squared is 4.810 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.902 mg/cm² LD_{50} is 0.797mq/cm² 95% confidence limits are 0.666 to 0.954mg/cm²

Appendix Table DCXLVI: Dose mortality effect of *L. camara* (ap/CH₃OH) extracts against *T. castaneum* after 36h of exposure

Regression equation: $Y = 3.070 + 1.850X$ Chi-squared is 1.697 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.043 mg/cm² LD_{50} is 1.105 mg/cm² 95% confidence limits are 0.827 to 1.477mg/cm²

Appendix Table DCXLVIII: Dose mortality effect of *L. camara* (r/PetE) extracts against *T. castaneum* after 24h of exposure

Dose				Ldos #U Kl %Kill Cr% E Pr Ex Pr Wk Pro Weght F Pro	
				3.567 0.552 30 15 50.000 50 5.00 5.126 4.990 19.02 5.097 3.057 0.485 30 11 36.667 37 4.67 4.438 4.690 16.74 4.465 2.548 0.406 30 2 6.667 7 3.52 3.626 3.529 9.06 3.719	

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Regression equation: $Y = -0.111 + 9.429X$ Chi-squared is 1.392 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.542 mg/cm² LD_{50} is 3.483mg/cm² 95% confidence limits are 3.178 to 3.818 mg/cm²

α **Appendices** \Box **IES, RU**

Regression equation: $Y = 0.685 + 8.666X$ Chi-squared is 1.312 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.498mg/cm² LD_{50} is 3.148 mg/cm² 95% confidence limits are 2.925 to 3.388 mg/cm²

Appendix Table DCL: Dose mortality effect of *L. camara* (r/PetE) extracts against *T. castaneum* after 48h of exposure

Regression equation: $Y = 2.064 + 6.880X$ Chi-squared is 0.710 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is $0.427mg/cm^2$ LD_{50} is 2.672mg/cm² 95% confidence limits are 2.474 to 2.885 mq/cm²

Appendix Table DCLI: Dose mortality effect of *M. piperita* (wp/PetE) extracts against *T. castaneum* after 30min of exposure

Regression equation: $Y = 1.427 + 2.783X$ Chi-squared is 0.829 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 1.284 mg/cm² LD_{50} is 1.923 mg/cm² 95% confidence limits are 1.228 to 3.011mg/cm²

Appendix Table DCLII: Dose mortality effect of *M. piperita* (wp/PetE) extracts against *T. castaneum* after 12h of exposure

Regression equation: $Y = 2.974 + 2.137X$ Chi-squared is 4.595 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.948 mg/cm² LD_{50} is 0.888 mg/cm² 95% confidence limits are 0.677 to 1.163mg/cm²

Regression equation: $Y = 3.276 + 2.287X$ Chi-squared is 8.995 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 0.754 mg/cm² LD_{50} is 0.567mg/cm² 95% confidence limits are 0.380 to 0.848mg/cm²

Appendix Table DCLIV: Dose mortality effect of *M. piperita* (wp/PetE) extracts against *T. castaneum* after 36h of exposure

Regression equation: $Y = 3.893 + 1.619X$ Chi-squared is 0.363 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.684 mg/cm² LD_{50} is 0.483mg/cm² 95% confidence limits are 0.339 to 0.688 mq/cm²

Appendix Table DCLV: Dose mortality effect of *M. piperita* (wp/PetE) extracts against *T. castaneum* after 48h of exposure

Regression equation: $Y = 4.288 + 1.472X$ Chi-squared is 0.084 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.484 mg/cm² LD_{50} is 0.305mg/cm² 95% confidence limits are 0.209 to 0.444 mg/cm²

Appendix Table DCLVI: Dose mortality effect of *M. piperita* (wp/CHCl₃) extracts against *T. castaneum* after 12h of exposure

Regression equation: $Y = 2.844 + 5.099X$ Chi-squared is 0.607 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.423 mg/cm² LD_{50} is 2.647 mg/cm² 95% confidence limits are 2.160 to 3.244mg/cm²

Regression equation: $Y = 0.152 + 3.647X$ Chi-squared is 5.101 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.329 mg/cm² LD_{50} is 2.135 mg/cm² 95% confidence limits are 1.798 to 2.536mg/cm²

Appendix Table DCLIX: Dose mortality effect of *M. piperita* (wp/CHCl₃) extracts against *T. castaneum* after 48h of exposure

Regression equation: $Y = 1.338 + 3.330X$ Chi-squared is 1.509 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.099mg/cm² LD_{50} is 1.257mg/cm² 95% confidence limits are 1.061 to 1.490 mg/cm²

Appendix Table DCLVIII: Dose mortality effect of *M. piperita* (wp/CHCl3) extracts against *T. castaneum* after 36h of exposure

Regression equation: $Y = 0.932 + 3.405X$ Chi-squared is 5.736 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.195 mg/cm² LD_{50} is 1.567mg/cm² 95% confidence limits are 1.345 to 1.824 mg/cm²

Appendix Table DCLX: Dose mortality effect of *M. piperita* (wp/CH₃OH) extracts against *T. castaneum* after 30min of exposure

Regression equation: $Y = 3.074 + 1.930X$ Chi-squared is 0.270 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.998mg/cm² LD_{50} is 0.995mg/cm² 95% confidence limits are 0.723 to 1.369mg/cm²

Appendices CLXV IES, RU

Regression equation: $Y = 3.434 + 1.957X$ Chi-squared is 0.780 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.800 mg/cm² LD_{50} is 0.631mq/cm² 95% confidence limits are 0.484 to 0.823 mq/cm²

Appendix Table DCLXIII: Dose mortality effect of *M. piperita* (wp/CH₃OH) extracts against *T. castaneum* after 36h of exposure

Regression equation: $Y = 4.231 + 1.328X$ Chi-squared is 0.032 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.580 mg/cm² LD_{50} is 0.380 mg/cm² 95% confidence limits are 0.253 to 0.569 mg/cm²

Appendix Table DCLXII: Dose mortality effect of *M. piperita* (wp/CH₃OH) extracts against *T. castaneum* after 24h of exposure

Regression equation: $Y = 3.823 + 1.867X$ Chi-squared is 5.784 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.630 mg/cm² LD_{50} is 0.427mg/cm² 95% confidence limits are 0.325 to 0.561 mg/cm²

Appendix Table DCLXIV: Dose mortality effect of *M. piperita* (wp/CH₃OH) extracts against *T. castaneum* after 48h of exposure

Regression equation: $Y = 4.372 + 1.668X$ Chi-squared is 0.074 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.377 mg/cm² LD_{50} is 0.238 mg/cm² 95% confidence limits are 0.166 to 0.342 mg/cm²

α **Appendices** \Box **IES, RU**

Regression equation: $Y = -1.788 + 12.349$ X Chi-squared is 0.167 with 1 degrees of freedom No significant heterogeneity

Log LD_{50} is 0.550 mg/cm²

 LD_{50} is 3.546mg/cm²

95% confidence limits are 3.284 to 3.828mg/cm²

Appendix Table DCLXVI: Dose mortality effect of *P. hysterophorus* (wp/PetE) extracts against *T. castaneum* after 24h of exposure

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Regression equation: $Y = 1.130 + 7.648X$ Chi-squared is 2.046 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.506mg/cm² LD_{50} is 3.206mg/cm² 95% confidence limits are 2.941 to 3.494mg/cm²

Appendix Table DCLXVII: Dose mortality effect of *P. hysterophorus* (wp/PetE) extracts against *T. castaneum* after 36h of exposure

Regression equation: $Y = 1.135 + 8.192X$ Chi-squared is 1.851 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is $0.472mg/cm^2$ LD_{50} is 2.963mg/cm² 95% confidence limits are 2.759 to 3.183 mg/cm²

Appendix Table DCLXVIII: Dose mortality effect of *P. hysterophorus* (wp/PetE) extracts against *T. castaneum* after 48h of exposure

Regression equation: $Y = 1.850 + 7.468X$ Chi-squared is 2.671 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is $0.422mg/cm^{2}$ LD_{50} is 2.642mg/cm² 95% confidence limits are 2.460 to $2.836mg/cm²$

Regression equation: $Y = 1.413 + 5.092X$ Chi-squared is 0.786 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.704 mg/cm² LD_{50} is 5.063 mg/cm² 95% confidence limits are 3.668 to 6.990 mq/cm²

Appendix Table DCLXX: Dose mortality effect of *P. hysterophorus* (wp/CHCl₃) extracts against *T. castaneum* after 24h of exposure

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Regression equation: $Y = 2.157 + 4.730X$ Chi-squared is 0.115 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.601 mg/cm² LD_{50} is 3.991mq/cm² 95% confidence limits are 3.371 to 4.725 mq/cm²

Appendix Table DCLXXI: Dose mortality effect of *P. hysterophorus* (wp/CHCl3) extracts against *T. castaneum* after 36h of exposure

Regression equation: $Y = 0.801 + 8.164X$ Chi-squared is 1.116 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.514 mg/cm² LD_{50} is 3.268mg/cm² 95% confidence limits are 3.058 to 3.493mg/cm²

Appendix Table DCLXXII: Dose mortality effect of *P. hysterophorus* (wp/CHCl₃) extracts against *T. castaneum* after 48h of exposure

Regression equation: $Y = 0.997 + 8.564X$ Chi-squared is 1.700 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is $0.467mg/cm^2$ LD_{50} is 2.934mq/cm² 95% confidence limits are 2.755 to 3.125mg/cm²

Regression equation: $Y = 2.297 + 5.796X$

Chi-squared is 0.974 with 1 degrees of freedom No significant heterogeneity

Log LD_{50} is 0.466 mg/cm²

 LD_{50} is 2.927mg/cm²

95% confidence limits are 2.301 to 3.723mg/cm²

Appendix Table DCLXXIV: Dose mortality effect of *P. hysterophorus* (wp/CH₃OH) extracts against *T. castaneum* after 12h of exposure

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Regression equation: $Y = 3.519 + 3.734X$ Chi-squared is 0.124 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.397mg/cm² LD_{50} is 2.493 mg/cm² 95% confidence limits are 1.970 to 3.154mg/cm²

Appendix Table DCLXXV: Dose mortality effect of *P. hysterophorus* (wp/CH₃OH) extracts against *T. castaneum* after 24h of exposure

Regression equation: $Y = 3.583 + 4.202X$ Chi-squared is 0.374 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is $0.337mg/cm²$ LD_{50} is 2.174 mg/cm² 95% confidence limits are 1.861 to 2.539 mg/cm²

Appendix Table DCLXXVI: Dose mortality effect of *P. hysterophorus* (wp/CH₃OH) extracts against *T. castaneum* after 36h of exposure

Regression equation: $Y = 3.723 + 4.938X$ Chi-squared is 3.862 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.259 mg/cm² LD_{50} is 1.814 mg/cm² 95% confidence limits are 1.614 to 2.038mg/cm²

Regression equation: $Y = 0.389 + 3.802X$ Chi-squared is 0.311 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.213mg/cm² LD_{50} is 1.631mg/cm² 95% confidence limits are 1.376 to 1.935mg/cm²

Regression equation: $Y = 1.074 + 2.614X$ Chi-squared is 0.517 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.502 mg/cm² LD_{50} is 3.174 mg/cm² 95% confidence limits are 1.778 to 5.665mg/cm²

Appendix Table DCLXXIX: Dose mortality effect of *Ph. niruri* (wp/PetE) extracts against *T. castaneum* after 12h of exposure

Regression equation: $Y = 1.668 + 2.771X$ Chi-squared is 3.743 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.203 mg/cm² LD_{50} is 1.595mg/cm² 95% confidence limits are 1.272 to 1.999mg/cm²

Appendix Table DCLXXX: Dose mortality effect of *Ph. niruri* (wp/PetE) extracts against *T. castaneum* after 24h of exposure

Regression equation: $Y = 2.150 + 2.613X$ Chi-squared is 7.160 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.091 mg/cm² LD_{50} is 1.233mg/cm² 95% confidence limits are 1.001 to $1.519mg/cm²$

Regression equation: $Y = 2.125 + 2.917X$ Chi-squared is 14.537 with 3 degrees of freedom

Variance has been adjusted for heterogeneity Log LD_{50} is 0.986mg/cm² LD_{50} is 0.967mq/cm² 95% confidence limits are 0.644 to 1.454 mg/cm²

Appendix Table DCLXXXIII: Dose mortality effect of *Ph. niruri* (wp/CHCl3) extracts against *T. castaneum* after 30min of exposure

Regression equation: $Y = 2.040 + 3.963X$ Chi-squared is 0.382 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.747 mg/cm² LD_{50} is 5.585 mg/cm² 95% confidence limits are 2.999 to 10.398 mg/cm²

Appendix Table DCLXXXII: Dose mortality effect of *Ph. niruri* (wp/PetE) extracts against *T. castaneum* after 48h of exposure

Regression equation: $Y = 3.655 + 1.905X$ Chi-squared is 0.310 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.706mg/cm² LD_{50} is 0.508mg/cm² 95% confidence limits are 0.374 to 0.691 mg/cm²

Appendix Table DCLXXXIV: Dose mortality effect of *Ph. niruri* (wp/CHCl3) extracts against *T. castaneum* after 12h of exposure

Regression equation: $Y = 2.756 + 3.317X$ Chi-squared is 2.316 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.676mg/cm² LD_{50} is 4.747 mg/cm² 95% confidence limits are 2.891 to 7.796mg/cm²

α **Appendices** \Box **IES, RU**

Regression equation: $Y = 1.913 + 5.585X$ Chi-squared is 3.489 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.553mg/cm² LD_{50} is 3.571mg/cm² 95% confidence limits are 3.085 to 4.133mg/cm²

Appendix Table DCLXXXVII: Dose mortality effect of *Ph. niruri* (wp/CHCl3) extracts against *T. castaneum* after 48h of exposure

Regression equation: $Y = 2.887 + 5.494X$ Chi-squared is 1.264 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.385mg/cm² LD_{50} is 2.425 mg/cm² 95% confidence limits are 2.212 to 2.659mg/cm²

Regression equation: $Y = 2.978 + 4.192X$ Chi-squared is 0.915 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.483 mg/cm² LD_{50} is 3.037mg/cm² 95% confidence limits are 2.645 to 3.488mg/ $cm²$

Appendix Table DCLXXXVIII: Dose mortality effect of *Ph. niruri* (wp/CH₃OH) extracts against *T. castaneum* after 30min of exposure

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Regression equation: $Y = 0.704 + 7.439X$ Chi-squared is 1.439 with 1 degree of freedom No significant heterogeneity Log LD_{50} is 0.578 mg/cm² LD_{50} is 3.781 mg/cm² 95% confidence limits are 3.218 to $4.442mg/cm²$

α **Appendices** \Box **IES, RU**

Regression equation: $Y = 2.403 + 4.731X$ Chi-squared is 3.356 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.549 mg/cm² LD_{50} is 3.541mg/cm² 95% confidence limits are 3.001 to 4.176mq/cm²

Appendix Table DCXC: Dose mortality effect of *Ph. niruri* (wp/CH₃OH) extracts against *T. castaneum* after 24h of exposure

Regression equation: $Y = 3.396 + 4.284X$ Chi-squared is 9.319 with 4 degrees of freedom No significant heterogeneity Log LD_{50} is 0.375mg/cm² LD_{50} is 2.369mg/cm² 95% confidence limits are 2.122 to 2.644 mg/cm²

Appendix Table DCXCI: Dose mortality effect of *Ph. niruri* (wp/CH₃OH) extracts against *T. castaneum* after 36h of exposure

Regression equation: $Y = 3.572 + 4.802X$ Chi-squared is 6.822 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.297mg/cm² LD_{50} is 1.983mg/cm² 95% confidence limits are 1.782 to 2.207mg/cm²

Appendix Table DCXCII: Dose mortality effect of *Ph. niruri* (wp/CH₃OH) extracts against *T. castaneum* after 48h of exposure

Regression equation: $Y = 3.744 + 5.001X$ Chi-squared is 4.097 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.251 mg/cm² LD_{50} is 1.783mg/cm² 95% confidence limits are 1.601 to 1.986mg/cm²

Chi-squared is 0.388 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.012 mg/cm² LD_{50} is 10.282mg/cm² 95% confidence limits are 0.090 to 1170.104mg/cm²

Appendix Table DCXCIV: Dose mortality effect of *Po. hydropiper* (wp/PetE) extracts against *T. castaneum* after 12h of exposure

Regression equation: $Y = 4.003 + 2.774X$ Chi-squared is 2.675 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.359mg/cm² LD_{50} is 2.288mg/cm² 95% confidence limits are 1.734 to 3.017mg/cm²

Appendix Table DCXCV: Dose mortality effect of *Po. hydropiper* (wp/PetE) extracts against *T. castaneum* after 24h of exposure

Regression equation: $Y = 3.603 + 5.746X$ Chi-squared is 1.309 with 4 degrees of freedom No significant heterogeneity Log LD_{50} is 0.243 mg/cm² LD_{50} is 1.750 mg/cm² 95% confidence limits are 1.613 to 1.899mg/cm²

Appendix Table DCXCVI: Dose mortality effect of *Po. hydropiper* (wp/PetE) extracts against *T. castaneum* after 36h of exposure

Regression equation: $Y = 3.667 + 5.883X$ Chi-squared is 1.255 with 4 degrees of freedom No significant heterogeneity Log LD_{50} is $0.227mg/cm^2$ LD_{50} is 1.685mg/cm² 95% confidence limits are 1.558 to $1.822mg/cm²$

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Regression equation: $Y = 3.674 + 6.737X$ Chi-squared is 4.203 with 4 degrees of freedom No significant heterogeneity Log LD_{50} is 0.197 mg/cm² LD_{50} is 1.573mg/cm² 95% confidence limits are 1.466 to 1.688 mq/cm²

Appendix Table DCXCIX: Dose mortality effect of *Po. hydropiper* (wp/CHCl3) extracts against *T. castaneum* after 24h of exposure

Regression equation: $Y = -4.148 + 14.052$ X Chi-squared is 3.121 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.651 mg/cm² LD_{50} is 4.478 mg/cm² 95% confidence limits are 4.222 to 4.749 mg/cm²

Appendix Table DCXCVIII: Dose mortality effect of *Po. hydropiper* (wp/CHCl3) extracts against *T. castaneum* after 12h of exposure

Regression equation: $Y = -6.749 + 17.880X$

Chi-squared is 7.43866e-04 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.657 mg/cm² LD_{50} is 4.540 mg/cm² 95% confidence limits are 4.313 to 4.780mg/cm²

Appendix Table DCC: Dose mortality effect of *Po. hydropiper* (wp/CHCl3) extracts against *T. castaneum* after 36h of exposure

────────────────────────────────────────────────────────────────────── Regression equation: $Y = -5.493 + 16.595X$ Chi-squared is 7.541 with 2 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is $0.632mg/cm^2$ LD_{50} is 4.288 mg/cm² 95% confidence limits are 3.948 to 4.658mg/cm²

Appendices CLXXV IES, RU

Regression equation: $Y = -2.647 + 12.658X$ Chi-squared is 11.101 with 3 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 0.604 mg/cm² LD_{50} is 4.019mg/cm² 95% confidence limits are 3.668 to 4.405 mq/cm²

Appendix Table DCCIII: Dose mortality effect of *Po. hydropiper* (wp/CH₃OH) extracts against *T. castaneum* after12h of exposure

Regression equation: $Y = 2.455 + 3.188X$ Chi-squared is 5.453 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.798mg/cm² LD_{50} is 0.629mg/cm² 95% confidence limits are 0.520 to 0.760 mg/cm²

Appendix Table DCCII: Dose mortality effect of *Po. hydropiper* (wp/CH₃OH) extracts against *T. castaneum* after 30min of exposure

Regression equation: $Y = 2.237 + 2.078X$ Chi-squared is 0.108 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.330mg/cm² LD_{50} is 2.137 mg/cm² 95% confidence limits are 1.215 to 3.759mg/cm²

Appendix Table DCCIV: Dose mortality effect of *Po. hydropiper* (wp/CH₃OH) extracts against *T. castaneum* after 24h of exposure

Regression equation: $Y = 2.313 + 3.880X$ Chi-squared is 2.791 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.693 mg/cm² LD_{50} is 0.493mg/cm² 95% confidence limits are 0.416 to 0.584 mg/cm²

Regression equation: $Y = 2.138 + 4.626X$

Chi-squared is 2.804 with 1 degrees of freedom No significant heterogeneity

Log LD_{50} is 0.619 mg/cm²

 LD_{50} is 0.416mg/cm²

95% confidence limits are 0.347 to 0.498 mg/cm²

Appendix Table DCCVI: Dose mortality effect of *Po. hydropiper* (wp/CH₃OH) extracts against *T. castaneum* after 48h of exposure

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Regression equation: $Y = 2.054 + 5.511X$ Chi-squared is 2.661 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.535 mg/cm² LD_{50} is 0.342 mg/cm² 95% confidence limits are 0.295 to 0.397mg/cm²

Appendix Table DCCVII: Dose mortality effect of *Pz. zeylanica* (wp/PetE) extracts against *T. castaneum* after 30min of exposure

Regression equation: $Y = 3.637 + 2.531X$ Chi-squared is 1.168 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.539mg/cm² LD_{50} is 3.457 mg/cm² 95% confidence limits are 2.413 to 4.951 mg/cm²

Appendix Table DCCVIII: Dose mortality effect of *Pz. zeylanica* (wp/PetE) extracts against *T. castaneum* after 12h of exposure

Regression equation: $Y = 4.165 + 3.066X$ Chi-squared is 3.204 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is $0.272mg/cm^{2}$ LD_{50} is 1.872 mg/cm² 95% confidence limits are 1.597 to 2.195mg/cm²

Regression equation: $Y = 4.473 + 3.610X$ Chi-squared is 4.544 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.150 mg/cm² LD_{50} is 1.400mg/cm² 95% confidence limits are 1.180 to 1.660mg/ $cm²$

Appendix Table DCCXI: Dose mortality effect of *Pz. zeylanica* (wp/PetE) extracts against *T. castaneum* after 48h of exposure

Regression equation: $Y = 5.272 + 2.784X$

Chi-squared is 0.815 with 1 degree of freedom

No significant heterogeneity

Log LD_{50} is -0.098 mg/cm²

- LD_{50} is 0.799mg/cm²
- 95% confidence limits are 0.471 to 1.353mg/cm²

Regression equation: $Y = 4.926 + 1.993X$ Chi-squared is 0.124 with 1 degree of freedom No significant heterogeneity Log LD_{50} is 0.037 mg/cm² LD_{50} is 1.090mg/cm² 95% confidence limits are 0.705 to 1.685mg/ $cm²$

Appendix Table DCCXII: Dose mortality effect of *Pz. zeylanica* (wp/CHCl₃) extracts against *T. castaneum* after 30min of exposure

Ldos					#U Kl %Kill Cr% E Pr Ex Pr Wk Pro Weght F Pro		
3.056					0.485 30 8 26.667 27 4.39 4.264 4.388 15.09 4.274		
2.547 2.037	0.406 30 0.309	$\overline{30}$	5 16.667 17 4.05 4.027		5 16.667 17 4.05 4.157 4.056 14.13 4.163	4.037 13.17 4.027	
1.528	0.184	30	3 10.000 10 3.72 3.859			3.720 11.10 3.852	
1.019	0.008	30	3 10.000 10 3.72 3.623			3.730 9.06 3.605	

Regression equation: $Y = 3.594 + 1.402X$ Chi-squared is 0.694 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.003 mg/cm² LD_{50} is 10.073 mg/cm² 95% confidence limits are 1.632 to 62.165mg/cm²

Regression equation: $Y = 4.068 + 1.902X$ Chi-squared is 0.393 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.490mg/cm² LD_{50} is 3.089mg/cm² 95% CONFIDENCE LIMITS ARE 2.075 to 4.601mq/cm²

Appendix Table DCCXV: Dose mortality effect of *Pz. zeylanica* (wp/CHCl₃) extracts against *T. castaneum* after 36h of exposure

Regression equation: $Y = 4.695 + 3.313X$ Chi-squared is 2.325 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.092 mg/cm² LD_{50} is 1.236mg/cm² 95% confidence limits are 0.998 to 1.532 mg/cm²

Appendix Table DCCXIV: Dose mortality effect of *Pz. zeylanica* (wp/CHCl3) extracts against *T. castaneum* after 24h of exposure

Regression equation: $Y = 4.412 + 2.673X$ Chi-squared is 1.698 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.220 mg/cm² LD_{50} is 1.659mg/cm² 95% confidence limits are 1.372 to 2.007mg/cm²

Appendix Table DCCXVI: Dose mortality effect of *Pz. zeylanica* (wp/CHCl₃) extracts against *T. castaneum* after 48h of exposure

Regression equation: $Y = 5.142 + 2.753X$ Chi-squared is 0.722 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is -0.0517 mg/cm² LD_{50} is 0.888 mg/cm² 95% confidence limits are 0.587 to 1.342mg/cm²

Regression equation: $Y = 2.963 + 10.877X$

Chi-squared is 0.072 with 1 degrees of freedom No significant heterogeneity

Log LD_{50} is 0.187 mg/cm²

 LD_{50} is 1.539mg/cm²

95% confidence limits are 1.404 to 1.687mg/cm²

Appendix Table DCCXVIII: Dose mortality effect of *S. nodiflora* (wp/PetE) extracts against *T. castaneum* after 24h of exposure

Regression equation: $Y = -4.595 + 8.327X$ Chi-squared is 5.552 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is $1.152mg/cm^2$ LD_{50} is 1.420mg/cm² 95% confidence limits are 1.298 to 1.553 mg/cm²

Appendix Table DCCXIX: Dose mortality effect of *S. nodiflora* (wp/PetE) extracts against *T. castaneum* after 36h of exposure

Regression equation: $Y = -3.906 + 7.926X$ Chi-squared is 3.716 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.124 mg/cm² LD_{50} is 1.329mg/cm² 95% confidence limits are 1.224 to 1.444 mg/cm²

Appendix Table DCCXX: Dose mortality effect of *S. nodiflora* (wp/PetE) extracts against *T. castaneum* after 48h of exposure

Regression equation: $Y = -4.246 + 8.435X$ Chi-squared is 8.047 with 2 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 1.096 mg/cm² LD_{50} is 1.248mg/cm²

95% confidence limits are 1.078 to 1.445mg/cm²

Regression equation: $Y = 0.741 + 7.047X$ Chi-squared is 2.661 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.604 mg/cm² LD_{50} is 4.021 mg/cm² 95% confidence limits are 3.333 to 4.852 mg/cm²

Appendix Table DCCXXII: Dose mortality effect of *S. nodiflora* (wp/CH₃OH) extracts against *T. castaneum* after 36h of exposure

Regression equation: $Y = 1.025 + 7.712X$ Chi-squared is 3.203 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.515 mg/cm² LD_{50} is 3.277mg/cm² 95% confidence limits are 2.995 to 3.585 mg/cm²

Appendix Table DCCXXIII: Dose mortality effect of *S. nodiflora* (wp/CH₃OH) extracts against *T. castaneum* after 48h of exposure

Regression equation: $Y = 2.819 + 4.911X$ Chi-squared is 1.631 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.444 mg/cm² LD_{50} is 2.780mg/cm² 95% confidence limits are 2.499 to 3.094 mg/cm²

Appendix Table DCCXXIV: Dose mortality effect of *Z. zerumbet* (ap/PetE) extracts against *T. castaneum* after 30min of exposure

Regression equation: $Y = 1.219 + 9.948X$ Chi-squared is 0.053 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.380 mg/cm² LD_{50} is 2.399mg/cm² 95% confidence limits are 2.209 to $2.605mg/cm²$

Regression equation: $Y = 2.369 + 7.692X$ Chi-squared is 6.058 with 2 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 0.342 mg/cm² LD_{50} is 2.198 mg/cm² 95% confidence limits are 1.883 to 2.567mg/cm²

Appendix Table DCCXXVI: Dose mortality effect of *Z. zerumbet* (ap/PetE) extracts against *T. castaneum* after 24h of exposure

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Regression equation: $Y = 2.567 + 8.632X$ Chi-squared is 6.957 with 2 degrees of freedom Variance has been adjusted for heterogeneity Log LD_{50} is 0.282 mg/cm² LD_{50} is 1.914 mg/cm² 95% confidence limits are 1.662 to 2.204 mg/cm²

Appendix Table DCCXXVII: Dose mortality effect of *Z. zerumbet* (ap/PetE) extracts against *T. castaneum* after 36h of exposure

────────────────────────────────────────────────────────────────────── Regression equation: $Y = 3.468 + 5.854X$ Chi-squared is 0.0002 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.262 mg/cm² LD_{50} is 1.827mg/cm² 95% confidence limits are 1.603 to 2.081mg/cm²

Appendix Table DCCXXVIII: Dose mortality effect of *Z. zerumbet* (ap/PetE) extracts against *T. castaneum* after 48h of exposure

Regression equation: $Y = -0.581 + 4.622X$ Chi-squared is 1.963 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.207mg/cm² LD_{50} is 1.612mq/cm² 95% confidence limits are 1.400 to $1.857mg/cm²$

Regression equation: $Y = 0.229 + 8.473X$

Chi-squared is 0.142 with 1 degrees of freedom No significant heterogeneity

Log LD_{50} is 0.563 mg/cm²

 LD_{50} is 3.657mg/cm²

95% confidence limits are 3.229 to 4.143 mg/cm²

Appendix Table DCCXXX: Dose mortality effect of *Z. zerumbet* (ap/CHCl3) extracts against *T. castaneum* after 24h of exposure

Regression equation: $Y = 1.959 + 5.784X$

Chi-squared is 1.540 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.526 mg/cm² LD_{50} is 3.355mg/cm²

95% confidence limits are 2.958 to 3.805mg/cm²

Appendix Table DCCXXXI: Dose mortality effect of *Z. zerumbet* (ap/CHCl₃) extracts against *T. castaneum* after 36h of exposure

Regression equation: $Y = 1.618 + 7.893X$ Chi-squared is 0.696 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.428 mg/cm² LD_{50} is 2.682 mg/cm² 95% confidence limits are 2.501 to 2.876 mg/cm²

Appendix Table DCCXXXII: Dose mortality effect of *Z. zerumbet* (ap/CHCl3) extracts against *T. castaneum* after 48h of exposure

Regression equation: $Y = 2.669 + 5.791X$ Chi-squared is 0.923 with 2 degrees of freedom No significant heterogeneity LOG LD_{50} IS 0.403mg/cm² LD_{50} IS 2.527 mg/cm² 95% confidence limits are 2.280 to 2.800mg/cm²

Regression equation: $Y = 3.553 + 2.806X$ Chi-squared is 0.091 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.516 mg/cm² LD_{50} is 3.280 mg/cm² 95% confidence limits are 1.836 to 5.859mg/cm²

Appendix Table DCCXXXIV: Dose mortality effect of *Z. zerumbet* (ap/CH₃OH) extracts against *T. castaneum* after 12h of exposure

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Regression equation: $Y = 3.293 + 4.981X$ Chi-squared is 0.158 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.343 mg/cm² LD_{50} is 2.201 mg/cm² 95% confidence limits are 1.925 to 2.517mq/cm²

Appendix Table DCCXXXV: Dose mortality effect of *Z. zerumbet* (ap/CH₃OH) extracts against *T. castaneum* after 24h of exposure

Regression equation: $Y = 3.610 + 5.055X$ Chi-squared is 1.535 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.275 mg/cm² LD_{50} is 1.883mg/cm² 95% confidence limits are 1.679 to $2.112mg/cm²$

Appendix Table DCCXXXVI: Dose mortality effect of *Z. zerumbet* (ap/CH₃OH) extracts against *T. castaneum* after 36h of exposure

Regression equation: $Y = 4.102 + 5.104X$ Chi-squared is 4.718 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.176 mg/cm² LD_{50} is 1.499mg/cm² 95% confidence limits are 1.332 to 1.687mg/cm² **Appendix Table DCCXXXVII:** Dose mortality effect of *Z. zerumbet* (ap/CH₃OH) extracts against *T. castaneum* after 48h of exposure

Regression equation: $Y = 0.268 + 4.227X$ Chi-squared is 0.416 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.119 mg/cm² LD_{50} is 1.316mg/cm² 95% confidence limits are 1.140 to 1.520 mq/cm²

Appendix Table DCCXXXIX: Dose mortality effect of *Z. zerumbet* (rh/PetE) extracts against *T. castaneum* after 12h of exposure

Regression equation: $Y = 1.364 + 2.820X$ Chi-squared is 1.929 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.289mg/cm² LD_{50} is 1.947mg/cm² 95% confidence limits are 1.479 to 2.564 mg/cm²

Regression equation: $Y = 0.956 + 2.809X$ Chi-squared is 0.690 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.440 mg/cm² LD_{50} is 2.753 mg/cm² 95% confidence limits are 1.745 to 4.344 mg/cm²

Appendix Table DCCXL: Dose mortality effect of *Z. zerumbet* (rh/PetE) extracts against *T. castaneum* after 24h of exposure

Regression equation: $Y = 2.154 + 2.484X$ Chi-squared is3.072 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.145 mg/cm² LD_{50} is 1.398mg/cm² 95% confidence limits are 1.110 to 1.761mg/cm²

α **Appendices** \Box **IES, RU**

Regression equation: $Y = 2.211 + 2.699X$ Chi-squared is6.276 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 1.033 mg/cm² LD_{50} is 1.079mg/cm² 95% confidence limits are 0.886 to 1.315mg/cm²

Appendix Table DCCXLIII: Dose mortality effect of *Z. zerumbet* (rh/CHCl₃) extracts against *T. castaneum* after 12h of exposure

────────────────────────────────────────────────────────────────────── Regression equation: $Y = 1.034 + 6.543X$ Chi-squared is 1.135 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.606mg/cm² LD_{50} is 4.038mg/cm²

95% confidence limits are 3.198 to 5.099mg/cm²

Appendix Table DCCXLII: Dose mortality effect of *Z. zerumbet* (rh/PetE) extracts against *T. castaneum* after 48h of exposure

Regression equation: $Y = 2.822 + 2.121X$ Chi-squared is1.726 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.0271 mg/cm² LD_{50} is 1.064mg/cm² 95% confidence limits are 0.789 to 1.436mg/ $cm²$

Appendix Table DCCXLIV: Dose mortality effect of *Z. zerumbet* (rh/CHCl3) extracts against *T. castaneum* after 24h of exposure

Regression equation: $Y = 0.613 + 8.444X$ Chi-squared is 0.371 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.520 mg/cm² LD_{50} is 3.308mg/cm² 95% confidence limits are 3.041 to 3.598mg/cm²

Regression equation: $Y = 1.894 + 6.436X$ Chi-squared is 0.124 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.483 mg/cm² LD_{50} is 3.038mg/cm² 95% confidence limits are 2.772 to 3.330mg/cm²

Appendix Table DCCXLVII: Dose mortality effect of *Z. zerumbet* (rh/CH₃OH) extracts against *T. castaneum* after 30min of exposure

────────────────────────────────────────────────────────────────────── Regression equation: $Y = 3.985 + 4.255X$ Chi-squared is 0.026 with 1 degrees of freedom No significant heterogeneity Log LD_{50} is 0.239 mg/cm² LD_{50} is 1.732mq/cm² 95% confidence limits are 1.469 to 2.043mg/cm²

Appendix Table DCCXLVI: Dose mortality effect of *Z. zerumbet* (rh/CHCl3) extracts against *T. castaneum* after 48h of exposure

Regression equation: $Y = 2.028 + 7.247X$ Chi-squared is 1.140 with 3 degrees of freedom No significant heterogeneity Log LD_{50} is 0.410 mg/cm² LD_{50} is 2.571 mg/cm² 95% confidence limits are 2.389 to 2.767mg/cm²

Appendix Table DCCXLVIII: Dose mortality effect of *Z. zerumbet* (rh/CH₃OH) extracts against *T. castaneum* after 12h of exposure

Regression equation: $Y = 0.585 + 3.887X$ Chi-squared is 1.096 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.136 mg/cm² LD_{50} is 1.368mg/cm² 95% confidence limits are 1.176 to 1.591mg/cm²

Regression equation: $Y = 0.404 + 4.437X$ Chi-squared is 3.198 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.036mg/cm² LD_{50} is 1.086mg/cm² 95% confidence limits are 0.941 to 1.253mg/cm²

Appendix Table DCCL: Dose mortality effect of *Z. zerumbet* (rh/CH₃OH) extracts against *T. castaneum* after 36h of exposure

Regression equation: $Y = 1.932 + 3.046X$ Chi-squared is 0.234 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 1.007 mg/cm² LD_{50} is 1.017 mg/cm² 95% confidence limits are 0.825 to 1.253mg/cm²

Appendix Table DCCLI: Dose mortality effect of *Z. zerumbet* (rh/CH3OH) extracts against *T. castaneum* after 48h of exposure

Regression equation: $Y = 2.333 + 2.965X$ Chi-squared is 0.218 with 2 degrees of freedom No significant heterogeneity Log LD_{50} is 0.899mg/cm² LD_{50} is 0.793mg/cm² 95% confidence limits are 0.650 to 0.969mg/cm²

Appendix Table DCCLXVII: Repellency of PetE extracts of *P. hysterophorus* (wp) against *A. gossypii*

Appendix Table DCCLXIX: Repellency of CH3OH extracts of *P. hysterophorus* (wp) against *A. gossypii*

Appendix Table DCCXC: Repellency of CH3OH extracts of *E. nummularius* (wp) against *T. castaneum* adults

Appendix Table DCCXCII: Repellency of CHCl₃ extracts of *L. camara* (ap) against *T. castaneum* adults

Appendix Table DCCXCVI: Repellency of CH3OH extracts of *L. camara* (r) against *T. castaneum* adults

Appendices \Box **IES, RU IES, RU**

Appendix Table DCCXCIX: Repellency of CH₃OH extracts of *M. piperita* (wp) against *T. castaneum* adults

Appendix Table DCCCI: Repellency of CHCI₃ extracts of *Mi. pudica* (wp) against *T. castaneum* adults

Appendix Table DCCCIII: Repellency of PetE extracts of *P. hysterophorus* (wp) against *T. castaneum* adults

Appendix Table DCCCIV: Repellency of CHCl3 extracts of *P. hysterophorus* (wp) against *T. castaneum* adults

Appendix Table DCCCV: Repellency of CH3OH extracts of *P. hysterophorus* (wp) against *T. castaneum* adults

Appendix Table DCCCVIII: Repellency of CH3OH extracts of *Ph. niruri* (wp) against *T. castaneum* adults

Appendix Table DCCCIX: Repellency of PetE extracts of *Po. hydropiper* (wp) against *T. castaneum* adults

Appendices \Box **IES, RU IES, RU**

Appendix Table DCCCXI: Repellency of CH3OH extracts of *Po. hydropiper* (wp) against *T. castaneum* adults

Appendix Table DCCCXIII: Repellency of CHCl³ extracts of *Pz. zeylanica* (wp) against *T. castaneum* adults

Appendix Table DCCCXIV: Repellency of CH3OH extracts of *Pz. zeylanica* (wp) against *T. castaneum* adults

Appendix Table DCCCXV: Repellency of PetE extracts of *S. nodiflora* (wp) against *T. castaneum* adults

Appendix Table DCCCXVII: Repellency of CH3OH extracts of *S. nodiflora* (wp) against *T. castaneum* adults

Appendix Table DCCCXVIII: Repellency of PetE extracts of *Z. zerumbet* (ap) against *T. castaneum* adults

Appendix Table DCCCXIX: Repellency of CHCI₃ extracts of *Z. zerumbet* (ap) against *T. castaneum* adults

Appendix Table DCCCXX: Repellency of CH3OH extracts of *Z. zerumbet* (ap) against *T. castaneum* adults

Appendix Table DCCCXXI: Repellency of PetE extracts of *Z. zerumbet* (rh**)** against *T. castaneum* adults

Appendix Table DCCCXXII: Repellency of CHCl3 extracts of *Z. zerumbet* (rh) against *T. castaneum* adults

Appendix Table DCCCXXIII: Repellency of CH3OH extracts of *Z. zerumbet* (rh) against *T. castaneum* adults

Appendix Table DCCCXXIV: Supporting spectra for the compound 1 (ENP) [Page 1]

2014/6/12 13:49:47 1 / 4

==== Shimadzu LabSolutions Analysis Report ====

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2014/6/12 13:49:47 2 / 4

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2014/6/12 13:49:47 3 / 4

2014/6/12 13:49:47 4 / 4

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Appendix Table DCCCXXV: Supporting spectra for the compound 2 (POM) [Page 1]

2014/6/12 13:59:21 1 / 8

==== Shimadzu LabSolutions Analysis Report ====

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Appendix Table DCCCXXV: Supporting spectra for the compound 2 (POM) [Page 2]

2014/6/12 13:59:21 2 / 8

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2014/6/12 13:59:21 3 / 8

Appendix Table DCCCXXV: Supporting spectra for the compound 2 (POM) [Page 4]

2014/6/12 13:59:21 4 / 8

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2014/6/12 13:59:21 5 / 8

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Appendix Table DCCCXXV: Supporting spectra for the compound 2 (POM) [Page 6]

2014/6/12 13:59:21 6 / 8

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2014/6/12 13:59:21 7 / 8

Event#: 1 Scan(E+) Ret. Time : [6.467->6.533]-[6.133<->6.767] Scan# : [389->

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2014/6/12 13:59:21 8 / 8

Event#: 2 Scan(E-) Ret. Time : [12.050->12.116]-[11.916<->12.216] Sca

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